# Improving Targeted Small Molecule Drugs to Overcome Chemotherapy Resistance

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#### Abstract

Conventional cancer treatments face the challenge of therapeutic resistance, which causes poor treatment outcomes. The use of combination therapies can improve treatment results in patients and is one of the solutions to overcome this challenge. Chemotherapy is one of the conventional treatments that, due to the non-targeted and lack of specificity in targeting cancer cells, can cause serious complications in the short and long-term for patients by damaging healthy cells. Also, the employment of a wide range of strategies for chemotherapy resistance by cancer cells, metastasis, and cancer recurrence create serious problems to achieve the desired results of chemotherapy. Accordingly, targeted therapies can be used as a combination treatment with chemotherapy to both cause less damage to healthy cells, which as a result, they reduce the side effects of chemotherapy, and by targeting the factors that cause therapeutic challenges, can improve the results of chemotherapy in patients. Small molecules are one of the main targeted therapies that can be used for diverse targets in cancer treatment due to their penetration ability and characteristics. However, small molecules in cancer treatment are facing obstacles that a better understanding of cancer biology, as well as the mechanisms and factors involved in chemotherapy resistance, can lead to the improvement of this type of major targeted therapy. In this review article, at first, the challenges that lead to not achieving the desired results in chemotherapy and how cancer cells can be resistant to chemotherapy are examined, and at the end, research areas are suggested that more focusing on them, can lead to the improvement of the results of using targeted small molecules as an adjunctive treatment for chemotherapy in the conditions of chemotherapy resistance and metastasis of cancer cells.

**Title:** Improving Targeted Small Molecule Drugs to Overcome Chemotherapy Resistance

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## Abstract:

Conventional cancer treatments face the challenge of therapeutic resistance, which causes poor treatment outcomes. The use of combination therapies can improve treatment results in patients and is one of the solutions to overcome this challenge. Chemotherapy is one of the conventional treatments that, due to the non-targeted and lack of specificity in targeting cancer cells, can cause serious complications in the short and long-term for patients by damaging healthy cells. Also, the employment of a wide range of strategies for chemotherapy resistance by cancer cells, metastasis, and cancer recurrence create serious problems to achieve the desired results of chemotherapy. Accordingly, targeted therapies can be used as a combination treatment with chemotherapy to both cause less damage to healthy cells, which as a result, they reduce the side effects of chemotherapy in patients. Small molecules are one of the main targeted therapies that can be used for diverse targets in cancer treatment due to their penetration ability and characteristics. However, small molecules in cancer treatment are facing obstacles that a better understanding of cancer biology, as well as the mechanisms and factors involved in chemotherapy resistance, can lead to the improvement of this type of major targeted therapy. In this review article, at first, the challenges that lead to not achieving the desired results in chemotherapy and how cancer cells can be resistant to chemotherapy are examined, and at the end, research areas are suggested that more focusing on them, can lead to the improvement of the results of using targeted small molecules as an adjunctive treatment for chemotherapy in the conditions of chemotherapy resistance and metastasis of cancer cells.

## Keywords:

Chemotherapy Resistance, Small Molecule, Tumor Microenvironment, Immunomodulation, DNA Repair Mechanisms, Cancer Stem Cell

## Introduction:

Cancer is a heterogeneous and highly complex disease that annually claims the lives of millions of people, making it one of the leading causes of death worldwide [1]. The number of deaths caused by cancer is growing increasingly and predictions indicate that it will continue to do so. Estimates show that in 2018, 9.6 million cancer-related deaths occurred [2]. However, by 2030, it is projected that around 30 million people may die annually from cancer [2]. That is why cancer is a very serious challenge for public health worldwide, and it becomes increasingly widespread and severe every year. Although there are innovative and diverse methods for cancer treatment, four treatment methods are conventional and usually employed to combat cancer, including surgery, radiation therapy, immunotherapy, and chemotherapy. Chemotherapy, however, faces many challenges such as non-specific targeting of cancer cells, treatment resistance, and cancer recurrence even after successful treatment. The use of targeted small-molecule drugs is one of the ways to improve the outcome of chemotherapy. Because of their low molecular weight (<1000 Da) and small size, they can bind to various targets outside and inside the cell [3]. Since 2001, when the first small molecule tyrosine kinase inhibitor (TKI) drug, imatinib, by the US Food and Drug Administration (FDA), was approved for clinical use, more than 80 small molecule drugs for cancer treatment have been approved by the US FDA and the National Medical Products Administration (NMPA) of China [4]. However, the therapeutic results of targeted small molecules are limited. Cancer cells use a variety of factors and strategies to fight and resist chemotherapy. For this reason, to improve the results of treatments based on small molecules as adjunctive therapy to overcome the challenges of chemotherapy, knowledge and deep understanding of chemotherapy resistance mechanisms and then determining the most effective and main factors as targets for small molecules is very crucial and important. In this study, most of these factors and mechanisms are comprehensively examined in four titles including tumor microenvironment, immunomodulation, DNA repair mechanisms, and cancer stem cells. It is crucial and essential for drug designers and scientists to extensively investigate these mechanisms for the proper and effective design of targeted small molecule drugs as adjunctive chemotherapy treatments.

