

The evolution of chemodiversity - From verbal to quantitative models

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Abstract

Plants harbour a great chemodiversity, i.e., diversity of specialized metabolites (SMs), at different scales. For instance, individuals can produce a large number of SMs and populations can differ in their metabolite composition. Given the ecological and economic importance of plant chemodiversity, it is important to understand how it arises and is maintained over evolutionary time. For other dimensions of biodiversity, i.e., species diversity and genetic diversity, quantitative models play an important role in addressing such questions. Here we provide a synthesis of existing hypotheses and quantitative models, i.e. mathematical models and computer simulations, for the evolution of plant chemodiversity. We describe each model's ingredients, i.e., the biological processes that shape chemodiversity, the scales it considers, and whether it has been formalized as a quantitative model. Although we identify several quantitative models, not all are dynamic and many influential models have remained verbal. To fill these gaps, we identify quantitative models used for genetic variation that may be adapted for chemodiversity. We end by outlining our vision for the future of chemodiversity modeling, presenting a flexible framework for the creation of individual-based models that address different scales of chemodiversity and combine different ingredients that bring this chemodiversity about.

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30 **Abstract**

31 Plants harbour a great *chemodiversity*, i.e., diversity of specialized metabolites (SMs), at different scales.
32 For instance, individuals can produce a large number of SMs and populations can differ in their metabolite
33 composition. Given the ecological and economic importance of plant chemodiversity, it is important to
34 understand how it arises and is maintained over evolutionary time. For other dimensions of biodiversity,
35 i.e., species diversity and genetic diversity, quantitative models play an important role in addressing such
36 questions. Here we provide a synthesis of existing hypotheses and quantitative models, i.e. mathematical
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39 has been formalized as a quantitative model. Although we identify several quantitative models, not all are
40 dynamic and many influential models have remained verbal. To fill these gaps, we identify quantitative
41 models used for genetic variation that may be adapted for chemodiversity. We end by outlining our vision
42 for the future of chemodiversity modeling, presenting a flexible framework for the creation of individual-
43 based models that address different scales of chemodiversity and combine different ingredients that bring
44 this chemodiversity about.

1 Introduction

Chemodiversity is the diversity of specialized metabolites (SMs) at different levels of organisation – from single tissues to entire communities (see Wetzels & Whitehead, 2020, for a review). SMs are compounds produced by an organism that are not directly involved in basic physiological mechanisms such as photosynthesis and reproduction, but may be important for the interaction with herbivores, pollinators, and conspecifics, or protect against abiotic stresses (Wetzels & Whitehead, 2020). For many SMs, the function is currently unknown. SMs are also often called 'secondary metabolites' (Dyer *et al.*, 2018; Moore *et al.*, 2014; Rokas *et al.*, 2020; Stone & Williams, 1992), 'phytochemicals' (in plants, Defosse *et al.*, 2021; Richards *et al.*, 2016), or 'natural products' (Firn & Jones, 2003). SMs belong to metabolite classes including alkaloids, terpenoids, or flavonoids. If individuals within a species differ in the metabolite composition within such metabolite classes, they are often divided into distinct chemotypes. For example, in *Tanacetum vulgare* (Lokki *et al.*, 1973) or *Thymus vulgaris* (Thompson *et al.*, 2003), different terpenoid chemotypes differing in terpenoid composition can be found.

Chemodiversity can be quantified in various ways (Petrén *et al.*, 2023; Wetzels & Whitehead, 2020): it can focus on a group of metabolites belonging to one or several metabolite classes or large parts of the metabolome, and it can be measured within as well as between units of scale. For example, both the number of SMs per individual and the differences in SMs between individuals in a population are aspects of chemodiversity.

The existence of widespread chemodiversity is puzzling from an evolutionary perspective. SMs are often synthesized in complex metabolic pathways that involve multiple enzymes modifying a precursor metabolite into the SM over several steps (Firn & Jones, 2003, 2009; Jones & Firn, 1991). Given the inherent costs of these pathways, one would expect evolution towards a small number of the most beneficial metabolites (Jones & Firn, 1991). Despite this, a high diversity of SMs has been found within and between plant populations and to some extent also in fungal and bacterial populations (Calf *et al.*, 2018; Defosse *et al.*, 2021; Li *et al.*, 2020; Rhoades *et al.*, 1976; Rokas *et al.*, 2020). Evidently, there are mechanisms for the maintenance of this chemodiversity. These mechanisms are what models around chemodiversity attempt to elucidate.

For other dimensions of biodiversity such as species diversity and genetic diversity, quantitative models – both mathematical and simulation models – have long been an important part of scientific inquiry (see e.g. Hubbell, 2001; Kimura, 1983; Wright, 1937). They are for instance used as proof-of-concept models to test the validity of verbal models (Servedio *et al.*, 2014), to generate predictions and hypotheses that can be tested empirically (Servedio *et al.*, 2014), and to estimate parameters from data. Running *in silico* experiments

76 with quantitative models can give ideas about those aspects of a system that are the most relevant for an
77 outcome or quantity of interest, and thus about which measurements should be taken in an experiment. In
78 that way they can make empirical studies more efficient, and sometimes provide a statistical model that can
79 be fit to data. Additionally, a good quantitative model can unify several studies that differ in methodology so
80 they form a coherent narrative (Otto & Rosales, 2019). Because of these various contributions of quantitative
81 models to scientific inquiry, we argue that chemodiversity should likewise be investigated in this way.

82 A better understanding of the evolution of chemodiversity that might be conferred by such models also
83 has an applied relevance. For instance, models that predict local adaptation and geographic structure
84 of chemodiversity (Calf *et al.*, 2018; Defosse *et al.*, 2021), could be taken into account in conservation
85 management, as implemented already for genetic diversity (see e.g. Frankham *et al.*, 2002, Ch. 16). Moreover,
86 models for the evolution of chemodiversity might help predict why some introductions of plants or herbivores
87 to new places succeed while others do not, which will be discussed further in Box 3.

88 In this article, we synthesize the work that has been done so far on developing verbal and, in particular,
89 quantitative models for chemodiversity. For each model, we discuss which biological processes it focuses
90 on and at which scale it addresses chemodiversity. We are not the first to review possible explanations for
91 chemodiversity. There are for instance the excellent reviews by Dyer *et al.* (2018), Kessler & Kalske (2018),
92 Moore *et al.* (2014), Stamp (2003), and Wetzal & Whitehead (2020); therefore we do not want to dwell
93 on reviewing the empirical support for the different explanations for chemodiversity. Instead, we focus on
94 how the verbal models described in those reviews have been tested through quantitative models, and discuss
95 how quantitative models for other types of diversity could be adapted to modeling chemodiversity. We also
96 discuss some of the empirical work that has been done on them to elucidate how quantitative models can
97 connect verbal models and empirical studies. We end by outlining an individual-based modeling framework
98 for chemodiversity and important avenues for future research.

99 2 Chemodiversity models

100 There is currently no single unifying theory that can explain chemodiversity at all scales and within as well
101 as between units at each scale. However, there are numerous hypotheses and verbal models (Fig. 1), each
102 of which focuses on a different set of “ingredients”, i.e., biological players and processes that can contribute
103 to shaping chemodiversity (Fig. 3, Box 1). Moreover, the different hypotheses and models cover different
104 scales of organization (Fig. 2, Box 2), and can be modeled in different ways (Fig. 4, Box 4). Although there

105 are connections and overlaps between many hypotheses and they are certainly not all mutually exclusive, to
106 provide some structure for this review, we have placed all models into a hierarchy and grouped similar models
107 together (Fig. 1). Note that sometimes different terms are used in different publications to describe the
108 same or similar models (for instance in the reviews of Dyer *et al.*, 2018; Stamp, 2003; Wetzel & Whitehead,
109 2020).

110 Although these various hypotheses on the development and maintenance of chemodiversity are commonly
111 used to generate hypotheses for empirical research (Li *et al.*, 2020; Whitehead *et al.*, 2021), there are fewer
112 quantitative models of these hypotheses. Here, we will review extant quantitative models of chemodiversity,
113 as well as research that could serve as a starting point for the creation of models to fill the numerous gaps
114 in the state of the art.

