Repurposing of thalidomide and its derivatives for the treatment of SARS-coV-2 infections: Hints on molecular action

Lakshmikirupa Sundaresan¹, Suvendu Giri², Himanshi Singh², and Suvro Chatterjee²

June 16, 2020

Abstract

Background and Purpose: SARS-coV-2 pandemic continues to cause an unprecedented global destabilization. There is an urgent need to develop vaccines or identify molecules to treat severe cases and repurposing of drugs is the best approach at this hour. Thalidomide, despite having an infamous history has been successfully repurposed and tested for various disease conditions including inflammatory diseases and tumor. Few reports emphasize the use of thalidomide with a SARS-coV-2 pneumonia patient being successfully treated with thalidomide. Experimental Approach: A meta-analysis comparing the transcriptomes of SARS-coV-2 infected tissues with thalidomide and lenalidomide-induced transcriptomic changes in transformed lung, endothelial and hematopoietic models was performed. Key Results: Thalidomide and lenalidomide exhibited pleiotropic effects affecting a range of biological processes including inflammation, immune response, angiogenesis, MAPK signaling, NOD-like receptor signaling, TLR signaling, leukocyte differentiation and innate immunity, the processes which are aberrantly regulated in severe COVID-19 patients. In addition, we show the similarities between the expression profiles of SARS-coV-2 infected lung and systemic lupus erythematous. Conclusion and Implications: The present study recommends thalidomide analogs as a "better fit" to treat severe cases of novel viral infections, healing the damaged network by compensating the impairment caused by the Coronavirus disease-2019 (COVID-19).

Repurposing of thalidomide and its derivatives for the treatment of SARS-coV-2 infections: Hints on molecular action

Lakshmikirupa Sunderesan[¥], Suvendu Giri^{#,*}, Himanshi Singh^{#,*}, and Suvro Chatteriee^{#,*}

Correspondence:

Suvro Chatterjee Ph.D,

AU-KBC Research Centre,

MIT Campus of Anna University,

Chromepet, Chennai-600044, India.

Tel.: $+91\ 44\ 2223\ 4885\times48/+91\ 44\ 2223\ 2711\times48;$

Fax: $+91 \ 44 \ 2223 \ 1034$

¹Hospital for Sick Children ²AU-KBC Research Centre

^{*}Vascular Biology Laboratory, AU-KBC Research Centre, Chennai, India.

[#]Department of Biotechnology, Anna University, Chennai, India.

^{*}Developmental & Stem Cell Biology Program, The Hospital for Sick Children, Toronto, ON M5G 1X8, Canada

E-mail address: soovro@yahoo.ca

Running title: Mechanisms of thalidomide in treating COVID-19

Word count: 2,733 Figure Count: 6 Table Count: 0

Keywords: SARS-coV-2, COVID-19, Thalidomide, Lenalidomide, Inflammation, Immune Response, Angiogenesis, Endothelium

What is already known about this subject

- Severe cases of COVID-19 infections show an aberrant surge in the host immune response and cytokine storm in lungs.
- There is an increased amount of angiogenesis observed in lungs of COVID-19 patients
- A patient with severe pneumonia-associated with COVID-19 has been successfully treated with thalidomide

What this study adds

- This study presents insights into the possible mechanisms by which thalidomide and lenalidomide would suppress the cytokine storm and immune response
- Thalidomide and its derivative lenalidomide modulate expression of several genes and key pathways aberrantly regulated in SARS-coV-2 infected tissues.
- There is a similarity in gene expression profiles of SARS-coV2 infected tissues with systemic lupus erythematosus

Abstract

Background and Purpose: SARS-coV-2 pandemic continues to cause an unprecedented global destabilization. There is an urgent need to develop vaccines or identify molecules to treat severe cases and repurposing of drugs is the best approach at this hour. Thalidomide, despite having an infamous history has been successfully repurposed and tested for various disease conditions including inflammatory diseases and tumor. Few reports emphasize the use of thalidomide with a SARS-coV-2 pneumonia patient being successfully treated with thalidomide.

Experimental Approach: A meta-analysis comparing the transcriptomes of SARS-coV-2 infected tissues with thalidomide and lenalidomide-induced transcriptomic changes in transformed lung, endothelial and hematopoietic models was performed.

Key Results: Thalidomide and lenalidomide exhibited pleiotropic effects affecting a range of biological processes including inflammation, immune response, angiogenesis, MAPK signaling, NOD-like receptor signaling, TLR signaling, leukocyte differentiation and innate immunity, the processes which are aberrantly regulated in severe COVID-19 patients. In addition, we show the similarities between the expression profiles of SARS-coV-2 infected lung and systemic lupus erythematous.

Conclusion and Implications: The present study recommends thalidomide analogs as a "better fit" to treat severe cases of novel viral infections, healing the damaged network by compensating the impairment caused by the Coronavirus disease-2019 (COVID-19).

Abbreviations: COVID-19 - Coronavirus Disease -19; BALF - Broncho-alveolar lavage fluid; PBMC - Peripheral Blood Mononuclear Cell; DEG- Differentially Expressed Genes; GSEA - Gene Set Enrichment Analysis; SLE - Systemic Lupus Erythematosus; TLR - Toll-like Receptor; HUVEC - Human Umbilical Vein Endothelial Cell

