# Pantoea abscess mimicking sarcoma in a HTLV-1-infected Indigenous Australian man: case report and literature review

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# Abstract

Gram-negative bacteria of the genus Pantoea are emerging bacterial causes of diverse sporadic and outbreak-linked infections. Chronic Pantoea abscesses are unusual and may give rise to a differential diagnosis of malignancy. Foreign body retention and host immune defects may be risk factors for such chronic infections.

# *Pantoea* abscess mimicking sarcoma in a HTLV-1-infected Indigenous Australian man: case report and literature review

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#### Author contributions

MS – investigation, writing – original draft; JD – investigation, writing – original draft; RC – conceptualization, methodology, writing – original draft; writing – review and editing; supervision

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Pantoea; abscess; sarcoma; Enterobacteriaceae; HTLV-1; Indigenous Australians

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# KEY CLINICAL MESSAGE

Gram-negative bacteria of the genus Pantoea are emerging bacterial causes of diverse sporadic and outbreaklinked infections. Chronic *Pantoea* abscesses are unusual and may give rise to a differential diagnosis of malignancy. Foreign body retention and host immune defects may be risk factors for such chronic infections.

# INTRODUCTION

Members of the genus *Pantoea* are Gram-negative motile non-capsulated and non-spore forming rods belonging to the Enterobacteriaceae family. The genus was established in 1989 and its 20 species are ubiquitous, being found in plants, insects, and animals as well as water, soil, and air.(1) *Pantoea* has traditionally been regarded as plant pathogen, but certain species are increasingly recognized as emerging opportunistic causes of human disease, including nosocomial outbreaks.(2, 3) As such, while immunocompromise and contact with plant material and healthcare environments are risk factors for infections caused by *Pantoea*, its widespread ecological niche implies that sporadic infections without these risk factors are likely to occur.(1, 3)

#### CASE REPORT

A 55-year-old Indigenous Australian male living in remote Central Australia was admitted to Alice Springs Hospital in December 2015 for pain and swelling at the right calf resulting in a limp. For context, Central Australia is a rural region of over 1,000,000 km<sup>2</sup> comprising approximately 10% of the total Australian landmass (Figure 1). It is sparsely populated by about 60,000 people.(4) Of these, approximately 40% are Indigenous Australians who are more likely to suffer from numerous chronic diseases and have poor health literacy than their non-Indigenous counterparts.(5) Alice Springs Hospital is the referral facility for this vast remote area.

The patient's relapsing-remitting symptoms had begun in 2012 following a fall which, according to him, did not cause traumatic injury or skin penetration. He was reviewed in the Orthopaedic clinic in 2014 where he was noted to have an antalgic gait with a range of motion at the right knee of 90-140° and crepitus on movement, and a palpable medial gastrocnemius lesion. MRI of the right leg showed heterogeneous thickening of the medial gastrocnemius/soleus measuring  $35 \times 28 \times 147$  mm with unusual tubular extension into the central aspect of the medial gastrocnemius favouring post-injury haematoma, but raising the possibility of a neoplastic process (Figure 2). Unfortunately, he was lost to hospital follow-up until October 2015 when a further Orthopaedic outpatient review showed no change in the morphology or character of the lesion and continued observation was recommended.

His past medical history included poorly-controlled type 2 diabetes mellitus complicated by macroalbuminuria, hypertension, hyperlipidemia, obesity, complete heart block requiring pacemaker insertion, osteoarthritis of the right knee and left shoulder, and right leg varicose veins. His regular medications were aspirin 100 mg daily, atorvastatin 40 mg daily, gliclazide MR 120 mg daily, metformin XR 2 g daily, perindopril/indapamide 10/2.5 mg daily, sitagliptin 100 mg daily, and tadalafil 20 mg as required. On examination at the time of admission, there were distended superficial veins below the knee, and marked hard swelling and tenderness of the ankle and right calf to the level of the popliteal fossa. He was afebrile with a pulse rate of 96, respiratory rate of 22, blood pressure of 99/90 mmHg, and oxygen saturation of 97% breathing air on admission. C-reactive protein was raised at 177 mg/L (normal range 0-5), as he had a neutrophil leukocytosis with total white cell and neutrophil counts of  $16.3 \times 10^9$ /L (4-11) and  $10.4 \times 10^9$ /L (2.0-7.5), respectively.