## 1. Tumor microenvironment (TME)

The tumor microenvironment (TME) provides a secure environment for cancer cells to evade the desired outcomes of various treatments. Essentially, the TME is a complex and dynamic ecosystem composed of diverse factors that play crucial roles in inhibiting apoptosis, proliferation, migration, immune evasion, treatment resistance, metastasis, metabolic reprogramming, and all stages of tumorigenesis [5]. The tumor microenvironment factors are generally divided into two main components: cellular components (such as tumor-associated macrophages (TAMs), tumor-infiltrating lymphocytes (TILs), myeloid-derived suppressor cells (MDSCs), cancer-associated fibroblasts (CAFs), and endothelial cells (ECs) which all of them are stromal cells), and non-cellular components (such as growth factors, various chemokines and cytokines, interstitial fluids, metabolites, extracellular matrix (ECM) and exosomes) [5, 6]. Therefore, targeting the TME is a potentially effective strategy for achieving fruitful outcomes of cancer therapy, and small molecules can easily penetrate the TME and ultimately reach tumor cells and affect them [6]. The tumor microenvironment is a hypoxic and low-pH environment [6]. The rapid growth of tumor cells causes hypoxia, which

subsequently causes the release of stimulating factors such as vascular endothelial growth factor (VEGF), matrix metalloproteinases (MMPs), and hypoxia-inducible factor-1 $\alpha$  (HIF- 1 $\alpha$ ) [5]. Also, the condition of hypoxia in TME affects the endothelial cells by secreting angiogenic factors from the tumor and subsequently promoting angiogenesis [7, 8]. In the TME, the most abundant stromal cells are cancer-associated fibroblasts (CAFs) that can produce abnormal extracellular matrix (ECM) which supports tumor cell migration [9]. TME reshaping creates the conditions for tumor cells to interact with surrounding fibroblasts, immune cells, and endothelial cells, leading to the induction of a variety of biological events, including angiogenesis, migration, proliferation, immune system suppression, and drug resistance, which ultimately causes tumor growth [9-18].

One of the important events in the TME is cell interaction and cell communication with the ECM because this interaction causes the release of factors that play a role in ECM remodeling and immune evasion, which ultimately strengthens treatment resistance [19]. Other important events are the generation of exosomes by benign and malignant cells, TME-specific metabolic patterns, and circulating deregulated microRNAs that increase treatment resistance [19]. There are many different types of immune cells in the TME that block the immune response, in addition, around the tumor cells, there are a set of inflammatory molecules that cause the failure to identify and eliminate cancer cells by the immune system, which together make TME a complex and heterogeneous space and they often cause an uncontrollable process in the growth and development of tumors [5, 20-24]. On the whole, a wide range of events and factors from biochemical agents, and a hypoxic environment to abnormal mechanical forces cause treatment resistance [19]. In the following, circumstances and various components of the tumor microenvironment are discussed:

# 1.1. Extracellular vesicles (EVs)

Extracellular vesicles (EVs) act as intermediaries in intercellular communication and are secreted by various types of cells [25]. These vesicles are composed of a lipid bilayer that protects their contents from enzymatic degradation [26]. They carry diverse biological active molecules such as lipids, proteins, and various nucleic acids including miRNA and lncRNA, and can regulate cellular processes and functions, leading to changes in gene expression and activation of multiple signaling pathways [25, 27-29]. Tumor-derived EVs can modulate the tumor microenvironment [30]. EVs by transferring surface markers and signaling molecules, nucleic acids, and oncogenic proteins to stromal cells, can prepare the tumor microenvironment for tumor growth, invasion, and metastasis [31-33]. Stromal and tumor cells, by releasing EVs, cause heterogeneity and complexity in the tumor microenvironment [25]. EVs create favorable conditions in the tumor microenvironment for tumor growth and resistance to anti-cancer drugs by transferring bioactive materials [25]. EVs contribute to drug resistance through various mechanisms, including drug export and sequestration, reduction of drug concentration in target sites, transferring drug efflux pumps, mediating communication between cancer and stromal cells, transfer of survival factors, apoptosis inhibitors, and non-coding RNAs [25]. EVs not only play a role in metastasis and drug resistance but also in immune suppression and angiogenesis, and also, by providing growth factors such as TGF- $\beta$  (transforming growth factor- $\beta$ ) and various miRNAs, they can convert mesenchymal stem cells and other bone marrow-derived cells into tumor-supporting cells [25, 34, 35].

One class of EVs is exosomes, which are involved in various cancer events such as apoptosis escape, immune suppression/evasion, cell proliferation, inflammatory responses, angiogenesis, invasion, metastasis, and chemotherapeutic sensitivity [5, 36-39]. Not only are exosomes involved in acquired drug resistance through various cellular and molecular processes in the tumor microenvironment (including DNA repair, epithelial-mesenchymal transition (EMT), immune surveillance, and cell cycle), but also they contribute to drug resistance through various pathways, including transporting drug efflux pumps, direct drug export, and miRNA signaling [40-42]. By transferring ABC transporters (drug efflux pumps) through exosomes, drug resistance is promoted in sensitive cells [36].

# 1.1.1. Exosomal miRNAs

miRNAs, or microRNAs, are a type of short ncRNAs that regulate various important biological functions

such as apoptosis, migration, proliferation, differentiation, drug resistance, and invasion by regulating gene expression [5, 43]. Cancer cells create an abnormal expression of miRNAs through genetic or epigenetic changes, which subsequently leads to abnormal expression of their target genes [44-49]. miRNAs act as elements that promote the formation and biological changes of TME [50-53]. Exosomal miRNAs derived from tumors cause heterogeneity and phenotypic changes in TME and subsequently promote uncontrolled tumor growth [20, 36, 54-58]. Exosomal miRNAs derived from tumors by matrix reprogramming in TME create a context for resistance to chemotherapy, tumor growth, immune system escape, and metastasis [5]. Exosomal miRNAs secreted by cancer stem cells that target immunosuppressive and anti-apoptotic pathways can impart and develop drug resistance in susceptible neighboring cells [36]. Exosomal miRNAs derived from cancer stem cells (CSCs) cause inhibition of pro-apoptotic FOXO3a property and activation of mTOR signaling pathway in sensitive cancer cells and they can inhibit apoptosis and subsequently promote tumor progression, therefore, drug-induced apoptosis is inhibited [36]. Horizontal transfer of exosomal miRNAs derived from cancer cells can induce a resistant phenotype of drug-resistant cells in sensitive cancer cells and create resistance to a wide range of anticancer drugs, moreover, miRNAs can be delivered from cancer cells to TME cells by exosomes and modulate the process of drug resistance response in TME [59-61]. Exosomal miRNAs derived from cancer stem cells and non-cancerous cells help to drug resistance by creating different effects on target cells in TME, in addition, exosomal miRNAs play a role in inducing resistance to specific molecular target drugs and cytotoxic drugs [36]. Due to the key role of miRNAs in cancer and their regulation of drug resistance in a tumor-specific manner by some miRNAs, exosomal miRNAs can be considered and used as potential cancer biomarkers for prediction and diagnosis in a broad or specific tumor approach [36]. Exosomal miRNAs derived from cancer cells and transferred to fibroblasts in the tumor microenvironment (TME) promote differentiation of cancer-associated fibroblasts (CAFs), subsequently, exosomal miRNAs secreted by CAFs induce drug resistance in cancer cells through induction of proliferation, metastasis, and inhibition of anti-tumor effects of cytotoxic drugs such as cell cycle arrest and apoptosis [36]. For example, the transfer of exosomal miR-21 derived from CAFs to ovarian cancer cells led to inhibition of apoptosis and downregulation in the expression of apoptotic peptidase activating factor 1 (APAF1), as a result, resistance to treatment with paclitaxel was increased [62]. Alterations in the Extracellular matrix promote the widespread growth of cancer cells, angiogenesis, metabolic reprogramming, and inflammatory response [5]. Primary tumor cells release exosomal miRNAs such as miR-21, miR-155, miR-210, miR-1247-30, and miR-124 that are transferred to normal fibroblasts (NFs), then, by targeting proteins such as SPHK1, PTEN, and SOCS1, as well as activating molecules such as FGF-2, FAP, TGF- $\beta$ , and bFGF induce NFs conversion into CAFs, ultimately, ECM undergoes reshaping [5, 36].