115 3 Model descriptions

116 3.1 Models based on resource constraints

117 The SMs an organism produces are limited by the resources the organism has access to. Furthermore, the
118 organism may optimize SM production subject to constraints based on available resources or other abiotic
119 or biotic factors (Fig. 3). These ideas are the basis for a group of models that aim to explain variation in
120 quantitative defense level depending on abiotic and biotic conditions. For instance, according to the **carbon**
121 **nutrient balance hypothesis**, the relative availability of carbon and nitrogen determines which types
122 of compounds are preferentially produced (Bryant *et al.*, 1983). For instance, in low-carbon (i.e. shady)
123 habitats, likely more nitrogen-based toxins are produced. While the hypothesis at times correctly predicts
124 the chemicals which are found in empirical studies, it has come under criticism for frequently failing to do
125 so (Hamilton *et al.*, 2008).

126 Many of the hypotheses in this group are based on the idea that a plant's investment in defense competes
127 with investment into growth, reproduction, and storage. According to the **resource availability hypoth-**
128 **esis** (Coley *et al.*, 1985; Hahn & Maron, 2016), plants in environments with low resource availability are
129 selected to grow slowly and be well defended, whereas plants grown at high resource availability are selected
130 to grow fast and invest less in defense. The **growth-differentiation hypothesis** (Herms & Mattson, 1992)
131 and the **coordinated resource allocation hypothesis** (Monson *et al.*, 2021) expand on the resource
132 availability hypothesis by modeling the maximum possible growth rate in more detail and by adding reserve
133 safety margins, i.e., storage pools that allow plants to plastically respond to biotic and abiotic challenges

134 (Fig. 3). Coley *et al.* (1985) also propose an equation (which is then extended by Herms & Mattson, 1992;
135 Monson *et al.*, 2021) modeling how plant growth rate depends on investment in defense. In these models (at
136 least with the selected parameter values), plant growth rate is optimized at intermediate defense levels and
137 the optimal strategy depends on the maximum possible growth rate in the absence of herbivores, i.e. resource
138 availability. Thus, if the microenvironments of individuals in a population differ in resource availability, these
139 hypotheses thus offer a quantitative prediction for differences in defense levels between individuals within a
140 population (Fig. 2). Analogously, differences between different populations of the same species are predicted
141 if the habitats of the populations differ in resource availability. Similarly, in a verbal model, Vannette &
142 Hunter (2011) assume that the expression of plant defenses has a nonlinear relationship with the density of
143 arbuscular mycorrhizal fungi (AMF), such that variation in plant defense between populations in different
144 habitats could be explained by variation in AMF density.

145 Differences in defense levels between individuals or populations in response to environmental conditions
146 are often based on phenotypic plasticity. Many SMs are not produced continuously, but induced only
147 when needed, being then beneficial. For example, exposure to higher ultraviolet-B radiation induces the
148 production of flavonoids, which are important SMs in photoprotection (Agati & Tattini, 2010). Using
149 a simple mathematical framework called **error-management theory**, Orrock *et al.* (2015) computed the
150 optimal herbivore cue strength at which a plant should express its induced defense. This threshold represents
151 a balance between the costs of a false alarm and the costs of being attacked unprepared. Since these costs
152 but also the exposure to herbivores might differ between different plant individuals and populations (Fig. 3),
153 this model could explain chemodiversity in expressed defense between individuals and between populations
154 (Fig. 2).

155 Clearly, plasticity cannot explain all differences in defense expression and many differences are also
156 genetically determined (see e.g. Garrido *et al.*, 2012). The SMs of populations and species as a whole can
157 be adapted to the typical resources in their location, consequently affecting their chemodiversity (Defossez
158 *et al.*, 2021; Hahn & Maron, 2016; Stamp, 2003). However, at the intraspecific level patterns appear to be
159 more complex, in part because locations with high resource availability also tend to be locations with high
160 herbivore pressure, such that it is still unclear how much of intraspecific defense variation can be explained
161 by the resource-availability hypothesis and other models based on constraints (Hahn & Maron, 2016).

162 **3.2 Apparency**

163 Feeny (1976) developed the **plant apparency hypothesis** as a verbal model where plants in a community

164 produce different SMs depending on how easy it is for herbivores to find them relative to other plants in the
165 community (Fig. 3): highly apparent plants (e.g. oak trees) produce certain SMs in high concentrations,
166 so called quantitative defenses, that affect most herbivores by acting as digestibility-reducers; less apparent
167 species (e.g. small annual plants) produce toxic SMs in low concentrations, so called qualitative defenses,
168 which some herbivores are immune to. The apparency hypothesis addresses chemodiversity at the community
169 level as well as between clades (see Fig. 2).

170 Partial support that the verbal apparency model can work in principle comes from a quantitative model
171 by Yamamura & Tsuji (1995). They used an optimal control theory approach to find the optimal investment
172 in growth vs. defense over the life time of the plant. With apparency understood as herbivory pressure,
173 their model results supported the apparency hypothesis. They found that more apparent plants produced
174 quantitative defenses whereas less apparent ones did not, and that for those plants who produced quantitative
175 defenses, the investment in defense increased with apparency. With apparency understood as length of the
176 growth period, only the first prediction was true and only under some parameter settings.

177 The plant apparency hypothesis has been criticized for being difficult to test in practice. SMs are not
178 easily divided into quantitative and qualitative defenses, but often have properties of both. This makes
179 it difficult to correlate apparency to the presence of quantitative and qualitative defenses (Stamp, 2003).
180 However, which SMs are beneficial to particular plants in a population still might differ depending on their
181 apparency.

182 **3.3 Mechanistic explanations of chemodiversity**

183 Hypotheses in this group focus on why having multiple defense SMs is beneficial to plants, but they do not
184 explicitly consider evolution on the herbivore side. Those hypotheses will be covered in the next section.

185 The **synergy hypothesis** proposes that the effect of a mixture of SMs is more than the sum of the effects
186 of each SM in isolation. These nonlinear effects of mixtures of SMs then explain why organisms produce
187 multiple SMs. On the molecular level, synergistic effects are explained by molecular interactions between
188 SMs, for example by one SM facilitating the movement of another SM across cell membranes (Richards *et al.*,
189 2016). The synergy hypothesis addresses chemodiversity within tissues and within individuals (see Fig. 2,
190 and Wetzal & Whitehead, 2020). In empirical studies, it was found that mixtures of SMs affect generalist
191 herbivores more than specialists (Richards *et al.*, 2016). Synergistic effects are difficult to study in the lab
192 in part because it is difficult to isolate large enough amounts of the SMs involved (Dyer *et al.*, 2018). In
193 this and similar situations, quantitative modeling can be a useful tool to formulate hypotheses and guide

194 experiments. So far, to our knowledge, such models do not exist.

195 The **moving target hypothesis** is a term proposed by Adler & Karban (1994) and used by Li *et al.*
196 (2020) to refer to an inducible defense where a plant in a population randomly changes its phenotype in
197 response to an attack by herbivores (Fig. 3). This is closely related to the 'novel weapons hypothesis' that
198 is proposed in biological invasions (Box 3). Herbivores then cannot adjust effectively to this defense as it
199 is unpredictable. Wetzel & Whitehead (2020) use the term more broadly to cover hypotheses that describe
200 changes in SMs within individuals over time, functioning like a moving target that herbivores have a hard
201 time adjusting to, even if it does not follow the precise mechanisms described in Adler & Karban (1994).
202 Relatedly, variation between plant individuals in a population can suppress herbivores because they need
203 to adjust their detoxification mechanism when moving between plants with different defense metabolites,
204 which costs energy and is not instantaneous (Pearse *et al.*, 2018). However, the conditions under which this
205 phenomenon would provide an advantage to individuals with rare metabolites and promote the evolution of
206 chemodiversity between individuals are not well understood yet and would probably depend on the details
207 of herbivore behavior.

208 Adler & Karban (1994) analysed a quantitative, stochastic, plant-herbivore model. In this model, plants
209 could have three defense strategies: a strategy that does not adjust to herbivore damage (constitutive
210 defense), one where plants adjust to herbivore damage by randomly changing their defense (moving target),
211 or one where all plants switch to whichever defense results in the highest growth rate in the current situation
212 (optimal inducible defense). They found that in an environment with a fluctuating number of herbivores,
213 the optimal inducible defense is superior when there is just one herbivore species. When there are at least
214 two herbivores with different resistances to defense strategies, the moving target strategy is superior as long
215 as the cost of defense is not too high. When the cost of defense is high, low levels of constitutive defense are
216 superior.

217 Eagle-eyed readers will have noticed that this model only included chemodiversity on the within popu-
218 lation and between-individual level, not on the within-individual level, although this is part of the verbal
219 model (Fig. 2). It is arguable that different defense strategies can be analogous to chemotypes with different
220 defensive properties. To make this an explicit chemodiversity model capable of investigating chemodiversity
221 on the individual level, a model would need to be added that simulates the actual chemotypes.