A CT scan was performed to characterize the lesion further. This again showed a cystic morphologically heterogenous structure, but also that it had increased in size to  $38 \times 49 \times 156$  mm with a larger solid component, again raising the possibility of sarcoma (Figure 3). This differential diagnosis was also supported by the peripherally-enhancing, multi-loculated nature of the lesion. The patient remained an inpatient under the General Surgical team for three days, where he received empirical cefazolin 1 g eight-hourly. Aspiration of the lesion for histological and microbiological examination was planned, but deferred by the patient to February 2016 when it was performed under ultrasound guidance.

The aspirate demonstrated an inflammatory exudate comprising primarily neutrophils and pigment-laden macrophages compatible with abscess. No malignant cells were seen, but culture yielded pure growth of *Pantoea* spp. susceptible to amoxicillin/clavulanate, cefazolin, ciprofloxacin, gentamicin, and trimethoprim/sulfamethoxazole but resistant to ampicillin. Identification to species level was precluded by laboratory technological limitations but, given the culture result, the patient was referred to the Infectious Diseases clinic where he was seen in April 2016. He reported never feeling systemically unwell or febrile, and that, subsequent to the aspiration, the pain and swelling had improved to the extent he was able to walk normally. The calf mass remained palpable and hard to touch (Figure 4), but CRP had decreased to 11 and he no longer had a leukocytosis. Investigations for immunodeficiency revealed no lymphocyte deficiency and negative serology for HIV and *Strongyloides*, but he was infected with HTLV-1, in keeping with the very high prevalence in Indigenous Central Australians.(6) A detailed occupational and exposure history was taken, in which he denied performing agricultural work or gardening in the course of his work as an Aboriginal Health Worker or recreationally, but for cultural reasons would go into the desert surrounding his rural community when required.

To confirm that the lesion was, indeed, an abscess a second aspiration and tissue core biopsy was performed. Again, no malignant cells were seen and pure growth of *Pantoea* spp. was cultured. Insufficient tissue was obtained to draw a definitive histological conclusion, but the patient refused a second attempt as well as any surgical intervention. As such, he was commenced on amoxicillin/clavulanate 875/125 mg in May 2016 for six weeks, ciprofloxacin being contraindicated because of his history of arrythmia.

Despite successful completion of antibiotic therapy, the lesion was still obvious at follow-up in October 2016 and the patient reported similar symptom exacerbations in the preceding months which would spontaneously resolve. His case was, thus, discussed at a specialist sarcoma multi-disciplinary meeting where the consensus opinion was that an abscess, rather than malignancy, was the most likely diagnosis. Ongoing surveillance with imaging was recommended but the patient was lost to follow-up until 2020, when an ultrasound scan requested by his general practitioner showed persistence of the mass which had organized into three separate collections measuring  $150 \times 25$ mm,  $27 \times 10$  mm, and  $31 \times 28$  mm. The patient declined further intervention for this problem and, at the time this report was written in 2022, remained systemically well. Apart from the six weeks of amoxicillin/clavulanate commenced in 2016, he had not had any extended antibiotic courses for the abscess.

#### DISCUSSION

To place our case in context, we searched the MEDLINE database on 27 September 2022 to determine the spectrum of pathologies caused by *Pantoea* and associated risk factors. We used the search term '*Pantoea*' with no restriction on publication date, and applying the 'human' and 'English language' filters. This search yielded 278 results, from which 69 case reports, case series, and outbreak reports were selected for data extraction. Patients included in these publications were stratified by immune status and age, with children defined as those <18 years old. We also extracted data on clinical syndromes of sporadic cases, as well as sources of outbreaks. The literature review is summarized in the flowchart shown in Figure 5.