Introduction

Novel coronavirus, SARS-coV-2 has been posing devastating effects on a global scale with a soaring number of infections and an alarming rate of mortality. Despite tremendous efforts, development of effective vaccines or anti-viral medications is months away leaving repurposing of drugs with known safety and efficacy profiles as the only viable choice. COVID-19 is clinically very challenging since the novel coronavirus triggers multiorgan turbulence devastating the homeostasis of the human system. Once the SARS-CoV-2 virus enters the respiratory tract, there are four different stages of the infection from symptoms to multi-organ failures. Phase I starts with the ACE2 receptor-mediated viral entry through nasal and oral routes followed by host immune system alert simultaneously with active viral replication in the upper respiratory tract (Phase II). Next, Phase III coincides with minor cytokine storm in the alveoli releasing the inflammatory cytokines making the blood vessels leaky around the air sac, and ultimately the second cytokine storm arrives with uncontrolled inflammatory and life-threatening symptoms, acute respiratory distress syndrome (ARDS), seizure, severe hypoxia and severed organ toxicity (Phase IV)^{1,2}. Manifestation of the bi-phasic cytokine storm occurs through the activation of a series of cytokines including granulocyte-colony stimulating factor (G-CSF), interferon gamma-induced protein 10 (CXCL10), monocyte chemoattractant protein 1 (MCP1), macrophage inflammatory protein 1α (MIP- 1α), tumour necrosis factor α (TNF- α), interleukin (IL)-2R and IL-6 overwhelming the system leading to indiscriminate damages in multiple organs 3-5. There is an increased amount of blood vessel growth in the lungs of COVID-19 patients compared to severe influenza ⁶. Due to such multi-layers of problems associated with the COVID-19 infections, researchers around the world are desperately in search for a drug, which would able to tackle all or few of these COVID-19 hallmarks.

Thalidomide, a small molecule drug, was marketed with an intent to relieve morning sickness in pregnant women. However, this resulted in an unexpected misfortune that resulted in the birth of several children with defects due to in-utero exposure ⁷. Later, thalidomide became a game changer for its the multi-faced pharmacological effects such as immunomodulation, anti-inflammation, anti-angiogenesis, and anti-viral effects ⁸. At this point of time, the world needs a "smart" solution. Thalidomide increases the hope for treating COVID-19 patients ⁹. Chen et al reports successful treatment of SARS-coV-2 associated pneumonia with combinatory treatment of thalidomide and a low-dose glucocorticoid ¹⁰ Two clinical trials, NCT04273581 and NCT04273529 have been registered to check the efficacy of thalidomide in treating COVID-19 patients. The adverse effects of thalidomide and its analogs are well documented. Various genes aberrantly expressed in SARS-coV-2 affected lungs are known targets of thalidomide (Table S1). Extensive information available on thalidomide's mechanisms, their efficacy and safety in hemophagocytic syndrome-induced cytokine storm¹¹ and idiopathic pulmonary fibrosis¹², severe H1N1 and paraquat poisoning lung injury^{13,14} argue for the possible action of thalidomide on COVID-19 induced lung effects and cytokine storm.

Transcriptome-based approach to connect diseases with drug responses is a recognized strategy in drug repurposing ¹⁵. With the fast-growing literature on SARS-coV-2 infections, we performed meta-analysis of whole transcriptome signatures of lungs, PBMC, BALF from SARS-coV-2 affected patients and A549 cells and compared with the gene expression signatures of thalidomide or lenalidomide-treated A549 (transformed adenocarcinoma cells), hematopoietic and endothelial cells. We hereby provide possible mechanistic actions of thalidomide in treating the SARS-coV-2 pathology. In addition, we suggest that the derivatives of thalidomide, lenalidomide and CC-220 might also be effective in the treatment of SARS-coV-2.

Methods

Data Collection

A total of 16 gene expression datasets including 15 publicly available expression datasets were meta-analyzed in this study. Transcriptomes of SARS-coV-2 infected lung tissues matched with healthy control and SARS-coV-2 treated A549 cells were obtained from GEO (Accession ID: GSE147507) ¹⁶. Transcriptome data of bronchoalveolar lavage fluid (BALF) and peripheral blood mononuclear cells (PBMC) were obtained from BIG Data Center (https://bigd.big.ac.cn/) (Accession ID: CRA002390) ¹⁷. The expression profiles of systemic lupus erythematosus (Accession ID: GSE112087) ¹⁸, bone marrow cells treated with lenalidomide (Accession ID: GSE106748) ¹⁹, lymphoma cells treated with lenalidomide (Accession ID: GSE60618) ²⁰, CD34 positive cells treated with pomalidomide (Accession ID: GSE144052), MERS infected PBMC (Accession:

GSE1739) 21 , lenalidomide treated PBMC (Accession ID: GSE84251) and CC-122 treated lymphoma cells (Accession ID: GSE75420) 22 were procured from GEO and differentially expressed genes (DEG) were identified using limma 23 . Library of Integrated Network-Based Cellular Signatures (iLINCS) is a database which contains the gene expression signatures of more than 21,000 compounds (http://www.ilincs.org/ilincs/). We obtained the gene expression signatures for A549 cells treated with 10μ M thalidomide for 6h (LINCSCP_4683) and 24h (LINCSCP_4463), 100μ M lenalidomide for 6h (LINCSCP_4650) and 24h (LINCSCP_4427).

Transcriptome sequencing

SARS-coV-2 affects the cardiovascular system to a great extent 6 and to understand the effects of thalidomide and its derivatives, human umbilical vein endothelial cells (HUVEC) were subjected to $20\mu\mathrm{M}$ thalidomide or $20\mu\mathrm{M}$ lenalidomide or $20\mu\mathrm{M}$ pomalidomide or vehicle control treatment for 8h. RNA was isolated using TRIzol® method and whole transcriptome sequencing was performed using Illumina HiSeq 2500 platform. The data can be accessed at GEO with the Accession ID GSE118979. The sequence reads were aligned with reference genome of *Homo sapiens* using TopHat2 (v2.0.8) and the followed by transcript compilation and gene identification was done using Cufflinks (v2.2.0) 24 . The DEG were identified using Cuffdiff program (v2.2.0) 24 .