# 1.1.1.1. Exosomal miRNAs and tumor-associated macrophages (TAMs)

The most abundant immune cell population in the TME is tumor-associated macrophages (TAMs) [36]. TAMs are highly plastic cells that are involved in various actions, including suppressing the immune system, promoting tumor angiogenesis, and increasing resistance of tumor cells to chemotherapy [63, 64]. Poor prognosis in many cancer types is directly related to the number of TAMs in the tumor microenvironment [65]. According to several studies, in several cancer types, including lung, skin, colon, head and neck, bladder, and ovarian cancer, exosomal miRNAs secreted by cancer cells can induce the transition of macrophages to the TAMs [66-71].

## 1.1.1.2. Exosomal miRNAs and Epithelial-mesenchymal transition (EMT):

Epithelial-mesenchymal transition (EMT) is a process in which cancer cells acquire increased motility and tremendous plasticity, caused by loss of cell junctions and apical-basal polarity and acquisition of mesenchymal characteristics [72, 73]. EMT cells are directly related to metastasis, cancer progression, and drug resistance in various cancers because they have an extremely invasive phenotype [74]. There is ample evidence to demonstrate that the EMT process in the TME can be modulated by exosomes released from cancer cells [41, 75, 76]. Xiao et al. showed that exosomal miRNAs involved in regulating EMT, such as miR-191 and let-7a derived from melanoma, induced EMT in primary melanocytes [77]. Additionally, studies have shown that exosomal miRNAs are involved in activating or stabilizing EMT in primary tumor cells [36].

## 1.1.1.3. Exosomal miRNAs and autophagy

Autophagy is a catabolic process that removes damaged or redundant macromolecules and organelles to maintain homeostasis and metabolic adequacy [78]. This process is involved in escalating tumor resistance against therapy and increasing tumor growth, as well as upregulating autophagy frequently occurring leading to cell survival and high energy supply during cancer initiation [36, 79, 80]. During a phenomenon called cytoprotective autophagy a process caused by high levels of autophagy and the creation of hypoxic TME, resulting in oxidative stress, ultimately delays apoptosis and subsequently contributes to treatment resistance [81, 82]. Upregulation of exosomes and autophagy can act to induce an adaptive response under cellular stress conditions in cancer cells [36]. Exosomes released from cancer cells can induce the construction of reactive oxygen species (ROS) and stimulate the secretion of tumor growth factors by upregulation of autophagy in target cells [83]. Moreover, during a drug treatment regimen, exosomes derived from cancer cells cause a resistant phenotype in target cells by upregulation of cytoprotective autophagy [84]. Exosomal miRNAs can control autophagy and play a role as mediators in therapeutic resistance [36, 85, 86]. For example, cisplatin-resistant non-small cell lung cancer (NSCLC) cells secreted exosomal miR-425-3p, which targeted the AKT1/mTOR signaling pathway and subsequently led to the upregulated autophagy activity and reduced cisplatin treatment outcomes [87].

# 1.1.2. Exosomal lncRNAs (Long Non-coding RNAs)

The transfer of exosomal lncRNAs between the tumor microenvironment and tumor cells is involved in events such as reprogramming the tumor microenvironment, growth, migration, and survival of cancer cells, as well as the development of mechanisms that cause resistance to chemotherapy [88]. In addition, because exosomal lncRNAs play a role in malignancy and response to treatment, they could be employed as biomarkers for many cancers [88]. There is evidence that lncRNAs promote drug resistance in diverse cancers [88]. For instance, lncRNA SBF2 (SET binding factor 2) induces temozolomide resistance in glioblastoma cells during chemotherapy [89]. During tumor growth, access to oxygen is limited due to poor vasculature within the developing solid tumor mass, which is called hypoxia [88]. Under hypoxia conditions, the hypoxia-inducible factor (HIF)-1 $\alpha$  pathway is activated in tumor cells, which during oxygen stress, plays a role as an adaptive mechanism [90]. Moreover, the hypoxia condition promotes cell survival through the transcription of several lncRNAs [91, 92].