222 The **interaction diversity hypothesis** (Kessler & Kalske, 2018; Whitehead *et al.*, 2021) suggests that
223 plants possess a variety of SMs because they interact with a large number of other organisms (both antagonists
224 and mutualists, as well as interspecific or intraspecific competitors, see Fig. 3) and different metabolites are

225 active in interaction with the different organisms. Independent selection on each metabolite, potentially in
226 coevolution with the respective interacting species, then leads to the emergence of chemodiversity. Although
227 the name of the hypothesis is rather new, the idea is relatively old and has also been called the “common-
228 sense scenario” (Berenbaum & Zangerl, 1996). Although the interaction diversity hypothesis mostly seeks to
229 explain the diversity of SMs within an individual (Fig. 2, with empirical support by Whitehead *et al.*, 2021),
230 the reasoning can be extended to explain chemodiversity at other scales. For example, if two populations or
231 species differ in the set of interacting animals, we would expect differences in their metabolite composition.
232 Although the ideas of multiple interaction partners driving chemodiversity is very prominent and well-
233 supported, there is currently no quantitative model based on this idea.

234 3.4 Evolutionary games

235 Models and hypotheses explaining plant chemodiversity or variation in defenses via evolutionary games
236 focus on the competition of plants ”playing” different defense strategies. That is, they have interactions
237 with other plants as important ingredient (Fig. 3). One way in which variation in defense traits can be
238 maintained between individuals in a population and within populations (see Fig. 2) is **negative frequency-
239 dependence** where the respective rare phenotype or genotype has an advantage. Sato *et al.* (2017) and
240 Augner *et al.* (1991) created quantitative models of evolutionary change in plant defense in populations over
241 time. Sato *et al.* (2017) found a pattern of negative frequency-dependent selection that could lead to stable
242 coexistence of undefended plants and defended plants which paid a cost for their defense. The underlying
243 mechanism is that when defended plants become too common, optimally foraging herbivores start to forage
244 also on defended plants despite lower profitability because it is too costly to search for the few undefended
245 plants. This matched up well with their data on a field site of *Arabidopsis halleri* that comprised hairy and
246 glabrous morphs. Similarly, Augner *et al.* (1991), showed in a game-theoretical model of a grazed population
247 with two morphs, that both morphs could coexist if the fitness benefit a defended player has if an undefended
248 opponent is grazed is higher than the profit a non-defended player makes if another non-defended opponent
249 is grazed. These models show that diversity, including chemodiversity, can exist when there is a benefit to
250 being the rare morph in a mixed population.

251 A coexistence mechanism closely related to negative frequency-dependence is a rock-paper-scissors game.
252 This appears to explain the coexistence of *Brassica nigra* genotypes with high levels of the defense metabolite
253 sinigrin, low-sinigrin *B. nigra* genotypes, and other competing species (Lankau & Strauss, 2007). Here the
254 high-sinigrin type has an advantage in the competition with other plant species and thus has an advantage

255 when *B. nigra* is rare. As *B. nigra* is then becoming more common and intraspecific competition becomes
256 stronger, it no longer pays to produce so much sinigrin which is not effective against conspecifics, and the
257 low-sinigrin type has an advantage. These dynamics were captured in an individual-based simulation model
258 by Lankau (2009).

259 **Associational effects** are the effects that plants have on each other without direct interaction with
260 each other but through interaction with other species, in particular herbivores (Hambäck *et al.*, 2014) (Fig.
261 3). These exist under many names. One kind of associational effect is social heterosis, which can explain
262 chemodiversity at the within-population scale. Social heterosis refers to a scenario wherein individuals have
263 different traits that are beneficial both to themselves and others in their neighbourhood but cannot all exist
264 in one organism at the same time. In this case, individuals in a diverse group have higher fitness than those
265 in a monoculture, as demonstrated in a quantitative model by Nonacs & Kapheim (2007). In chemodiversity,
266 this could take the form of a population of plants where different plants have different chemical profiles which
267 repel different insect herbivores, both from the individual plant itself and its neighbours. If the production
268 of SMs is costly, it may be detrimental to an individual to produce too many SMs, but beneficial to be in
269 an environment where other individuals produce SMs different from oneself. In this way, social heterosis
270 generates negative frequency-dependent selection on a single or community scale (Fig. 2) as rare types are
271 more likely to find themselves in mixed neighbourhoods with other types. This type of associational effect
272 is also called associational resistance (increased resistance through associational effects).

273 Another example of associational effects of chemodiversity is the model by Hambäck *et al.* (2014), which
274 modeled associational effects in a community of plants with different traits involved in both visual and
275 olfactory detection by herbivores. Volatile SMs form odor plumes that can be detected by herbivores. The
276 model included both repellent and attractant traits, different rates of detection of these plumes from outside
277 the patch by herbivores, the relative attraction to different types of plants, and the within- and without-
278 patch migration rates. Their model found scenarios where mixed-trait plots had associational resistance to
279 herbivory, scenarios where mixed-trait plots had associational susceptibility, and scenarios where one trait
280 displayed associational susceptibility and the other associational resistance, depending on the nature of the
281 trait of the plant and whether the compared patches comprised the same number of plants. Hambäck *et al.*
282 (2014) did not model evolutionary changes, nor were SMs modeled beyond 'plant type'- which could be taken
283 to correspond to chemotypes with different effects. However, their model could be modified in the future to
284 include these aspects, and in a multi-chemotype system these kinds of associational effects should certainly
285 be taken into account. Similarly, (Till-Bottraud & Gouyon, 1992) used a quantitative model to study the

286 evolution of the proportion of leaves containing cyanogenic compounds per plant (chemodiversity within
287 individuals) assuming that herbivores leave a patch after tasting the first leaf with cyanogenic compounds.
288 In this way, there were associational effects between leaves both between different plants in the population
289 and between leaves on the same plant.

290 3.5 Coevolution

291 Coevolution is a phenomenon where the evolution of two or more interacting species is influenced by the
292 evolution of the respective other species in the interaction (Ehrlich & Raven, 1964). In the case of the
293 evolution of chemodiversity, chemodiversity can be conceptualized in different ways in the co-evolutionary
294 framework. In some studies, the richness or evenness of SMs is used as a trait of individuals (e.g. Calf *et al.*,
295 2018), in others, the individual SMs are treated as separate traits (Speed *et al.*, 2015). Most coevolutionary
296 models and hypotheses for chemodiversity focus on coevolution with herbivores and envision plants and
297 herbivores to be in an "arms race". There is evidence from field, lab and phylogenetic studies for the
298 coevolutionary arms race hypothesis of chemodiversity on a population, species and lineage level (Becerra
299 *et al.*, 2009; Jander, 2014; Richards *et al.*, 2016; Thorsteinson, 1953).

300 Many quantitative models of **coevolution with herbivores** can be found in the literature (e.g. Ashby &
301 Boots, 2017; Sandoval-Castellanos & Núñez-Farfán, 2023; Gilman *et al.*, 2012; Speed *et al.*, 2015). However,
302 the one that most explicitly addresses chemodiversity is by Speed *et al.* (2015). This model deals with
303 chemodiversity within individual plants and within populations (see Fig. 2). In this individual-based model,
304 the traits that make plants produce toxins have direct resistance counterparts in insect herbivores, so that
305 an insect that feeds on a plant with trait A has to have the corresponding resistance A to the same degree
306 as the plant to be able to reproduce. We have dubbed this style of model a **toxin-resistance matching**
307 **model**. When plants and insects were allowed to evolve toxins and resistances through random mutation
308 and selection (Fig. 3), an arms race developed. Selection favoured plants which produced whatever toxin the
309 insects were least resistant to at a time, while other toxins became less prevalent until the insect resistances
310 evolved in turn and a different toxin would be the toxin they were least resistant to. This caused cycles in
311 which multiple toxins were present in the plants at any time in different concentrations, giving one possible
312 explanation of chemodiversity within individuals, within populations, and in populations through time (Fig.
313 2). Which chemicals dominate in the population at any given time is independent of the chemicals which
314 dominate in separate populations which are going through their own coevolutionary processes, thus causing
315 chemodiversity between populations and within the species.