Differential Expression and Enrichment Analysis

The raw counts from the SARS-coV-2 transcriptomic profiles were subjected to differential expression analysis by DESEq2 v1.26.0²⁵. Subsequently the genes were pre-ranked using the p-values from DESeq2 analysis and subjected to pre-ranked gene set enrichment analysis (GSEA) ²⁶. Gene sets with FDR <0.05 were considered to be statistically significant and were visualized using EnrichmentMap plugin ²⁷ of Cytoscape²⁸. For drug signatures from iLINCS, differentially expressed genes with p<0.05 were considered to be statistically significant. Enrichment of kinase perturbation was carried out using Enrichr ²⁹.

KINOMEscan Kinase Screening

We analyzed our previously published KINOMEscan kinase screening dataset of thalidomide 30 in order to investigate how thalidomide affects the immune system. Kinases whose activities were reduced at least by 60% were considered for further enrichment analysis.

Comparative analysis using Toppcluster

The differentially expressed genes (Q<0.05 for transcriptome and p<0.05 for drug signatures) from all the gene expression profiles were compared for overlapping genes and over-represented pathways using Toppcluster and the networks were visualized using in Cytoscape 28 .

Identification of protein targets using PharmMapper

For identifying the protein targets of the thalidomide, we utilized the PharmMapper server (http://www.lilab-ecust.cn/pharmmapper/)³². The server identifies possible physiological protein targets of any drug molecule by using a pharmacophore-based mapping approach. The three-dimensional structure of thalidomide was obtained from PubChem and processed on the PharmMapper server choosing only human protein target sets. The top 100 target proteins were selected based on the ranking associated with a fit score (pKd value) for further enrichment analysis using Enrichr²⁹.

Results

Meta-analysis of SARS-coV-2 affected lung biopsies and PBMCs reveal enrichment of various pathways pertaining to immune response

SARS-coV-2 infection caused a massive surge in inflammatory response, cytokine production and cytokine-mediated signaling. There was a substantial up-regulation of immune response including the processes of hematopoietic development and lymphocyte activation (Figure 1A,3A,3B). Activation of viral life cycle and anti-viral interferon signaling was observed in infected lungs and A549 cells (Figure 1B, 5). Over-representation of pathways including NOD-like receptor signaling, MAPK cascade, Measles and Influenza-

A were seen in the infected lung as identified by gene set enrichment analysis (Figure 1A). Targets of transcription factors, ISRE and IRF were upregulated in SARS-coV-2 affected lungs similar to up-regulation observed in the PBMC of SLE patients (Figure 1B, 1C). Enrich analysis revealed that the genes up-regulated in SARS-coV-2 lung were down-regulated when SYK was knocked down or inhibited (Figure 1D). Expression to kinase (X2K) analysis showed the possible perturbation of various MAP kinases, ABL1 and JNK1 (Figure 1E,1F). Human phenotype enrichment analysis shows thrombocytopenia, poor wound healing, abnormality of lymphatic system, serositis and abnormal anticoagulant pathways in SARS-coV-2 infected lungs (Figure S1).

Effect of Thalidomide on Kinases Implicated in Immune Response and MAPK signaling

The kinase screening assay identified key kinases involved in the regulation of immune response are functionally modulated by thalidomide. The most kinase affected was LCK and SYK (Figure 2C), critical modulators of T cell receptor signaling. Many LCK substrates, SPI1, TBK1, FOXP3, EGR1, ESR1, IRF1, CBL and STAT1 as well as SYK phosphorylation targets such as OAS1 and MX1 were up-regulated in SARS-coV-2 lung. Various processes mediating immune response including JUN phosphorylation, IKappaB phosphorylation, JAK-STAT pathway, leukocyte mediated immunity, neutrophil degranulation and activation, B cell receptor signaling and MAPK cascade were found to be affected by thalidomide (Figure 2D). PharmMapper results showed strong affinity for LCK, HCK and SYK along with other proteins involved in innate and adaptive immune response (Table S2, S3, S4). We studied the effects of thalidomide and its derivatives on endothelium and identified the down-regulation of several angiogenic genes (Figure S5 and Table S1).

Up-regulated pathways and GO Biological Processes in SARS-coV-2 infection suppressed by thalidomide and lenalidomide

SARS-coV-2 infected lungs, PBMC and A549 cells showed significant upregulation of expression of genes involved in inflammation, cytokine signaling, MAPK signaling and activation of cells mediating the immune response whereas BALF exhibited a slightly different immune profile where were leukocyte and neutrophil activation was suppressed (Figure 3A). Comparison of differentially expressed genes of all the signatures yielded interesting results. Many of the processes up-regulated in SARS-coV-2 infected tissues were suppressed by thalidomide and lenalidomide in A549 cells and endothelial cells (Figure 2A,2B,3A). Thalidomide-treated A549 cells showed suppression of key genes including SYK, JUN, PIK3CA and HLA genes implicated in immune response (Figure 3B, 3C). Thalidomide and lenalidomide down-regulated various pro-inflammatory and angiogenic genes aberrantly expressed in SARS-cov-2 infected lungs including CCL2 and TSC22D3 which are NF-xB modulators in A549 cells (Figure S3, S4). Thalidomide and lenalidomide treatment resulted in significantly suppression of cytokine response, angiogenesis, inflammation, Fc Epsilon receptor signaling and MAPK cascade (Figure 2A). In addition, lenalidomide down-regulated STAT1 expression, leukocyte differentiation, TLR signaling as well as IRF activation (Figure 2B, 4B). Many genes implicated in NOD-like receptor signaling overexpressed in SARS-coV-2 were suppressed by lenalidomide in A549 and lymphoma cells (Figure 4C). B cell receptor signaling was activated in SARS-coV-2 affected PBMC whereas T cell activation was observed in SARS-coV-2 lungs. Translation of viral mRNA was exclusively observed in SARS-coV-2 infected BALF whereas genes implicated in viral entry and life cycle were up-regulated in SARS-coV-2 infected lungs, BALF and A549 cells. Genes involved in viral entry and type I interferon signaling were down-regulated in thalidomide-treated A549 cells and lenalidomide-treated lymphoma, A549 and HUVEC (Figure 5A, 5C).