## 1.1.2.1. Exosomal lncRNAs and autophagy

Because of the replenishment of energy supply and protection from environmental stress, cancer cells upregulate autophagy, accordingly, autophagy is a significant event in cancer [93]. Studies have shown that because lncRNAs are key regulators of autophagy, they can protect cancer cells against chemotherapy and environmental stress [88]. In addition, autophagy induced by lncRNAs has a key role in the proliferation and survival of tumor cells mediated by CAFs [94].

## 1.1.2.2. Exosomal lncRNAs and metabolic programming

Metabolic activity in cancer cells is different from healthy cells [88]. In hypoxia conditions, healthy cells begin to perform aerobic glycolysis (Warburg effect), while cancer cells are highly dependent on it [95]. The cancer cells which are proliferated continuously require rapid production of ATP [88]. Furthermore, the rapid production of ATP leads to the synthesis of many glycolytic intermediates which go to subsidiary pathways and supply energy for proliferating cancer cells [96]. In the metabolic reprogramming of cancer cells, several signaling pathways play a role, including:

phosphatidylinositol 3-kinase (PI3K)/Akt, c-Jun N-terminal kinase (JNK),

extracellular signal-regulated kinase (ERK), and Ras [97]. Because lncRNAs can regulate these signaling pathways, lncRNAs can affect the metabolism of cancer cells [88].

# 1.2. Extracellular matrix (ECM):

The ECM is a complex and diverse network of more than a hundred proteins, including proteoglycans (heparan sulfate, chondroitin sulfate), fibrous proteins (elastin, collagen), glycoproteins (laminins, tenascin C (TNC), fibronectin 1 (FN1)), and glycosaminoglycans (hyaluronic acid), which plays a role as a scaffold of organs and tissues, as well as constitutes the largest component of the TME [6, 19, 98]. Although many tumor cells and diverse types of stromal cells can produce ECM proteins, CAFs have a main and significant role in the organization and composition of ECM [99, 100]. Moreover, the ECM contains cytokines, chemokines, and growth factors released by stromal and tumor cells [6]. The ECM regulates cell behavior and plays an important role in tissue function and maintenance [101, 102]. Accordingly, disruption of the mechanisms involved in the regulation of ECM degradation, production, and remodeling causes pathological conditions such as cancer and fibrosis [101, 103, 104]. Furthermore, necessary signals for cellular differentiation, growth, and migration are provided by ECM [102]. Each component of the extracellular matrix, through cell surface receptors, induces signaling pathways to cells and plays a role in tumor events including differentiation, survival, migration, and metabolism [105, 106]. Moreover, ECM heterogeneity provides evading growth suppressors, resisting cell death, and sustaining growth signals for cells, as a result, has a key role in tumor proliferation, also in tumor angiogenesis, metastasis, and invasion [106, 107]. The ECM is responsible for cellular migration out of the TME and cellular adhesion, although it functions as a physical scaffold for cells [19]. The presence of stored diverse soluble factors such as growth factors, cytokines, chemokines, and angiogenic factors in the extracellular matrix creates a continuous inflammatory condition that ultimately leads to the expansion of the cellular repertoire [19, 108, 109]. In this inflammatory condition, the deposition of a large amount of ECM protein occurs via facilitation in the transformation of stromal fibroblasts into myofibroblasts, which results in contraction and an increase in stiffness [109, 110]. One of the determining and important factors in TME involved in cancer treatment resistance is cell adhesion to ECM [111]. The interactions of ECM components such as laminin, fibronectin, and collagen with integrin provide a context for drug resistance mediated by cell adhesion [111]. The efficacy of many drugs depends on the composition of the cancer's ECM [102]. The dynamic adaptation of ECM is involved in the invasion and progression of cancer and especially drug resistance, for instance, ECM can adequately hinder drug delivery by increasing the remodeling of microvascular endothelial cells [112]. Indeed, ECM remodeling creates a physical obstacle that delays or prevents drug delivery, thereby it increases drug resistance [113]. Furthermore, ECM can activate survival proteins through survival pathways including MAPK, p53, PI3K/AKT, and subsequently promote chemoresistance [102]. Alterations in stiffness and elasticity of ECM impact drug delivery to cancer cells, as well as pressure and diffusion, are factors related to drug delivery in the interstitial spaces [102]. Drugs are delivered to tumors through the pressure of blood circulation and the interstitial areas [111]. In the interstitial areas, the ECM organization causes an increase of fluid pressure through the physical obstacles of the tumor mass and subsequently extremely prevents the desired results of drug delivery [111]. In addition, an increase in the fluid flux from the neoplasms to the surrounding environment due to the ample proliferation of cancer cells prevents the adequate transfer of drugs [114]. Indeed, the density of ECM cells plays a key role in the low efficiency of drug delivery [115].