As is evident from the flowchart, *Pantoea* infections are rare, with only 248 cases reported worldwide between 1991 and 2022, 53 of which were outbreak cases stemming from various contaminated environmental sources. (7-11) Sporadic adult and paediatric cases were seen in equal proportions, with immunocompetent patients greatly outnumbering their immunocompromised counterparts. Bacteraemia, (14-36) followed by skin and soft tissue infections (SSTI), (31, 32, 37-42) were the commonest manifestations of *Pantoea* infection, but the range of syndromes was wide, including peritonitis, (32, 43-52) bone and joint infection, (32, 53-60) intraabdominal abscess, (61-63) pneumonia, (31, 64) urinary tract infection, (31, 32) ocular infection, (65-71) and rhinosinusitis. (72) One case each of endocarditis, (15) prosthetic joint infection, (55) and post-neurosurgical meningitis (73) was also reported, as were four cases of neonatal sepsis. (74, 75) Only in 19 patients, all of whom had either SSTI, (37, 41, 42) septic arthritis, (53, 56, 58, 59) or ocular infection (66, 69) was there a history of penetrating plant trauma, indicating that this risk factor is less important than previously assumed. The vast majority (84%) of infections were caused by *P. agglomerans*, with only four other culprit species reported: *P. dispersa*, (16, 17, 25, 27, 28, 72) *P. ananatis*, (19, 21, 68) *P. calida*, (24, 73) and *P. stewartii*. (18) In four other cases, identification to species level was unable to be performed, (39, 66, 67, 71)

like in ours. However, the diversity of *Pantoeas* pecies is not adequately differentiated by many laboratory identification methods and, as such, many cases attributed to *P. agglomerans* may, in fact, have been cause by other species. (76-78)

Our patient, therefore, exhibited a very atypical manifestation of a *Pantoea* SSTI mimicking malignancy, to the extent that both clinical and radiological findings raised concerns for sarcoma. Sarcoma is highly unlikely in our case, given the repeated pure growth of *Pantoea* spp., the long intervening period without death or deterioration, and the expert opinion provided by the specialist sarcoma multi-disciplinary meeting. While he denied any penetrating trauma, whether plant-related or otherwise, it is possible that micro-abrasions may have occurred when he fell, providing a portal of entry. Furthermore, given his rural residence, it is likely that plant or animal material was present on the ground, and his poor diabetic control likely contributed to the development and progression of infection.

Only one other case of *Pantoea* pseudotumour has been reported, although this patient from India provided a clear history of penetrating plant trauma due to his work in agriculture.(42) Like our case, this patient also reported a distant history of a fall four years prior to presentation. Regardless, both cases, despite the different management approaches taken, resulted in good outcomes. While surgical drainage of the lesion is ideal, as in the latter case, it is interesting that in our patient the infection was successfully contained with him remaining well ten years after symptom onset, even without surgical management. An important clinical lesson, therefore, is that foreign bodies, especially of plant origin which are not well-visualised on plain radiographs, may be retained following such trauma and act as foci of chronic inflammation leading to pseudotumour formation.

Our case raises some interesting questions. The first is whether the patient's HTLV-1 infection further predisposes to the establishment of a chronic bacterial infection. HTLV-1, unlike HIV, does not result in overt immunodeficiency, but associations between HTLV-1 and non-bacterial infections, such as scabies and strongylodiasis, are well-described.(79) However, little is known about how HTLV-1 mediates concurrent bacterial infections,(80) making this an important research question of clinical significance for the many infected Indigenous Central Australians in whom rates of bacterial infection far exceed those of their non-Indigenous countrymen,(79) as well as people living with this neglected tropical disease worldwide.

The second question arises from the ability of *Pantoea* to secrete products with bioremediative and immunogenic potential, facilitating its adaptation to diverse ecological niches, including in hostile environments.(1) It may well be that this has aided the establishment of a well-contained infective focus in our patient, given that macrophage activation and epithelial-mesenchymal transformation due to inflammatory mediators released by *P. agglomerans* leading to fibrosis has been recently reported.(81) Such a process may have succeeded in walling off the abscess, thus preventing cell-mediated immunity from eradicating the infection but also preventing the development of sepsis. Unfortunately, our laboratory was unable to speciate the causative organism, but other species may also have this capability. As such, research to elucidate the mechanisms of action of pathogen mediators released during *Pantoea* infections may be clinically useful.