Similarity of SARS-coV-2 expression profile with that of SLE, lymphoma and multiple myeloma

Comparative enrichment analysis of gene expression profiles for disease-specific phenotypes revealed the similarity of SARS-infected tissues with pneumonia, influenza, lymphoma, systemic lupus erythematosus (SLE), multiple myeloma, asthma, auto-immune diseases, asthma, pneumonia, and atherosclerosis while SARS-coV-2 PBMC showed exclusive overlap with lymphoma, multiple myeloma and chronic lymphocytic leukemia (CLL) (Figure 6A). The gene expression profile of SARS-coV-2 infected lungs was very similar to that of PBMC from SLE patients (Figure 6C). There was a significant overlap between SARS-coV-2

and SLE in up-regulated genes involved in immune regulation (Figure 3B), interferon signaling (Figure 5B) and disease-specific phenotypes (Figure 6B, 6D, 6E). Many genes deregulated in lymphoma and multiple myeloma were up-regulated in SARS-coV-2 infections as well (Figure 6B, 6E). Several aberrantly expressed genes involved in these disease phenotypes were down-regulated in thalidomide and lenalidomide treated A549 and endothelium (Figure 6).

Discussion

SARS-coV-2 infection causes surge in a number of pathways related to inflammation, cytokine signaling, leukocyte and lymphocyte activation, innate and adaptive immune response marking the phenomenon of "cytokine storm". As the whole immune system is affected during the SARS-coV-2 infection, immunomodulators would be highly beneficial in treating the symptoms. A COVID-19 patient with pneumonia was treated successfully with thalidomide and low dose glucocorticoid. There was a significant decrease in the inflammatory cytokines including IL1-, IL-6 and IFN- and increase in the CD4+ and CD8+ T cells and NK cells. Thalidomide reduced the severity of many COVID-19 symptoms such as lung lesions, exudation due to its pleiotropic effects on the human system¹⁰. Hemophagocytic syndrome, a hyperinflammatory disorder is also another condition in which cytokine storm occurs. It is frequently present with extranodal natural killer/T cell lymphoma (ENKTL). Thalidomide was effective in suppressing the cytokine storm through inhibition of NF-xB based transcription of IFN- and TNF genes 11 and thalidomide along with P-Gemox was highly effective in treating ENKTL patients in a Phase II clinical trial ³³. Comparison of SARS-coV-2 expression profiles with drug signatures through enrichment analysis revealed striking actions of thalidomide and lenalidomide in A549 and endothelial cells. The results suggest that thalidomide and lenalidomide could reverse the devastating effects of SARS-coV-2 infections on immune system. We selected A549, an adenocarcinomic human alveolar basal epithelial cell line to test our hypothesis that thalidomide would be effective against the cytokine storms. The A549 cell line is an appropriate model for testing cytokine storm targeting drugs since a previous study established this model by infecting the cells with influenza A/H1N1 virus (PR-8) or nonstructural protein 1 (NS1) plasmid to test the mechanisms behind inflammatory cytokines/chemokines mediated "cytokine storm" 34 Studies have utilized A549 cells to show the effects of thalidomide on lung fibrosis ^{35–37}. A limitation of this study is that only 978 genes called "landmark genes" are profiled in the iLINCS drug signatures. However, the profiles are highly reproducible and represent the whole transcriptome^{15,38}. Our models of A549 and HUVEC effectively capture the effects of thalidomide in lungs as well as endothelium.

It is also emerging that SARS-coV-2 infections perturb vascular plexus significantly and there is a substantial increase in the growth of new blood vessels and evidence of intussusceptive angiogenesis with overexpression of angiogenesis and hypoxia genes in the lungs of COVID-19 patients ⁶. Cytokine storm and atherosclerosis are tightly connected in SARS-coV-2 ³⁹which is consistent with our analysis revealing the enrichment of atherosclerosis in the SARS-coV-2 signatures (Figure 6). Thalidomide is a renowned modulator of vascular system, and it is known to transcriptionally or functionally target various genes (Table S1) up-regulated genes in the lungs of COVID-19 patients^{6,16}. As SARS-coV-2 infection has a huge impact on the hematopoietic system ⁴⁰ affecting the myeloid cell maturation, we meta-analyzed the effects of thalidomide and its derivatives on PBMC, bone marrow cells as well as lymphoma cells. Thalidomide and lenalidomide exhibited attenuation of cytokine signaling and inflammation in addition to its anti-angiogenic action (Figure 3A). The drugs affected most of the pathways up-regulated in SARS-coV-2 affected lungs and PBMC (Figure 1A, 3A) in A549 cells, mandating direct investigations in SARS-coV-2 infected models.