316 Other quantitative coevolutionary models do not directly address chemodiversity, but contain elements
317 which could be used in chemodiversity models. Gilman *et al.* (2012) developed a model similar to the one by
318 Speed *et al.* (2015) and also show that coevolving with multiple traits can give the 'victim' species an edge,
319 although their model is not explicitly about chemodiversity. Ashby & Boots (2017) developed a host-parasite
320 model which uses a toxin-resistance matching model to model host defenses and parasite resistance traits.
321 This model displayed dynamics of both stable equilibrium where either all traits or none were present, and
322 infinite cyclic evolution. In the case of infinite cycles, there were two types of cycles occurring in the same
323 simulation: fast-cycling of the possible resistances at the same number of resistance loci in a plant and
324 slow cycling of number of resistances that existed in a plant. This was possible through a dynamic where
325 immediate benefits or detriments of the exact resistances an individual possessed caused rapid changes in
326 the resistances present, while subtle fitness costs of maintaining resistances created a slow dwindling of
327 the number of resistances until a threshold was reached where having many was beneficial again. Both
328 types of cycles displayed negative frequency-dependent-selection and are thus linked to the game theory
329 models discussed above. These dual cycles are a feature not found in the model of Speed *et al.* (2015) or
330 in the model by Gilman *et al.* (2012) which is very similar to the model by Ashby & Boots (2017) but
331 only found stable equilibria. While the models of Ashby & Boots (2017) and Gilman *et al.* (2012) are not
332 specific to chemodiversity, these and other matched-trait models of coevolution could easily be translated to
333 a chemodiversity context by making the resistance traits correspond to metabolites.

334 Some authors have proposed that chemodiversity can come about when individual plants produce multiple
335 toxins to make it more difficult for herbivores to adapt to counter every single toxin of the set a plant produces
336 (Speed *et al.*, 2012; Wetzels & Whitehead, 2020). This **slowed adaptation hypothesis**, as coined by Wetzels
337 & Whitehead (2020) is the outcome of an evolutionary process for which toxin-resistance matching is the
338 mathematical description. The slowed adaptation hypothesis may play a role in biological invasions (Box
339 3). However, this is not the only possible verbal explanation for a mechanism of toxin-resistance matching.
340 In the model by Speed *et al.* (2015), the observed chemodiversity is the result of constant, moving target-
341 like innovation and the remains of innovations past, rather than of slowed adaptation of herbivores to a
342 standing set of toxins. Additionally, while all models described in this review that have an explicit or
343 implicit description of individual metabolites use a toxin-resistance model, this is not the only possible way
344 to model interactions between herbivores with metabolites and combinations of metabolites. Quantitative
345 models which explore synergy between metabolites, for example, would require the effects of metabolites to
346 differ depending on which other metabolites are present.

347 Another coevolutionary hypothesis which is based on spatial structure is the **geographic mosaic of**
348 **coevolution hypothesis**. Differences in SM composition between populations may derive from opposite
349 selection pressures of specialists, which use certain SMs for host plant finding, versus generalists that are
350 repelled or deterred by the same SMs (Enge *et al.*, 2012; van der Meijden, 1996). Geographic differences in
351 herbivore abundance may thus result in a selection pressure mosaic (Thompson, 1999; Zangerl & Berenbaum,
352 2003). In a general mathematical model for the geographic mosaic of coevolution that could also be applied
353 to toxin-resistance matching, Gomulkiewicz *et al.* (2000) showed that a spatial setup with coevolutionary
354 hot spots (mutual selection) and cold spots (only one species exerts selection on the other, but not vice
355 versa) could under some conditions allow the maintenance of polymorphism in both species and differences
356 in allele frequency between populations. For a plant toxin coevolving with a herbivore, this would mean
357 that the model could explain coexistence of toxic and nontoxic plant individuals in the same patch as well
358 as differences in the frequency of toxic plants between patches (see Fig. 2).

359 A recent coevolutionary model that does not fit clearly into any of the previously discussed categories
360 investigates the circumstances under which non-linearity of costs and benefits of herbivore resistance can
361 lead to a mix of herbivory-resistant and herbivory-tolerating plants in a population (Sandoval-Castellanos
362 & Núñez-Farfán, 2023). The model is an individual-based model that compares additive and multiplicative
363 versions of a fitness function that includes the fitness costs and benefits of resistance and tolerance as well as
364 the fitness cost of inbreeding depression. It concludes that the nonlinear fitness function needs to be concave,
365 the allocation of costs and benefits multiplicative, selfing non-heritable, and tolerance costly to promote a
366 mix of strategies. The 'resistance' in this model does not necessarily refer to SMs, but the model could be
367 easily modified to include different SMs.

368 To really make the models specific to chemodiversity, introducing ingredients like the branching pathways
369 through which chemicals are created would be useful. Additionally, while toxin-resistance matching has been
370 used in most quantitative coevolutionary models, empirical studies frequently test linear additive or nonlinear
371 synergistic effects of SMs (Dyer *et al.*, 2018; Richards *et al.*, 2016). Therefore, developing models that use
372 these additive or synergistic effects could bring the empirical and quantitative modeling research in this field
373 together.

374 **Coevolution with seed dispersers** is another way in which coevolution can potentially promote
375 chemodiversity. For example, in the genus *Piper*, fruit toxicity affects dispersal by coevolved frugivores.
376 This was statistically modeled by Baldwin *et al.* (2020) based on empirical data collected in a system where
377 *Carollia* fruit bats consume *Piper* fruits. They found that higher levels of defensive amides correlate with

378 shorter gut retention times and lower dispersal distances, while lower concentrations of amides correlate with
379 higher dispersal distances as well as optimal ripeness of the fruit and maximum attractiveness to the fruit
380 bats. In this way, the ripe *Piper* seeds are dispersed as far as possible, while unripe fruits are less likely to be
381 eaten in the first place. Statistical models like this can be used to make quantitative predictions for future
382 empirical studies. In its current form, such a model could explain differences in amide concentration between
383 fruits within an individual or between individuals when fruits differ in ripeness (Fig. 2). For purposes of
384 studying chemodiversity, it would be interesting to not only include total amide concentrations as a predicting
385 variable, but also diversity of amides. It may be possible to shine a light on how coevolution of seed dispersal
386 influences chemodiversity on the within-plant and between-population scale. Moreover, gut retention time
387 in frugivores is not the only way in which chemodiversity may play a role in the coevolution of fruit-bearing
388 plants and their seed dispersers. For example, SMs also may act as olfactory cues to frugivores, inhibit seed
389 germination, and act as toxins (Cipollini & Levey, 1997). So far, we did not find any quantitative models
390 addressing the potentially important consequences of coevolution with seed dispersers for the evolution of
391 plant chemodiversity.

392 **3.6 Screening hypothesis**

393 The models reviewed in the previous sections generally assume that every metabolite has a specific biological
394 activity in the interaction with one or more species. Metabolites without a specific advantageous activity
395 would be expected to be selected away because of their inherent costs. The screening hypothesis first
396 proposed by Jones & Firn (1991) (see also Firn & Jones, 2003; Firn, 2009) challenges this view. Based on
397 the observation that many metabolites produced by plants and fungi have no known function, they argue
398 that pronounced specific biological activity is in fact a rare phenomenon. Thus, in order to defend themselves
399 against a variety of herbivores, plants need to “screen” a large number of “candidate metabolites”. They keep
400 a diversity of metabolites, even many without a function, because this diversity and the underlying multitude
401 of enzymes allows them via mutation to rapidly generate new metabolites and thus increase their chances
402 to find at least some that have strong activity. Since maintaining a diversity of enzymes and metabolites
403 still has costs, the screening hypothesis predicts that the metabolic pathways would be selected such that
404 they can produce a high number of metabolites with as little enzymatic machinery as possible, mostly via
405 promiscuous enzymes that can take multiple substrates, leading to grid-like metabolic pathways. Keeping a
406 diversity of metabolites around can then also allow plants to rapidly respond to new herbivores or herbivores
407 that have evolved a counter-defense (Jones & Firn, 1991).

408 In summary, the screening hypothesis is based on the ingredients of random mutation and natural selec-
409 tion, metabolic pathways, and interactions with other organisms (Fig. 3). Though the focus in the original
410 formulation is on herbivores, by the same logic, a diversity of metabolites can help plants to develop metabo-
411 lites that are beneficial in the interaction with other plants or with mutualists and soil communities. The
412 screening hypothesis has been formulated as a verbal model and to our knowledge, it has not been formulated
413 as a quantitative model. Nor are the ideas of grid-like metabolic pathways, promiscuous enzymes, and rare
414 metabolic activity incorporated into other quantitative models.