# CONCLUSION

In conclusion, we have reported, to our knowledge, the second case of *Pantoea* pseudotumour in the literature, and the first in a patient with HTLV-1 infection. Through our analysis of this case, we have also identified areas in which further research will have clinically beneficial implications.

#### FUNDING

None required.

# POTENTIAL CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

#### PATIENT CONSENT STATEMENT

The authors confirm that the patient provided written consent for publication of this report.

# REFERENCES

- 1. Walterson AM, Stavrinides J. *Pantoea* : insights into a highly versatile and diverse genus within the Enterobacteriaceae. FEMS Microbiol Rev. 2015; 39(6):968-84.
- Dutkiewicz J, Mackiewicz B, Lemieszek MK, Golec M, Milanowski J. Pantoea agglomerans : a marvelous bacterium of evil and good.Part I. Deleterious effects: Dust-borne endotoxins and allergens - focus on cotton dust. Ann Agric Environ Med. 2015;22(4):576-88.
- Dutkiewicz J, Mackiewicz B, Kinga Lemieszek M, Golec M, Milanowski J. Pantoea agglomerans : a mysterious bacterium of evil and good. Part III. Deleterious effects: infections of humans, animals and plants. Ann Agric Environ Med. 2016;23(2):197-205.
- 4. Northern Territory Department of Health and Families. Central Australia regional plan 2010–2012. Darwin, Australia: Northern Territory Government; 2010.
- Australian Institute of Health and Welfare. The Health and Welfare of Australia's Aboriginal and Torres Strait Islander peoples 2015. Canberra, Australia: Australian Institute of Health and Welfare; 2015.
- 6. Einsiedel L, Pham H, Talukder MR, Taylor K, Wilson K, Kaldor J, et al. Very high prevalence of infection with the human T cell leukaemia virus type 1c in remote Australian Aboriginal communities: Results of a large cross-sectional community survey. PLOS Negl Trop Dis. 2021;15(12):e0009915.
- Boszczowski I, Nóbrega de Almeida Júnior J, Peixoto de Miranda EJ, Pinheiro Freire M, Guimarães T, Chaves CE, et al. Nosocomial outbreak of *Pantoea agglomerans* bacteraemia associated with contaminated anticoagulant citrate dextrose solution: new name, old bug? J Hosp Infect. 2012;80(3):255-8.
- 8. Bicudo EL, Macedo VO, Carrara MA, Castro FF, Rage RI. Nosocomial outbreak of *Pantoea agglome*rans in a pediatric urgent care center. Braz J Infect Dis. 2007;11(2):281-4.