COVID-19 coincides with a strong neuro-endocrine modulation because the disease devastates functions of the organs, and naturally the reciprocal communication between the organs of the endocrine stress system gets a set-back ⁴¹. ACE2 is expressed along the hypothalamus, pituitary and adrenal (HPA) axis which is implicated in the stress response and adrenal glands has the highest concentration of virus particles next to lung ⁴². A high expression of ACE2 in brain is believed to be the reason for the possible infection of the central nervous system in SARS patients ⁴³. Chronic elevated stress levels have been reported in SARS and SARS-coV-2 patients even long after the outbreak ¹⁰. Notably, thalidomide is also known for

its neuro-endocrine modulation properties. Thalidomide modulates CNS by reducing the generation of pro-inflammatory cytokines such as IL-1, IL-6, IL-8 and TNF- α through NF- α B inhibition⁴⁴. There was a down-regulation of genes involved in circadian wake cycle (Figure 1B, S2) including PER3 in the PBMC of COVID-19 patients hinting on the possible sleep disturbances in SARS-coV-2 patients. Thalidomide being a well-known antiemetic and sedative action on the neuroendocrine axis would relax the patients which is supported by the report that thalidomide was effective in treating the anxiety and digestive symptoms in the COVID-19 patient¹⁰.

The anti-inflammatory properties of thalidomide and its analogs through reduction of IL-1β, TNF-α expression and NF-xB inhibition are well established⁴⁵. SARS-coV-2 infections showing elevated NF-xB signaling and rampage activation of immune response. Unlike other RNA viruses, SARS-coV-2 suppresses TNF receptor-associated factors 3 (TRAF3) activation, inhibiting NF-xB and IRFs, leading to suppression of early pro-inflammatory and antiviral responses. Whereas later stages of the infection show an enhanced expression of IRF targets in the lungs with an activation of IL-1, IL-6 and TNF-α expression and inhibition of type I interferon signaling ⁴⁶. Activation of IRF and ISRE transcriptional targets in SARS-coV-2 affected lungs is in agreement with previous studies reporting the SARS biology ⁴⁷. Thalidomide inhibited LCK activity affecting STAT1 phosphorylation, cytokine mediated signaling, NF- xB signaling, osteoclast differentiation and MAPK signaling through modulation of various upstream activators and downstream effectors. Lenalidomide, in addition, suppressed leukocyte differentiation, TLR signaling along with IRF activation in A549 and lymphoma cells. The effects of thalidomide and lenalidomide observed in our study are consistent with the previous studies where thalidomide and lenalidomide has been shown to inhibit IRF and STAT1 phosphorylation resulting in the downregulation of interferon expression and TLR signaling ^{48,49}.

The expression profile of SARS-coV-2 infected lungs, PBMC as well as A549 cells show resemblance with profiles of lymphoma, multiple myeloma and SLE. Therefore, drugs which are effective in treating SLE, lymphoma and multiple myeloma might be effective against SARS-coV-2 infection. Thalidomide and its derivatives show impressive efficacy in treating multiple myeloma and certain forms of lymphoma ⁴⁵. Remarkably, hydroxychloroquine, an FDA approved SLE drug is currently being used in the management of critically ill SARS-coV-2 patients⁵⁰. CC-220, another thalidomide analog shows very promising results in phase I/II clinical trials against SLE^{51,52}. CC-220 through suppression of Ikaros and Aiolos expression ⁵³, transcription factors which are essential for differentiation of leukocyte and NK cells thus modulating the innate immune system. As innate immune system pathways are deregulated in SARS-coV-2 infected lung and PBMC, further studies are warranted to investigate the efficacy and safety of CC-220 in treating COVID-19

Any treatment strategy with thalidomide and its analogs including repurposing thalidomide for COVID-19, should consider thalidomide-induced adverse effects including neuropathy and venous thromboembolism (VTE) ⁵⁴. There have been many reports on COVID-19 patients develop blood clots ³⁹, a dangerous issue which might be aggravated with the use of thalidomide and lenalidomide. In addition, lenalidomide might cause cytokine release syndrome in chronic lymphocytic leukemia patient ⁵⁵. Therefore, a very careful dosage regimen has to be followed with all these drugs as serious adverse effects have been observed during dose escalation earlier.

Conclusion

Our study sheds light on the possible mechanisms through which thalidomide and lenalidomide might be effective in the management of SARS-coV-2 pathology. Thalidomide and derivatives effectively modulating various aberrantly regulated pathways infections with abundant pharmacological information available make them promising candidates for the treatment of novel coronavirus infections.

Acknowledgement

This project was partially supported by a grant from University Grant Commission-Faculty Recharge Programme (UGC-FRP), Government of India to SC.

Contributors

LS, SG, HS and SC contributed to study design, experiments, and data collection while LS, SG, and SC helped in manuscript preparation.

Conflict of Interest

The authors have none to declare.

Data Availability Statement

The data generated in this study have been deposited to Gene Expression Omnibus. The datasets that support this study are available in GEO at https://www.ncbi.nlm.nih.gov/geo/, BIG Data Center at https://bigd.big.ac.cn/ and iLINCS at http://www.ilincs.org/ilincs/. The appropriate references have been mentioned in the Methods section.