415 The screening hypothesis mostly addresses chemodiversity within units of scale, i.e. it attempts to explain
416 why a plant individual, or a plant population, species or clade produces so many different metabolites (Fig.
417 2). Although this has been less discussed by the authors of the screening hypothesis, if a complex metabolic
418 network allows plants to rapidly generate new metabolites, this could also help explain differences between
419 individuals, populations, and species. This appears to be supported by the observation that many enzymes
420 involved in generating chemodiversity of SMs are less conserved than those that are involved in more primary
421 metabolism (Weng *et al.*, 2012).

422 The screening hypothesis has been criticized because evolutionary foresight appears to be required for
423 evolution to create a complex network of pathways just based on the chance that some of its products might
424 eventually prove useful (Pichersky *et al.*, 2006). Proponents of the hypothesis counter that no evolutionary
425 foresight is required if the rapid adaptation made possible through the network is sufficiently beneficial to be
426 selected for (Firn & Jones, 2006). A formal mathematical description or computer model could demonstrate
427 to what extent these criticisms are warranted. Such models have, to our knowledge, not yet been developed,
428 but could therefore play an important role in the debate around the merits of the screening hypothesis.

429 **3.7 Population genetics models**

430 Chemodiversity is, at least in part, underpinned by genetic variation. Thus, it would seem natural to extend
431 classical population genetic models to study chemodiversity.

432 In biodiversity research (Hubbell, 2001) and in population genetics (Kimura, 1983), **neutral models** play
433 an important role as null models for more interesting ecological and evolutionary processes though there is also
434 much debate over how useful they are (see e.g. Clark, 2009; Kern & Hahn, 2018). While neutral biodiversity
435 models assume that species within a guild are equivalent in terms of their birth, death, and migration rates,
436 neutral models in population genetics assume that allele frequencies change just through mutation and drift
437 without selection. We argue that similar neutral models should also be developed for chemodiversity. That

438 is, diversity in such models would come about and be maintained through random mutation and genetic
439 drift without selection (Box 1). A neutral model for chemodiversity should additionally include pathways
440 and enzymes that are encoded by the modeled genes and that thus also evolve neutrally (Fig. 3). A
441 neutral model could in principle be formulated at any scale of organization, though a neutral model derived
442 from neutral models in population genetics would most straightforwardly address chemodiversity within
443 populations, within and between individuals (Fig. 2). This model would serve as a null model for the origin
444 and maintenance of chemodiversity. Any explanations of the origin and maintenance of chemodiversity which
445 propose that there is a selective advantage to producing a diversity of SMs can then be compared to it. We
446 have not found any sources where a model of chemodiversity is compared to such a population genetic null
447 model, but we think that such an approach would be extremely useful. However, patterns of chemical traits
448 between species have been compared to null models for trait evolution on a phylogeny. For example, Volf
449 *et al.* (2018) used Brownian motion and white noise models. So far, however, these models cannot address
450 intraspecific chemodiversity.

451 An explanation for chemodiversity that is slightly more complex than neutrality is **mutation-selection-**
452 **drift balance** (see e.g. Wright, 1937). Mutation-selection-drift models in population genetics generally
453 assume that mutations are either unconditionally beneficial or deleterious. There are no complex selection
454 patterns like balancing selection or negative frequency-dependent selection. Thus variation under this model
455 is observed due to mutations that are transiently segregating in the population before they are getting either
456 lost or fixed. In a chemodiversity extension of such models, new SMs would come about through random
457 mutation in existing metabolic pathways. Here it is assumed that at least some SMs improve the organisms'
458 fitness and are under positive selection, while others may be detrimental or are not worth the cost of their
459 production and are thus under negative selection. The probability that a mutation is fixed or lost and the
460 expected time to do so then depends on the strength of selection relative to genetic drift. During their time
461 to loss or fixation, mutations and their effects on the chemical phenotype would then transiently contribute
462 to chemodiversity. Like neutral models, mutation-selection-drift models could explain chemodiversity within
463 populations, and within and between individuals (Fig. 2), but potentially also at all other scales. For
464 example if different SMs randomly become fixed in different populations and clades this could explain
465 differences between these units. Models based on mutation-selection-drift may thus be highly applicable
466 to chemodiversity.

467 4 Vision

468 It would be easy to fall into the trap of assuming that the different frameworks and hypotheses described
469 here must be in competition, and that through modeling and empirical work, we can find one correct
470 and all others incorrect. Instead, we agree with Dyer *et al.* (2018) that the hypotheses do not have to
471 contradict each other. In fact, many of the hypotheses and selection mechanisms described here may be at
472 play at once within the same plant species. While the members of a particular set of SMs cannot all have
473 both synergistic and toxin-resistance matching fitness effects at the same time, this set of SMs might have
474 synergistic effects on herbivory at the plant level, and also be under negative frequency-dependent selection
475 on the population level. This set can then simultaneously be in a co-evolutionary arms race with herbivores
476 on the lineage level, where new SMs come about within a pathway through the process well-described by the
477 screening hypothesis. Another set of SMs in the same plant species may only exist transiently due to random
478 mutations in a pathway under mutation-selection-drift balance. For this reason, it is useful to think of the
479 different hypotheses and selection methods as different possible factors in the evolution and maintenance of
480 chemodiversity, and not as mutually exclusive. At the same time, some of these hypotheses may well be
481 invalid, or some might contribute less to explaining chemodiversity than others. Quantitative models can
482 serve to investigate the validity and explanatory power of these hypotheses. Combining multiple hypotheses
483 in a single quantitative model would enable us to compare the relative explanatory power of different models
484 (Box 4).

485 When we make a model to test a verbal hypothesis, we need to incorporate enough detail to describe
486 what that hypothesis is about. That means that a test of the coevolutionary arms race needs to incorporate
487 a focal species and an interacting co-evolving species. For the screening hypothesis it is necessary that a
488 model of the metabolic pathways is included that works as described by Firm & Jones (2003); Jones & Firm
489 (1991), but it is not fundamentally necessary that any other species is involved. When modeling a very
490 specific system, it is useful to ensure that all aspects of that system are carefully considered, and all relevant
491 details are included, and all assumptions are justified (Servedio *et al.*, 2014). For example, if one were to
492 model a biological invasion as described in Box 3, it would make sense to include elements of the slowed
493 adaptation and novel weapons/moving target hypotheses (but see Box 4).

494 We have shown here that there is a great body of work on the description of current chemodiversity
495 and its evolutionary history, and there are meaningful hypotheses for the evolution of chemodiversity and
496 maintenance of chemodiversity. For many of the hypotheses, we also found a small number of quantitative
497 modeling studies. Quantitative models that explicitly address chemodiversity where individuals produce a

498 diverse set of metabolites are few and far between. We found such a model for a coevolutionary toxin-
499 resistance matching model where negative frequency-dependence was found Speed *et al.* (2015). The other
500 models did not explicitly consider metabolites but rather a more abstract numerical. These are nevertheless
501 rendered in red in Fig. 1 and Fig. 2, because they can be extended to explicitly include this chemodiversity.
502 So when one looks at Fig. 2, there is work to be done both to extend these models, particularly for those
503 hypotheses which could possibly explain chemodiversity on levels that are not between-individual or within-
504 population. Such models will often be sufficiently complicated that it might not be possible to capture them
505 in a system of equations that can then be solved analytically. Thus, numerical solutions and individual-
506 based models as in Speed *et al.* (2015) appear to be the most productive ways to model the evolution of
507 chemodiversity in these cases. In Box 4 we outline a modular approach to constructing such an individual-
508 based model.

509 **5 Conclusion**

510 In conclusion, while there has been much verbal theory-crafting on chemodiversity and there is a small
511 number of interesting quantitative models for many of the verbal hypotheses, there is much space for the
512 development of mathematical and simulation models. This can be done by expanding existing models in
513 topics related to chemodiversity, adapting existing models of diversity, or by combining different submodels
514 to create coherent models of chemodiversity. In this way, the existing theory can be tested for validity, the
515 explanatory power of different hypotheses can be explored, and the results can be used to interpret empirical
516 data to better understand the dynamics underlying extant chemodiversity.