- Koo HS, Kim JS, Eom JS, You JY, Park JY, Kim HS, et al. Pseudooutbreak of *Pantoea* species bacteremia associated with contaminated cotton pledgets. Am J Infect Control. 2006;34(7):443-6.
- Borrego Garcia E, Ruiz Sancho AL, Plaza Lara E, Díaz Gómez L, Delgado Ureña A. Bacteremia outbreak due to *Pantoea agglomerans* in hemodialysis, an infection by an unexpected guest. Nefrologia (Engl Ed). 2020;40(5):573-575.
- Yablon BR, Dantes R, Tsai V, Lim R, Moulton-Meissner H, Arduino M, et al. Outbreak of *Pantoea agglomerans* Bloodstream Infections at an Oncology Clinic-Illinois, 2012-2013. Infect Control Hosp Epidemiol. 2017;38(3):314-319.
- Habsah H, Zeehaida M, Van Rostenberghe H, Noraida R, Wan Pauzi WI, Fatimah I, et al. An outbreak of *Pantoea* spp. in a neonatal intensive care unit secondary to contaminated parenteral nutrition. J Hosp Infect. 2005;61(3):213-8.
- Liberto MC, Matera G, Puccio R, Lo Russo T, Colosimo E, Focà E. Six cases of sepsis caused by Pantoea agglomerans in a teaching hospital. New Microbiol. 2009;32(1):119-23.
- Cheng A, Liu CY, Tsai HY, Hsu MS, Yang CJ, Huang YT, et al. Bacteremia caused by *Pantoea agglomerans* at a medical center in Taiwan, 2000-2010. J Microbiol Immunol Infect. 2013;46(3):187-94.
- Wong KW. Pantoea agglomerans as a rare cause of catheter-related infection in hemodialysis patients. J Vasc Access. 2013;14(3):306.
- 16. Asai N, Koizumi Y, Yamada A, Sakanashi D, Watanabe H, Kato H, et al. *Pantoea dispersa* bacteremia in an immunocompetent patient: a case report and review of the literature. J Med Case Rep. 2019;13(1):33.
- Ruan XL, Qin X, Li M. Nosocomial bloodstream infection pathogen Pantoea dispersa : a case report and literature review. J Hosp Infect. 2022;127:77-82.
- Cobo F, González A, Pérez-Carrasco V, García-Salcedo JA. Pantoea stewartii : A new pathogen as a cause of bacteremia? Enferm Infecc Microbiol Clin (Engl Ed). 2022;40(5):278-280.
- Yoshimura M, Tokushige C, Maruyama JI, Kawano Y, Ishikura H, Matsunaga A, et al. Emerging resistance to beta-lactams in *Pantoea ananatis* isolated from an immunocompetent patient with bacteremia. Diagn Microbiol Infect Dis. 2022;102(4):115633.
- 20. Hagiya H, Otsuka F. Pantoea dispersa bacteremia caused by central line-associated bloodstream infec-