References

- 1. Sungnak W, Huang N, Bécavin C, et al. SARS-CoV-2 entry factors are highly expressed in nasal epithelial cells together with innate immune genes. $Nat\ Med$. 2020;26(5):681-687. doi:10.1038/s41591-020-0868-6
- 2. Xu Z, Shi L, Wang Y, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. Lancet Respir Med . 2020;8(4):420-422. doi:10.1016/S2213-2600(20)30076-X
- 3. Chen L, Liu H, Liu W, et al. Analysis of clinical features of 29 patients with 2019 novel coronavirus pneumonia. Zhonghua Jie He Hu Xi Za Zhi . 2020;43(0):E005.
- 4. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet. 2020;395(10223):497-506. doi:10.1016/S0140-6736(20)30183-5
- 5. Tay MZ, Poh CM, Rénia L, MacAry PA, Ng LFP. The trinity of COVID-19: immunity, inflammation and intervention. *Nat Rev Immunol*. Published online April 28, 2020:1-12. doi:10.1038/s41577-020-0311-8
- 6. Ackermann M, Verleden SE, Kuehnel M, et al. Pulmonary Vascular Endothelialitis, Thrombosis, and Angiogenesis in Covid-19. $N\ Engl\ J\ Med$. Published online May 21, 2020:NEJMoa2015432. doi:10.1056/NEJMoa2015432
- 7. Stephens TD, Bunde CJW, Fillmore BJ. Mechanism of action in thalidomide teratogenesis. *Biochem Pharmacol* . 2000;59(12):1489-1499. doi:10.1016/S0006-2952(99)00388-3
- 8. Rehman W, Arfons LM, Lazarus HM. The rise, fall and subsequent triumph of thalidomide: lessons learned in drug development. Ther $Adv\ Hematol\ .\ 2011;2(5):291-308.\ doi:10.1177/2040620711413165$
- 9. Rismanbaf A. Potential Treatments for COVID-19; a Narrative Literature Review. *Arch Acad Emerg Med* . 2020;8(1):e29. Accessed May 21, 2020. http://www.ncbi.nlm.nih.gov/pubmed/32232214
- 10. Chen C, Qi F, Shi K, et al. *Thalidomide Combined with Low-Dose Glucocorticoid in the Treatment of COVID-19 Pneumonia*. Preprints; 2020. Accessed May 20, 2020. www.preprints.org
- 11. Wen H, Ma H, Cai Q, et al. Recurrent ECSIT mutation encoding V140A triggers hyperinflammation and promotes hemophagocytic syndrome in extranodal NK/T cell lymphoma. $Nat\ Med$. 2018;24(2):154-164. doi:10.1038/nm.4456
- 12. Somogyi V, Chaudhuri N, Torrisi SE, Kahn N, Müller V, Kreuter M. The therapy of idiopathic pulmonary fibrosis: what is next? $Eur\ Respir\ Rev$. 2019;28(153):190021. doi:10.1183/16000617.0021-2019
- 13. Zhu H, Shi X, Ju D, Huang H, Wei W, Dong X. Anti-Inflammatory Effect of Thalidomide on H1N1 Influenza Virus-Induced Pulmonary Injury in Mice. Inflammation . 2014;37(6):2091-2098. doi:10.1007/s10753-014-9943-9

- 14. Li D, Zhang X-W, Jiang X-Q, et al. Protective effects of thalidomide on pulmonary injuries in a rat model of paraquat intoxication. J Inflamm . 2015;12(1):46. doi:10.1186/s12950-015-0093-0
- 15. Subramanian A, Narayan R, Corsello SM, et al. A Next Generation Connectivity Map: L1000 Platform and the First 1,000,000 Profiles. *Cell*. Published online 2017. doi:10.1016/j.cell.2017.10.049
- 16. Blanco-Melo D, Nilsson-Payant BE, Liu WC, et al. Imbalanced Host Response to SARS-CoV-2 Drives Development of COVID-19. *Cell* . Published online 2020. doi:10.1016/j.cell.2020.04.026
- 17. Xiong Y, Liu Y, Cao L, et al. Transcriptomic characteristics of bronchoalveolar lavage fluid and peripheral blood mononuclear cells in COVID-19 patients. $Emerg\ Microbes\ Infect\ .\ 2020;9(1):761-770.\ doi:10.1080/22221751.2020.1747363$
- 18. Oon S, Monaghan K, Ng M, et al. A potential association between IL-3 and type I and III interferons in systemic lupus erythematosus. *Clin Transl Immunol* . 2019;8(12). doi:10.1002/cti2.1097
- 19. Lopez-Millan B, Diaz de la Guardia R, Roca-Ho H, et al. IMiDs mobilize acute myeloid leukemia blasts to peripheral blood through downregulation of CXCR4 but fail to potentiate AraC/Idarubicin activity in preclinical models of non del5q/5q- AML. Oncoimmunology . 2018;7(9). doi:10.1080/2162402X.2018.1477460
- 20. Gopalakrishnan R, Matta H, Tolani B, Triche T, Chaudhary PM. Immunomodulatory drugs target IKZF1-IRF4-MYC axis in primary effusion lymphoma in a cereblon-dependent manner and display synergistic cytotoxicity with BRD4 inhibitors. *Oncogene* . 2016;35(14):1797-1810. doi:10.1038/onc.2015.245
- 21. Reghunathan R, Jayapal M, Hsu LY, et al. Expression profile of immune response genes in patients with severe acute respiratory syndrome. *BMC Immunol* . 2005;6. doi:10.1186/1471-2172-6-2
- 22. Hagner PR, Man HW, Fontanillo C, et al. CC-122, a pleiotropic pathway modifier, mimics an interferon response and has antitumor activity in DLBCL. *Blood* . 2015;126(6):779-789. doi:10.1182/blood-2015-02-628669
- 23. Smyth GK. Linear Models and Empirical Bayes Methods for Assessing Differential Expression in Microarray Experiments. $Stat\ Appl\ Genet\ Mol\ Biol\ .\ 2004; 3(1):1-25.\ doi:10.2202/1544-6115.1027$
- 24. Trapnell C, Roberts A, Goff L, et al. Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks. $Nat\ Protoc$. 2012;7(3):562-578. doi:10.1038/nprot.2012.016
- 25. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol* . 2014;15(12):550. doi:10.1186/s13059-014-0550-8
- 26. Subramanian A, Tamayo P, Mootha VK, et al. Gene set enrichment analysis: A knowledge-based approach for interpreting genome-wide expression profiles. $Proc\ Natl\ Acad\ Sci\ U\ S\ A$. 2005;102(43):15545-15550. doi:10.1073/pnas.0506580102
- 27. Merico D, Isserlin R, Stueker O, Emili A, Bader GD. Enrichment Map: A Network-Based Method for Gene-Set Enrichment Visualization and Interpretation. Ravasi T, ed. *PLoS One* . 2010;5(11):e13984. doi:10.1371/journal.pone.0013984
- 28. Shannon P, Markiel A, Ozier O, et al. Cytoscape: A software Environment for integrated models of biomolecular interaction networks. *Genome Res.* 2003;13(11):2498-2504. doi:10.1101/gr.1239303
- 29. Kuleshov M V., Jones MR, Rouillard AD, et al. Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. *Nucleic Acids Res.* 2016;44(W1):W90-W97. doi:10.1093/nar/gkw377
- 30. Sundaresan L, Kumar P, Manivannan J, et al. Thalidomide and Its Analogs Differentially Target Fibroblast Growth Factor Receptors: Thalidomide Suppresses FGFR Gene Expression while Pomalidomide Dampens FGFR2 Activity. *Chem Res Toxicol* . 2019;32(4):589-602. doi:10.1021/acs.chemrestox.8b00286