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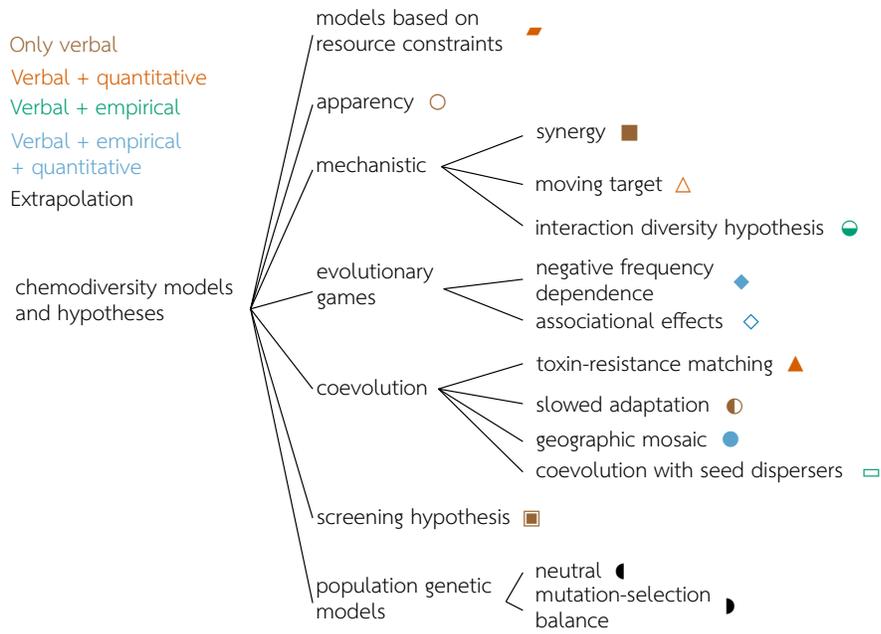