tion. Braz J Infect Dis. 2014;18(6):696-7.

- 21. De Baere T, Verhelst R, Labit C, Verschraegen G, Wauters G, Claeys G, et al. Bacteremic infection with *Pantoea ananatis*. J Clin Microbiol. 2004;42(9):4393-5.
- 22. Naha K, Ramamoorthi, Prabhu M. Spontaneous septicaemia with multi-organ dysfunction-a new face for *Pantoe agglomerans*? Asian Pac J Trop Med. 2012;5(1):83-4.
- 23. Uche A. *Pantoea agglomerans* bacteremia in a 65-year-old man with acute myeloid leukemia: case report and review. South Med J. 2008;101(1):102-3.
- 24. Yamada K, Kashiwa M, Arai K, Satoyoshi K, Nishiyama H. *Pantoea calida* bacteremia in an adult with end-stage stomach cancer under inpatient care. J Infect Chemother. 2017;23(6):407-409.
- Yang Y, Hu H, Zhou C, Zhang W, Yu Y, Liu Q, et al. Characteristics and accurate identification of *Pantoea dispersa* with a case of spontaneous rupture of hepatocellular carcinoma in China: A case report. Medicine (Baltimore). 2022;101(2):e28541.
- Christakis GB, Perlorentzou SP, Aslanidou M, Savva L, Zarkadis IK. Bacteremia caused by Pantoea agglomerans and Enterococcus faecalis in a patient with colon cancer. J BUON. 2007;12(2):287-90.
- 27. Schmid H, Schubert S, Weber C, Bogner JR. Isolation of a *Pantoea dispersa* -like strain from a 71-yearold woman with acute myeloid leukemia and multiple myeloma. Infection. 2003;31(1):66-7.
- Mehar V, Yadav D, Sanghvi J, Gupta N, Singh K. Pantoea dispersa : an unusual cause of neonatal sepsis. Braz J Infect Dis. 2013;17(6):726-8.
- Cicchetti R, Iacobini M, Midulla F, Papoff P, Mancuso M, Moretti C. Pantoea agglomerans sepsis after rotavirus gastroenteritis. Pediatr Infect Dis J. 2006;25(3):280-1.
- Barros Pinto MP, Marques G. An unexpected microbiological finding in a blood film. Br J Haematol. 2019;187(1):9.
- Büyükcam A, Tuncer Ö, Gür D, Sancak B, Ceyhan M, Cengiz AB, et al. Clinical and microbiological characteristics of *Pantoea agglomerans* infection in children. J Infect Public Health. 2018;11(3):304-309.
- Cruz AT, Cazacu AC, Allen CH. Pantoea agglomerans, a plant pathogen causing human disease. J Clin Microbiol. 2007;45(6):1989-92.
- 33. Lalas KM, Erichsen D. Sporadic *Pantoea agglomerans* bacteremia in a near-term female: case report and review of literature. Jpn J Infect Dis. 2010;63(4):290-1.
- Aly NY, Salmeen HN, Lila RA, Nagaraja PA. Pantoea agglomeransbloodstream infection in preterm neonates. Med Princ Pract. 2008;17(6):500-3.
- Oliveira MI, Batalha S, Gouveia C, Maia R, Kjöllerstrom P. Pantoea Species Bacteremia in a Child With Sickle Cell Disease: Looking for a Culprit. J Pediatr Hematol Oncol. 2017;39(6):e307-e308.
- Fernández-Muñoz H, Lassaletta A, González MJ, Andión M, Madero L. Pantoea agglomerans bacteremia in a child with acute lymphoblastic leukemia during induction therapy. J Pediatr Hematol Oncol. 2015;37(4):328.
- Alpiste FO, Ezquerra GM, Pujol RM. Wound infection by *Pantoea agglomerans* after penetrating plant injury. Indian J Dermatol Venereol Leprol. 2022;88(5):633-635.
- Vaiman M, Lazarovich T, Lotan G. Pantoea agglomerans as an indicator of a foreign body of plant origin in cases of wound infection. J Wound Care. 2013;22(4):182, 184-5.
- Kircheva A, Paskalev D. Fulminant infection of the lower limb in a diabetic patient. J Wound Care. 2007;16(5):223-4.
- 40. Ajam M, Shokr M, Ajam F, Lieberman R. Rare case of implantable cardioverter defibrillator infection caused by *Stenotrophomonas maltophilia* and *Pantoea calida*. BMJ Case Rep. 2019;12(8):e230506.
- 41. Harris EJ. Retained Hawthorn fragment in a child's foot complicated by infection: diagnosis and excision aided by localization with ultrasound. J Foot Ankle Surg. 2010;49(2):161-5.
- 42. Jain S, Bohra I, Mahajan R, Jain S, Chugh TD. *Pantoea agglomerans* infection behaving like a tumor after plant thorn injury: an unusual presentation. Indian J Pathol Microbiol. 2012;55(3):386-8.
- Sastre A, González-Arregoces JE, Romainoik I, Mariño S, Lucas C, Monfá E, et al. Peritonitis caused by *Pantoea agglomerans* in peritoneal dialysis. Nefrologia. 2017;37(1):108-109.
- 44. Kazancioglu R, Buyukaydin B, Iraz M, Alay M, Erkoc R. An unusual cause of peritonitis in peritoneal dialysis patients: *Pantoea agglomerans*. J Infect Dev Ctries. 2014;8(7):919-22.