- 31. Kaimal V, Bardes EE, Tabar SC, Jegga AG, Aronow BJ. ToppCluster: a multiple gene list feature analyzer for comparative enrichment clustering and network-based dissection of biological systems. *Nucleic Acids Res*. 2010;38(Web Server):W96-W102. doi:10.1093/nar/gkq418
- 32. Liu X, Ouyang S, Yu B, et al. Pharm Mapper server: a web server for potential drug target identification using pharmacophore mapping approach. $Nucleic\ Acids\ Res\ .\ 2010;38(suppl_2):W609-W614.$ doi:10.1093/nar/gkq300
- 33. Huang H, Yan G, Su H, et al. Clinical Outcome of an Multicentre, Randomized, Phase II Clinical Trial for Patients with Extranodal NK/T Cell Lymphoma Treated By P-Gemox or Aspametdex. Blood. 2019;134(Supplement_1):1569-1569. doi:10.1182/blood-2019-123478
- 34. Phung TTB, Sugamata R, Uno K, et al. Key role of regulated upon activation normal T-cell expressed and secreted, nonstructural protein1 and myeloperoxidase in cytokine storm induced by influenza virus PR-8 (A/H1N1) infection in A549 bronchial epithelial cells. $Microbiol\ Immunol\ .\ 2011;55(12):874-884.$ doi:10.1111/j.1348-0421.2011.00396.x
- 35. Lin Y-C, Shun C-T, Wu M-S, Chen C-C. A Novel Anticancer Effect of Thalidomide: Inhibition of Intercellular Adhesion Molecule-1-Mediated Cell Invasion and Metastasis through Suppression of Nuclear Factor- B. Clin Cancer Res. 2006;12(23):7165-7173. doi:10.1158/1078-0432.CCR-06-1393
- 36. El-Aarag B, Kasai T, Masuda J, Agwa H, Zahran M, Seno M. Anticancer effects of novel thalidomide analogs in A549 cells through inhibition of vascular endothelial growth factor and matrix metalloproteinase- $2.Biomed\ Pharmacother$. 2017;85:549-555. doi:10.1016/j.biopha.2016.11.063
- 37. Sun X, Xu Y, Wang Y, Chen Q, Liu L, Bao Y. Synergistic Inhibition of Thalidomide and Icotinib on Human Non-Small Cell Lung Carcinomas Through ERK and AKT Signaling. *Med Sci Monit* . 2018;24:3193-3203. doi:10.12659/MSM.909977
- 38. Cheng L, Li L. Systematic Quality Control Analysis of LINCS Data. CPT Pharmacometrics Syst Pharmacol . 2016;5(11):588-598. doi:10.1002/psp4.12107
- 39. Vinciguerra M, Romiti S, Greco E. Atherosclerosis as Pathogenetic Substrate for Sars-Cov2 "Cytokine Storm." Preprints . 2020:2020040430.
- 40. Guan W, Ni Z, Hu Y, et al. Clinical characteristics of 2019 novel coronavirus infection in China. N Engl J Med . Published online February 9, 2020:2020.02.06.20020974. doi:10.1101/2020.02.06.20020974
- 41. Steenblock C, Todorov V, Kanczkowski W, et al. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and the neuroendocrine stress axis. $Mol\ Psychiatry$. Published online May 7, 2020:1. doi:10.1038/s41380-020-0758-9
- 42. Lv Q, Yang Q, Cui Y, et al. Effects of taurine on ACE, ACE2 and HSP70 expression of hypothalamic-pituitary-adrenal axis in stress-induced hypertensive rats. Adv Exp Med Biol . 2017;975:871-886. doi: $10.1007/978-94-024-1079-2_69$
- 43. Chrousos GP, Kaltsas G. Post-SARS sickness syndrome manifestations and endocrinopathy: How, why, and so what? Clin Endocrinol (Oxf) . 2005;63(4):363-365. doi:10.1111/j.1365-2265.2005.02361.x
- 44. Jung YJ, Tweedie D, Scerba MT, Greig NH. Neuroinflammation as a Factor of Neurodegenerative Disease: Thalidomide Analogs as Treatments. Front Cell Dev Biol . 2019;7. doi:10.3389/fcell.2019.00313
- 45. Vallet S, Palumbo A, Raje N, Boccadoro M, Anderson KC. Thalidomide and lenalidomide: Mechanism-based potential drug combinations. *Leuk Lymphoma* . 2008;49(7):1238-1245. doi:10.1080/10428190802005191
- 46. Felsenstein S, Herbert JA, McNamara PS, Hedrich CM. COVID-19: Immunology and treatment options. Clin Immunol . 2020;215:108448. doi:10.1016/j.clim.2020.108448