Figure 1

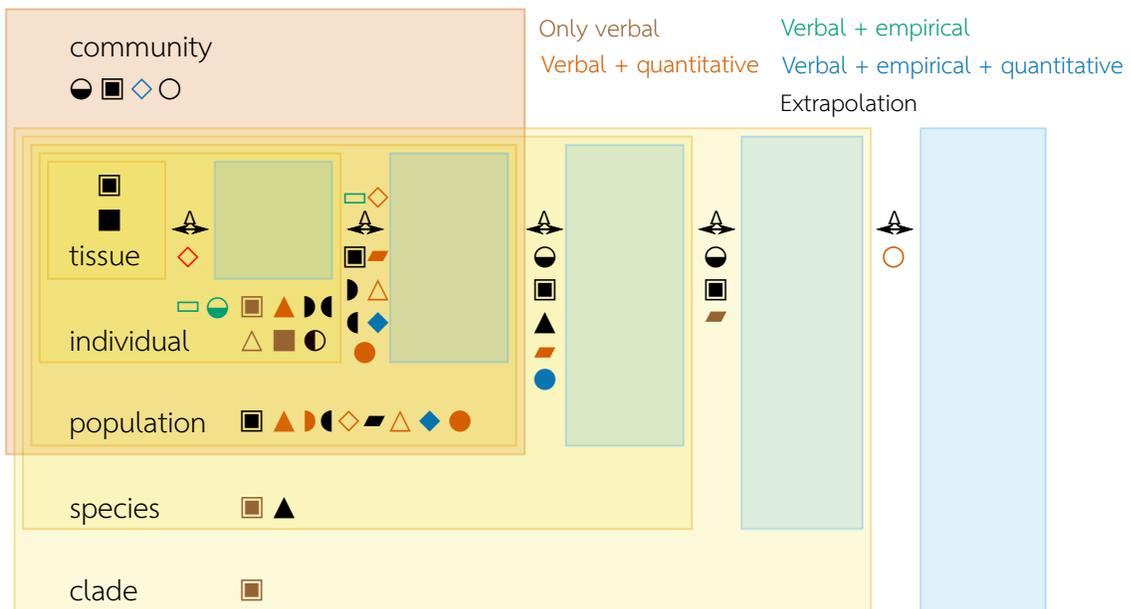


Figure 2

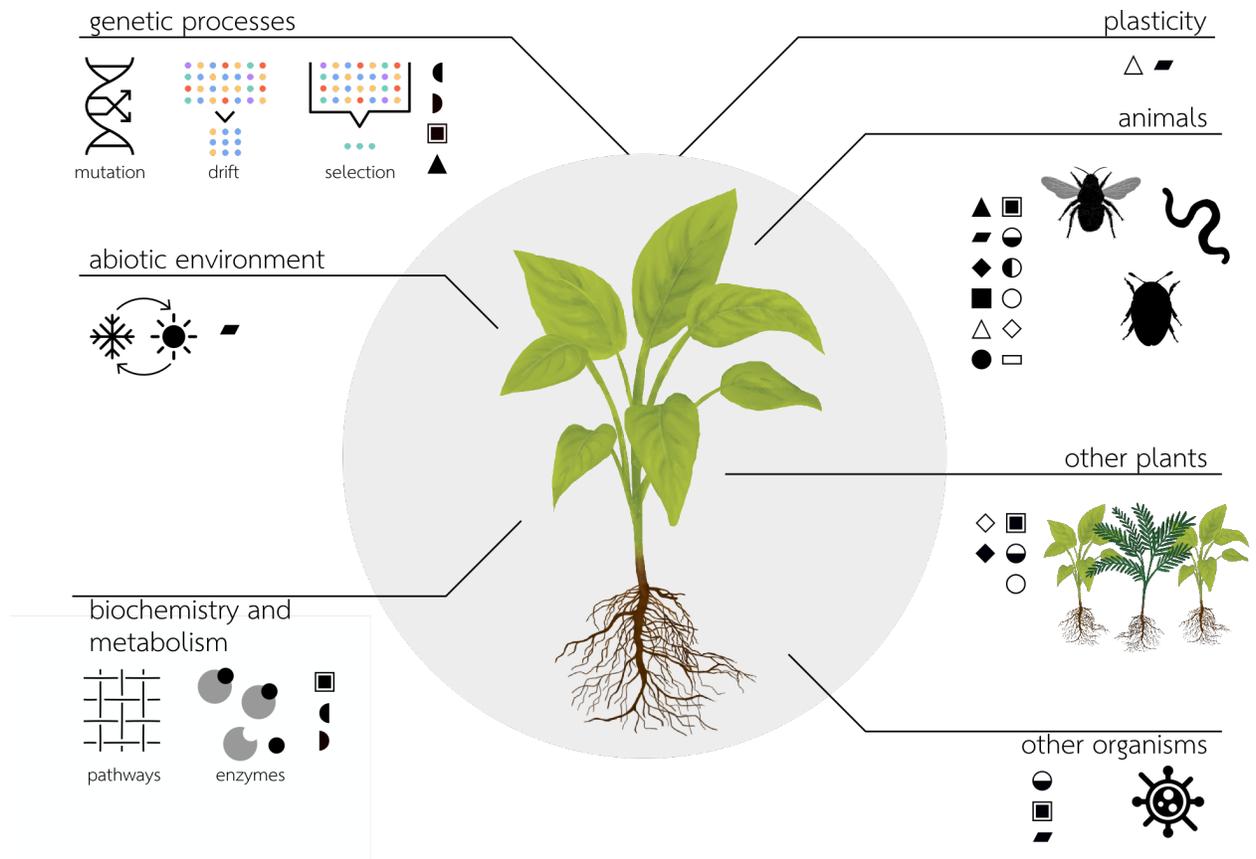


Figure 3

Type of phenotype <input type="radio"/> discrete chemotypes <input type="radio"/> metabolite classes <input checked="" type="radio"/> individual metabolites <i>skip the section 'Chemical model'</i>	
Link between genotype and phenotype <input checked="" type="radio"/> phenotype is identical to genotype <input type="radio"/> genotype distinct from phenotype <input checked="" type="checkbox"/> <i>skip the section 'Chemical model'</i> <input type="checkbox"/> gene regulation <input type="checkbox"/> dominance <input type="checkbox"/> stochasticity <input type="checkbox"/> plasticity	
Chemical model <input type="radio"/> no chemical model <input type="radio"/> enzyme kinetics, but no pathways <input checked="" type="radio"/> specific pathways <input checked="" type="checkbox"/> <input type="checkbox"/> abstract pathways <input checked="" type="checkbox"/>	
Inheritance of traits <input checked="" type="radio"/> no genetic mutations <input type="radio"/> mutations at reproduction <input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> deletion/duplication <input type="checkbox"/> gene function mutations <input checked="" type="checkbox"/> <input checked="" type="checkbox"/> Reproductive system <input type="radio"/> asexual <input checked="" type="radio"/> sexual <input type="radio"/> diploid <i>skip to next section</i> Selection of parents for offspring <input type="radio"/> random <input checked="" type="radio"/> fitness based <input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <i>skip section on phenotype and fitness</i> <input type="radio"/> deterministic proportional <input checked="" type="radio"/> stochastic lottery <input type="checkbox"/> include genetic linkage	
Abiotic environment <input checked="" type="checkbox"/> <input type="checkbox"/> light <input type="checkbox"/> water <input type="checkbox"/> nutrients <input type="checkbox"/> temperature <input type="checkbox"/> elevation	
Animal interactions <input type="radio"/> no interactions <input checked="" type="radio"/> interactions <input type="checkbox"/> seed dispersers <input type="checkbox"/> <input type="checkbox"/> pollinators <input checked="" type="checkbox"/> herbivores <input checked="" type="checkbox"/> intraspecific trait variation <input type="checkbox"/> mobility <input type="checkbox"/> coevolution <input type="checkbox"/> <input type="checkbox"/> multiple species <input checked="" type="checkbox"/>	Scale requiring this selection: 1 = within-tissue 2 = between-tissue 3 = within-plant 4 = between-plant 5 = within-population 6 = between-population 7 = within-community 8 = between-community
<input type="checkbox"/> pathogens <input type="checkbox"/> non-animal mutualists	
Spatial model <input type="radio"/> no spatial model <input checked="" type="radio"/> spatial model <input type="checkbox"/> raster <input checked="" type="checkbox"/> <input checked="" type="checkbox"/> patches/islands _{6, 8} <input type="checkbox"/> plant parts _{1, 2, 3} <input type="checkbox"/> physical dynamics of volatile compounds <input type="radio"/> <input checked="" type="checkbox"/>	
Relation between phenotype and fitness <input type="radio"/> synergic <input checked="" type="checkbox"/> <input type="radio"/> additive <input type="radio"/> threshold <input type="radio"/> toxin-resistance <input checked="" type="checkbox"/> <input checked="" type="checkbox"/> <input type="checkbox"/> multiple plant species _{7, 8}	

716

717

Figure 4

Box 1: Ingredients of chemodiversity models

Here we explain the biological processes that act as key ingredients (Fig. 3) in current models and hypotheses for chemodiversity and, without aspiring to completeness, give some exemplary empirical studies supporting their role in shaping chemodiversity. How the ingredients are used in the various theoretical models is explained in section 3 and Box 4.

- **Genetic processes:** SMs are produced by enzymes, which are coded for in genes, which in turn have regulatory genes. Therefore, genetic processes can give rise to chemodiversity. Genes for enzymes in the same metabolic pathway are often organized as operons, i.e. located closely together in the genome (Hamberger & Bak, 2013). Within these operons, gene duplication and subsequent divergence is common, which facilitates a flexible gene network that can undergo rapid evolution (Hamberger & Bak, 2013).
- **Mutations** may change the function of the enzyme coded for by a gene, so it has a different substrate profile or catalyzes different reactions and therefore produces different SMs than its ancestor. Meanwhile changes in gene regulation may change the abundance of SMs, and duplication and deletion may open new pathways of SM synthesis or close them off. Random mutations thus bring about genetic and consequently chemical diversity.
- **Genetic drift** is the phenomenon of changing allele frequencies through chance effects on reproduction and survival rather than natural selection based on fitness. For example, a gene might become fixed or lost randomly within one population but not another, resulting in chemodiversity between populations.
- **Natural selection** occurs when SMs which confer net fitness benefits become more common, and SMs that confer net fitness costs become more rare. Similarly, if there is a fitness benefit to lack of specificity in the production of SMs, this trait may be selected for.
- **The abiotic environment**, such as water and nutrient availability, temperature, or elevation, can determine how much a plant can invest in growth and defense (Bryant *et al.*, 1983; Coley *et al.*, 1985; Herms & Mattson, 1992; Hamilton *et al.*, 2008; Monson *et al.*, 2021; Smakowska *et al.*, 2016). Moreover, changes in the abiotic environment may cause stress responses that lead to changes in the concentrations of certain SMs or induce biosynthesis of new SMs (Agati & Tattini, 2010).
- **Biochemistry and metabolism:** Metabolic pathways for the production of SMs can involve multiple enzyme-catalyzed reactions and can be branched. Which new SMs can be produced by changes in these

747 pathways is constrained by the extant pathways. Many of the enzymes involved in SM synthesis are
748 “promiscuous”, meaning that they accept multiple substrates and thus can produce multiple different
749 SMs at low cost (Aharoni *et al.*, 2005). Thus, small modifications to the enzyme suite can have a
750 profound impact on the resulting metabolite composition (Moghe & Last, 2015; Shoji, 2019).

- 751 • **Phenotypic plasticity:** Phenotypic plasticity can explain chemodiversity even among genetically
752 identical individuals in a population. For example, an individual in a nutrient-poor micro-environment
753 may invest these resources differently from an individual in a nutrient-rich micro-environment, leading
754 to variation in chemical composition between individuals in a population (Defosse *et al.*, 2021; Stamp,
755 2003).
- 756 • **Interactions with animals** play a role in the majority of hypotheses and models for the evolution of
757 chemodiversity and their importance has broad empirical support (Calf *et al.*, 2018; Hambäck *et al.*,
758 2014; Li *et al.*, 2020; Whitehead *et al.*, 2021). For example, in the bittersweet nightshade *Solanum*
759 *dulcamara*, slugs showed a preference for leaves from populations which produced fewer glycoalkaloids
760 (Calf *et al.*, 2018). These plants belonged to populations in which few slugs were present, hinting at
761 local adaptation of plant populations. SMs can also serve as visual, olfactory or gustatory signals to
762 attract pollinators and seed dispersers, and influence seed dispersion by influencing gut retention time
763 (Baldwin *et al.*, 2020; Borghi *et al.*, 2021; Cipollini & Levey, 1997; Nevo *et al.*, 2018).
- 764 • **(Indirect) interactions with other plants:** Plants can interact directly with each other, for exam-
765 ple through SMs that inhibit the germination or growth of heterospecific competitors (i.e allelopathy).
766 In the black mustard *Brassica nigra* for example, the production of sinigrin was found to be advanta-
767 geous when among heterospecifics but not when among conspecifics (Lankau & Strauss, 2007, 2008).
768 Interactions with other plants can also be indirect, often through interactions with other species in the
769 environment such as herbivores (associational effects, Hambäck *et al.*, 2014; Salazar & Marquis, 2022).
- 770 • **Interactions with other organisms:** Apart from animal interaction partners, numerous microbial
771 mutualists are guided by plant SMs to their host, and the formation of symbiotic structures is induced
772 by SMs (De la Peña & Loyola-Vargas, 2014), for example in interaction with rhizobia (Cooper, 2004)
773 and arbuscular mycorrhizal fungi (AMF) (Akiyama *et al.*, 2005). Differences in root exudates among
774 populations within species have been found to lead to distinct soil chemical communities (Mueller
775 *et al.*, 2020). Mutualists can in turn also affect patterns of chemodiversity. For example, endophytes

776 and a symbiosis with AMF resulting in functional arbuscular mycorrhiza can modify the chemical
777 composition of different plant tissues or organs (Schweiger & Müller, 2015; Yadav *et al.*, 2022).

778 **Box 2: Scales of chemodiversity**

779 Chemodiversity can be quantified at various scales of organization (Wetzel & Whitehead, 2020). Terms
780 such as richness and evenness, α , β and γ diversity that are familiar from species diversity are frequently
781 used. However, different authors use different definitions for these terms (see e.g. discussion in Kessler &
782 Kalske, 2018; Li *et al.*, 2020; Wetzel & Whitehead, 2020), depending on the scales their work focuses on. We
783 argue that using the same terms to speak about different scales can be confusing, especially when comparing
784 studies, and when there are more than two scales that may be of interest. Therefore, rather than redefining
785 the terms α , β and γ depending on the scales a particular study or model works with, we name these scales
786 explicitly.

787 We argue that instead of using α , β , and γ diversity, it is more helpful to explicitly state the scales under
788 consideration (tissue, individual, population, species, clade, or community) and additionally specify whether
789 diversity within units at those scales or differences between units at those scales are considered (Fig. 2).
790 For example, one can quantify chemodiversity within individuals as the average number of SMs produced
791 per individual, and between-population chemodiversity as number of SMs that are produced only by one
792 of the populations. Note that between-unit chemodiversity is not the same as within-unit chemodiversity
793 at the next higher scale. For example, the average number of metabolites that are not shared between
794 randomly picked individuals in the population is not the same as the total number of metabolites in the
795 whole population. At each scale, chemodiversity could be quantified using different metrics, e.g. metrics
796 focusing on richness, evenness, or disparity (Petrén *et al.*, 2023). In Fig. 2, we place all hypotheses at the
797 scale or scales at which they have been shown to explain, claim to explain, or can be logically inferred to
798 explain chemodiversity.