- Lim PS, Chen SL, Tsai CY, Pai MA. Pantoea peritonitis in a patient receiving chronic ambulatory peritoneal dialysis. Nephrology (Carlton). 2006;11(2):97-9.
- 46. Habhab W, Blake PG. Pantoea peritonitis: not just a "thorny" problem. Perit Dial Int. 2008;28(4):430.
- 47. Chen KJ, Chen TH, Sue YM. *Citrobacter youngae* and *Pantoea agglomerans* peritonitis in a peritoneal dialysis patient. Perit Dial Int. 2013;33(3):336-7.
- 48. Kahveci A, Asicioglu E, Tigen E, Ari E, Arikan H, Odabasi Z, et al. Unusual causes of peritonitis in a peritoneal dialysis patient: *Alcaligenes faecalis* and *Pantoea agglomerans*. Ann Clin Microbiol Antimicrob. 2011;10:12.
- Ferrantino M, Navaneethan SD, Sloand JA. Pantoea agglomerans : an unusual inciting agent in peritonitis. Perit Dial Int. 2008;28(4):428-30.
- Borràs M, Roig J, Garcia M, Fernández E. Adverse effects of *Pantoea* peritonitis on peritoneal transport. Perit Dial Int. 2009;29(2):234-5.
- Magnette C, Tintillier M, Horlait G, Cuvelier C, Pochet JM. Severe peritonitis due to Pantoea agglomerans in a CCPD patient. Perit Dial Int. 2008;28(2):207-8.
- 52. Lau KK, Ault BH, Jones DP. Polymicrobial peritonitis including *Pantoea agglomerans* from teething on a catheter. South Med J. 2005;98(5):580-1.
- 53. Duerinckx JF. Case report: subacute synovitis of the knee after a rose thorn injury: unusual clinical picture. Clin Orthop Relat Res. 2008;466(12):3138-42.
- 54. Arlet G, Grateau G, Lionnet F. Bilateral tibial chronic osteomyelitis due to *Pantoea agglomerans* in a patient with sickle cell disease. Rheumatology (Oxford). 2007;46(8):1247.
- 55. Hischebeth GT, Kohlhof H, Wimmer MD, Randau TM, Bekeredjian-Ding I, Gravius S. Detection of *Pantoea agglomerans* in hip prosthetic infection by sonication of the removed prosthesis: the first reported case. Technol Health Care. 2013;21(6):613-8.
- 56. De Champs C, Le Seaux S, Dubost JJ, Boisgard S, Sauvezie B, Sirot J. Isolation of *Pantoea agglome-rans* in two cases of septic monoarthritis after plant thorn and wood sliver injuries. J Clin Microbiol. 2000;38(1):460-1.
- 57. Rave O, Assous MV, Hashkes PJ, Lebel E, Hadas-Halpern I, Megged O.*Pantoea agglomerans* foreign body-induced septic arthritis. Pediatr Infect Dis J. 2012;31(12):1311-2.
- 58. Ulloa-Gutierrez R, Moya T, Avila-Aguero ML. *Pantoea agglomerans* and thorn-associated suppurative arthritis. Pediatr Infect Dis J. 2004;23(7):690.
- 59. Kratz A, Greenberg D, Barki Y, Cohen E, Lifshitz M. *Pantoea agglomerans* as a cause of septic arthritis after palm tree thorn injury; case report and literature review. Arch Dis Child. 2003;88(6):542-4.
- 60. Labianca L, Montanaro A, Turturro F, Calderaro C, Ferretti A. Osteomyelitis caused by *Pantoea* agglomerans in a closed fracture in a child. Orthopedics. 2013;36(2):e252-6.
- Gourgiotis S, Kantounakis I, Falidas E, Exarchou E, Kalamara P, Villias C. Pyogenic Intraperitoneal Pantoea agglomeransAbscesses. Surg Infect (Larchmt). 2016;17(1):119-20.
- Fullerton DG, Lwin AA, Lal S. Pantoea agglomerans liver abscess presenting with a painful thigh. Eur J Gastroenterol Hepatol. 2007;19(5):433-5.
- Rodrigues AL, Lima IK, Koury A Jr, de Sousa RM, Meguins LC. Pantoea agglomerans liver abscess in a resident of Brazilian Amazonia. Trop Gastroenterol. 2009;30(3):154-5.
- 64. Shubov A, Jagannathan P, Chin-Hong PV. *Pantoea agglomerans* pneumonia in a heart-lung transplant recipient: case report and a review of an emerging pathogen in immunocompromised hosts. Transpl Infect Dis. 2011;13(5):536-9.
- Sudhalkar A, Majji AB, Chhablani J, Manderwad G. Pantoea agglomerans endophthalmitis: clinical features and outcomes. Retina. 2014;34(8):1702-6.
- Lee NE, Chung IY, Park JM. A case of *Pantoea* endophthalmitis. Korean J Ophthalmol. 2010;24(5):318-21.
- 67. Kletke SN, Brissette AR, Gale J. Endogenous bacterial endophthalmitis caused by *Pantoea* species: a case report. Can J Ophthalmol. 2014;49(1):e1-2.
- 68. Comba OB, Pehlivanoglu S, Bayraktar Z, Albayrak S, Karakaya M. *Pantoe Agglomerans* Endophthalmitis after Phaco Surgery: The First Case in Literature. Ocul Immunol Inflamm. 2020;28(3):479-482.