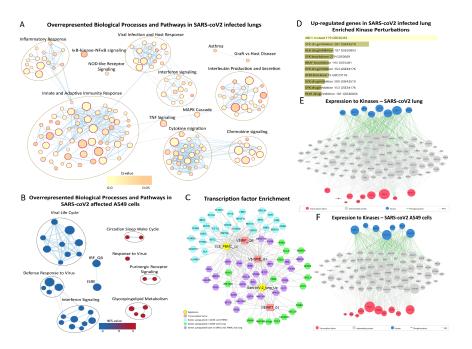
- 47. Spiegel M, Pichlmair A, Martinez-Sobrido L, et al. Inhibition of Beta Interferon Induction by Severe Acute Respiratory Syndrome Coronavirus Suggests a Two-Step Model for Activation of Interferon Regulatory Factor 3. $J\ Virol\ .\ 2005;79(4):2079-2086.\ doi:10.1128/JVI.79.4.2079-2086.2005$
- 48. Millrine D, Tei M, Gemechu Y, Kishimoto T. Rabex-5 is a lenalidomide target molecule that negatively regulates TLR-induced type 1 IFN production. *Proc Natl Acad Sci U S A* . 2016;113(38):10625-10630. doi:10.1073/pnas.1611751113
- 49. Millrine D, Miyata H, Tei M, et al. Immunomodulatory drugs inhibit TLR4-induced type-1 interferon production independently of Cereblon via suppression of the TRIF/IRF3 pathway. Int Immunol . 2016;28(6):307-315. doi:10.1093/intimm/dxw005
- 50. Taccone FS, Gorham J, Vincent J-L. Hydroxychloroquine in the management of critically ill patients with COVID-19: the need for an evidence base. $Lancet\ Respir\ Med$. Published online April 2020. doi:10.1016/S2213-2600(20)30172-7
- 51. Schafer PH, Ye Y, Wu L, et al. Cereblon modulator iberdomide induces degradation of the transcription factors Ikaros and Aiolos: immunomodulation in healthy volunteers and relevance to SLE. *Ann Rheum Dis* . 2018;77(10):1516-1523. doi:10.1136/annrheumdis-2017-212916
- 52. Lonial S, van de Donk NWCJ, Popat R, et al. First clinical (phase 1b/2a) study of iberdomide (CC-220), a CELMoD, in combination with dexamethasone (DEX) in patients (pts) with relapsed/refractory multiple myeloma (RRMM). J Clin Oncol . 2019;37(15_suppl):8006-8006. doi:10.1200/JCO.2019.37.15_suppl.8006
- 53. Mazzurana L, Forkel M, Rao A, et al. Suppression of Aiolos and Ikaros expression by lenalidomide reduces human ILC3-ILC1/NK cell transdifferentiation. $Eur\ J\ Immunol\ .\ 2019;49(9):1344-1355.$ doi:10.1002/eji.201848075
- 54. Palumbo A, Palladino C. Venous and arterial thrombotic risks with thalidomide: Evidence and practical guidance. Ther Adv Drug Saf. 2012;3(5):255-266. doi:10.1177/2042098612452291
- 55. Aue G, Njuguna N, Tian X, et al. Lenalidomide-induced upregulation of CD80 on tumor cells correlates with T-cell activation, the rapid onset of a cytokine release syndrome and leukemic cell clearance in chronic lymphocytic leukemia. Haematologica. 2009;94(9):1266-1273. doi:10.3324/haematol.2009.005835

Figure legends

- Figure 1: Characteristics of SARS-coV-2 infected lungs and A549 cells. Gene Set Enrichment Analysis (GSEA) of genes modulated in A) SARS-coV-2 infected lung and B) A549 cells. C) Transcription factor enrichment of genes up-regulated in SARS-coV-2 infection. D) Genes activated in SARS-coV-2 overlapping with genes down-regulated upon kinase perturbations. Up-regulated genes mapped to their transcription factors and kinases in E) SARS-coV-2 lung and F) A549 cells.
- Figure 2: Effect of Thalidomide and Lenalidomide on Immune System. GSEA network of gene expression profiles of A) thalidomide and B) lenalidomide treated A549 cells. C) Activities of kinase involved in immunomodulation and MAPK signaling in the presence of thalidomide D) Biological Pathways enriched by kinases affected by thalidomide.
- Figure 3: Comparative Enrichment of Biological Process. A) GO Biological Process enrichment comparison of gene expression signatures B) SARS-coV-2 infected tissues B) Overexpression of immune response genes in SARS-coV-2 tissues and suppression by thalidomide and lenalidomide C) B cell receptor signaling activated in PBMC of SARS-coV-2 patient and T cell activation in SARS-coV-2 lungs D) Modulation of ERK1 and ERK2 cascade in SARS-coV2 lung and thalidomide in A549.
- Figure 4: Activation of A) inflammation, B) TLR signaling, C) NOD-like signaling, D) Osteoblast differentiation and E) Cytokine signaling in SARS-coV-2 and genes targeted by thalidomide and lenalidomide in the signaling pathways.

Figure 5: Viral life cycle and defense response to virus. A) Up-regulation of pathways pertaining to viral entry, life cycle and antiviral response was observed in SARS-coV-2 infected lungs, PBMC, BALF and A549 cells and their modulation by thalidomide and lenalidomide. Overexpression of genes implicated in B) Interferon signaling and C) Type I interferon signaling in SARS-coV-2 infected lung and A549.

Figure 6: Enrichment of Disease-specific Phenotypes. A) Similarity of SARS-coV-2 signatures with disease-specific phenotypes. Differentially modulated genes in SARS-coV-2 and thalidomide treatment similar to profiles of B) lymphoma and D) Multiple Myeloma. C) Overlapping of SARS-coV-2 expression profile of SARS-coV-2 affected lung with systemic lupus erythematosus.



Effect of Thalidomide and Lenalidomide Treatment in A549 cells