799 **Box 3: Biological invasions as testing ground for chemodiversity** 800 **models**

801 Connecting models for the evolution of chemodiversity to empirical data can be challenging because in natural
802 populations we can usually only observe a snapshot of the evolutionary process at the current time, but the
803 long evolutionary history remains hidden. Biological invasions of plants or interacting animals offer exciting
804 opportunities for testing these hypotheses because here the interaction partners often experience sudden large
805 changes in selection regime, with different locations providing 'replicate experiments', sometimes resulting in
806 rapid evolution on ecological time scales (Cox, 2004). Moreover, understanding the mechanisms facilitating
807 invasions may be important to develop conservation strategies.

808 Invasive species may particularly benefit from high intraspecific variation in chemical composition, i.e.
809 chemodiversity on a population level. For example, tansy *Tanacetum vulgare* (Asteraceae) individuals are
810 highly diverse in their terpenoid profiles, with different terpenoids occurring in different concentrations (Wolf
811 *et al.*, 2011). Terpenoids have various ecological functions (Cheng *et al.*, 2007), and effects on antagonists are
812 highly species-specific. Therefore, a high chemodiversity within invasive plant populations may impede the
813 adaptation of herbivores and microorganisms native to the habitat (Tewes *et al.*, 2018; Wolf *et al.*, 2011), as
814 predicted by the slowed adaptation hypothesis. Furthermore, the simple unfamiliarity of native antagonists
815 with the novel SMs is thought to provide a competitive advantage to *T. vulgare*. This is called the 'novel
816 weapons hypothesis' (Callaway & Ridenour, 2004) in the context of biological invasions and similar to the
817 moving target hypotheses described above in that both confront antagonists with random and unfamiliar
818 SMs.

819 Furthermore, multiple introductions from different source populations to areas where the plant became
820 invasive may have led to hybridisation of individuals of different chemotypes, producing novel mixtures of
821 terpenoid profiles and thus novel chemotypes. Indeed, a few chemotypes were only found in the invasive
822 range of *T. vulgare* (Wolf *et al.*, 2011). Moreover, potential inbreeding in invasive populations and envi-
823 ronmental conditions different from the originating environment may lead to rapid evolutionary changes
824 (Schrieber & Lachmuth, 2016), which may result in novel SM variation. Overall, changes in chemodiversity
825 and the functions in species interactions in non-native populations have been largely neglected (Tewes *et al.*,
826 2018), although chemical composition can highly vary between native and non-native populations (Pankoke
827 *et al.*, 2019; Tewes *et al.*, 2018) and due to hybridization (Piola *et al.*, 2013), offering great opportunities to
828 develop theoretical models and test them with empirical studies. The novel-weapons hypothesis has already

829 been mathematically modeled (Allstadt *et al.*, 2012), albeit not as a chemodiversity model, and is ripe for
830 expanding upon (see Box 4).

831 **Box 4: Constructing a generalized chemodiversity individual-based** 832 **model**

833 When developing an individual-based model to investigate chemodiversity, one needs to carefully consider
834 what to include in the model, based on what one wants to investigate with it. Different questions require
835 different models with different ingredients. However, the parts that different models are built from can be
836 re-used across models. Considering different mechanisms that could describe how a particular ingredient
837 (Box 1) influences chemodiversity also means that you can construct your model in a modular way so that
838 you can compare these mechanisms to each other and see which model explains empirical results best.

839 This does not mean that every individual-based model of chemodiversity has to be able to test all possible
840 mechanisms involved with the evolution and maintenance of chemodiversity. A model should include no
841 more complexity than it requires (Servedio *et al.*, 2014), and there is also the risk of overfitting in overly
842 complex models. Nevertheless, one should be mindful of alternative explanations when making a model that
843 investigates a particular set of hypotheses. A model can show a verbal hypothesis to be flawed if the stated
844 logic cannot be mathematically reproduced, and a model that can test multiple proposed mechanisms can
845 give clues to which of these mechanisms may have the strongest impact on the origin and maintenance of
846 chemodiversity (Otto & Rosales, 2019).

847 We propose the workflow described by Fig. 4 and the more elaborate Supplement 1. This is a workflow
848 for creating individual-based models of chemodiversity based on individual plants, and is most useful for
849 models that describe the origin and maintenance of chemodiversity up to the multi-species community or
850 multiple-population level.

851 For the example from Box 3, a possible set of selections to investigate chemodiversity in biological invasion
852 is marked in Fig. 4. Here, the model of Allstadt *et al.* (2012), which is already an IBM with two types
853 of plants (resident and invader) competing in a raster, is expanded by selecting several options that are
854 not considered in the original model. As empirical studies tend to focus on variation within a metabolite
855 class (here for an example in invasion biology, see Wolf *et al.* (2011)), chemotype in this proposed model is
856 modeled through individual metabolites. The model has sexual reproduction without mutations but with

857 genetic linkage to focus on the population genetics of biological invasions, and interactions with herbivores
858 are modeled explicitly in this version. Lastly, the origin of the invading plants is modeled as an abstract
859 island that individuals with the invading genotype might arrive from.

860 **Caption Figure 1:** Tree showing a rough conceptual grouping of chemodiversity models and hypothe-
861 ses. Each model or hypothesis has a unique symbol, which is then used in Fig. 3 to indicate which ingredients
862 are part of the model, in Fig. 2 to indicate at what scales the model can explain chemodiversity, and in Fig.
863 4 to indicate which model components are essential to any model that investigates these hypotheses. Note
864 that similarity in shape between symbols here does not indicate any relationship between models. Models
865 with brown symbols have only a verbal model, green symbols a verbal model and empirical evidence, red a
866 quantitative model, and blue both a quantitative model and empirical evidence. Black symbols stand for
867 models that have, to our knowledge, not been previously used in the context of chemodiversity but that we
868 argue can be extended to modeling chemodiversity (i.e. they are our extrapolation).

869 **Caption figure 2:** Schematic overview of the scales at which the different models and hypotheses
870 (potentially) explain the evolution of chemodiversity. Please refer to Fig. 1 for the models each symbol refers
871 to. Models with brown symbols have only a verbal model, green symbols additionally have empirical evidence,
872 red a quantitative model, and blue both a quantitative model and empirical evidence. Black symbols stand
873 for models that have, to our knowledge, not been previously used in the context of chemodiversity but that we
874 argue can be extended to modeling chemodiversity (i.e. they are our extrapolation). For example, the filled
875 blue diamond next to the label “population” indicates that there is a quantitative model and also empirical
876 evidence suggesting that negative frequency-dependence can explain the maintenance of chemodiversity
877 within a population and the filled red circle between the two “individual” boxes indicates that a quantitative
878 model based around a geographic mosaic exists that suggests that it can maintain differences between
879 individuals. For some symbols, we used different colors in different places in the figure to indicate a distinct
880 type of support for chemodiversity at the various scales. The community level is orthogonal to the species
881 level because populations can be considered nested both in their species and in a multi-species community.

882 **Caption figure 3:** Overview of the key ingredients of chemodiversity models. To see which hypothe-
883 sis/model a symbol stands for, please refer to Fig. 1. The ingredients listed in the figure are further explained
884 in Box 1. Models and hypotheses are listed as connected to particular ingredients when these ingredients
885 are key components of the model as described below in section 3 and Box 4.

886 **Caption figure 4:** Flowchart of the decision making process for making a individual-based model to
887 investigate chemodiversity. Every section represents one important 'module' in the model, with one or more
888 elements to pick decisions on. Top-level decisions are aligned left, while indentation indicates that a decision
889 should only be considered if the option above was picked. Circles represent mutually exclusive decisions, with
890 the possible options presented in a horizontal row. Squares represent decisions where multiple options can
891 be picked, and are presented in vertical columns. The yellow ovals are a set of possible options for an IBM
892 inspired by (Allstadt *et al.*, 2012) which could investigate the development of chemodiversity under biological
893 invasion (Box 3). Symbols behind options or section names indicate that this option or any option from this
894 section is necessary for a model that aims to investigate the hypothesis corresponding to this symbol (Fig.
895 1), while numbers indicate this option is necessary to investigate chemodiversity on the corresponding scale
896 (Fig. 2). An interactive R program with a more detailed decision tree can be found in SI. 1.