- 69. Manoharan G, Lalitha P, Jeganathan LP, Dsilva SS, Prajna NV. *Pantoea ananatis* as a cause of corneal infiltrate after rice husk injury. J Clin Microbiol. 2012;50(6):2163-4.
- 70. Seok S, Jang YJ, Lee SW, Kim HC, Ha GY. A case of bilateral endogenous *Pantoea agglomerans* endophthalmitis with interstitial lung disease. Korean J Ophthalmol. 2010;24(4):249-51.
- Zuberbuhler B, Carifi G, Leatherbarrow B. Acute dacryocystitis in a 2-year old child caused by pantoea. Orbit. 2012;31(1):13-4.
- Su YW, Huang WH, Yeh CF. Pantoea dispersa rhinosinusitis: clinical aspects of a rare sinonasal pathogen. Eur Arch Otorhinolaryngol. 2022;279(9):4389-4395.
- Fritz S, Cassir N, Noudel R, De La Rosa S, Roche PH, Drancourt M. Postsurgical Pantoea calida meningitis: a case report. J Med Case Rep. 2014;8:195.
- Tiwari S, Beriha SS. Pantoea species causing early onset neonatal sepsis: a case report. J Med Case Rep. 2015;9:188.
- Bergman KA, Arends JP, Schölvinck EH. Pantoea agglomeranssepticemia in three newborn infants. Pediatr Infect Dis J. 2007;26(5):453-4.
- 76. Rezzonico F, Stockwell VO, Tonolla M, Duffy B, Smits TH. *Pantoea* clinical isolates cannot be accurately assigned to species based on metabolic profiling. Transpl Infect Dis. 2012;14(2):220-1.
- Rezzonico F, Smits TH, Duffy B. Misidentification slanders *Pantoea agglomerans* as a serial killer. J Hosp Infect. 2012;81(2):137-9.
- Delétoile A, Decré D, Courant S, Passet V, Audo J, Grimont P, et al. Phylogeny and identification of *Pantoea* species and typing of *Pantoea agglomerans* strains by multilocus gene sequencing. J Clin Microbiol. 2009;47(2):300-10.
- Einsiedel L, Spelman T, Goeman E, Cassar O, Arundell M, Gessain A. Clinical associations of Human T-Lymphotropic Virus type 1 infection in an indigenous Australian population. PLoS Negl Trop Dis. 2014;8(1):e2643.
- 80. Rosadas C, Taylor GP. HTLV-1 and Co-infections. Front Med (Lausanne). 2022;9:812016.
- Lemieszek MK, Rzeski W, Golec M, Mackiewicz B, Zwoliński J, Dutkiewicz J, et al. Pantoea agglomerans chronic exposure induces epithelial-mesenchymal transition in human lung epithelial cells and mice lungs. Ecotoxicol Environ Saf. 2020;194:110416.

#### FIGURE LEGENDS

Figure 1. Map of Australia highlighting the Central Australia region (shaded) and the location of the town of Alice Springs (red marker).

Figure 2. MRI axial section of the right calf showing the location of the pseudotumour (arrow) between the gastrocnemius (G) and soleus (S) muscles.

Figure 3. CT sagittal section of the right leg showing the extent of the multi-loculated pseudotumour (arrows) within the calf.

Figure 4. Photograph of the right calf mass four years after its appearance.

Figure 5. Literature review flowchart showing sources and clinical syndromes for outbreaks and sporadic Pantoea infections, respectively. Immunocompromise in the paediatric population included age <28 days.