Although all ECM components play a key role in tumor progression, the role of collagen stands out [98]. Collagen can affect the behavior of tumor cells through discoidin domain receptors, tyrosine kinase receptors, integrins, and several signaling pathways [116]. Furthermore, collagen can affect the activity of cancer cells by interacting with ECM molecules including MMPs, lamins, fibronectin, and hyaluronic acid [117]. Exosomes and miRNAs are other components that have close interactions with collagen in cancer [118, 119]. In the status that tumors are rich in collagen, the hypoxic condition is common, leading to the promotion of cancer progression [116]. However, heparan sulfate (CGKRK peptide), gelatin (anginex, small geletic-1 binding peptide), laminin (IKVAV), fibronectin extra domain A and B (anti-EDB aptide), and aggrecan (a conjugate of quaternary ammonium) are some components of ECM that have therapeutic value [98].

## 1.3. Cancer-associated fibroblasts (CAFs):

Non-malignant stromal cells in the tumor microenvironment are crucial for drug resistance and tumor progression [112]. Stromal cells are polarized by their interaction with tumor cells, and through education of tumor cells, they secrete diverse molecules, resulting in pharmacokinetics regulation, immunosuppression, and metabolic regulation, subsequently, they reduce therapeutic outcomes [112]. The most common type of stromal cells are CAFs. CAFs by involvement in ECM remodeling and metabolic reprogramming, as well as by secretion of diverse factors such as cytokines (IL-6, IL-8, IL-10, etc.), growth factors (hepatocyte growth factor, stromal cell-derived factor, fibroblast growth factor, transforming growth factor), various chemokines, metabolites, and exosomes (contain lncRNAs, miRNAs and mitochondrial DNA (mtDNA)) which activate multiple signaling cascades, cause drug resistance, and ultimately tumor recurrence [111, 112, 120].

# 1.4. Mesenchymal stem cells (MSCs):

MSCs (or mesenchymal stromal cells) have the potential for self-renewal and differentiation into various cell types [102, 106, 112]. Moreover, MSCs can be involved in many cancer events such as epithelial-mesenchymal transition, angiogenesis, anti-apoptosis, metastasis, immunosuppression, pro-survival, and treatment resistance especially drug resistance [112]. These cells can be inherently resistant to chemotherapy and in various cancer types, they expand this resistance among cancer cells [121-127]. MSCs can contribute to drug resistance and quiescence of cancer cells by creating a pre-metastatic niche for tumor cells [128]. Furthermore, they can move to inflammatory areas, and subsequently, infiltrate the tumor [106]. Indeed, MSCs migrate to the tumor site via the action of factors derived from tumor cells, then, infiltrate the tumor and produce the necessary factors for cancer cells [102]. Mesenchymal stem cells can cause drug resistance through the following actions:

1. Direct cell-to-cell contact between MSCs and cancer cells that triggers multiple signaling cascades in tumors [112, 129].

2. Genetic mutations in MSCs: Genetic changes occur not only in tumor cells but also in non-malignant cells, leading to latent tumor recurrence in radiotherapy- and chemotherapy-treated patients [112, 130].

3. Secretion of soluble factors: MSCs can release a variety of fatty acids, cytokines, and growth factors that result in drug resistance [102, 112, 131, 132].

4. Differentiation of MSCs into CSC or CAF: Some characteristics of CSCs are enhancing colony-forming capacity and pluripotency, obtaining drug resistance along with loss of anchorage dependence, having the capability to promote of metastasis and tumor progression, and inherently resistant to chemotherapy, moreover, CAFs have a key role in drug resistance and CAF-MSC cells (a type of CAF cells possessing resembling function and phenotype of bone marrow-derived mesenchymal stem cell (BM-MSC) in the tumor stroma) are involved in tumor growth, the decline in cell apoptosis, increase in cell proliferation, and resistance to chemotherapy; therefore, by differentiating MSCs into CSC or CAF, they can cause treatment resistance [112].

5. Release of exosomes: According to multiple studies, exosomes released from MSCs promote chemotherapy resistance through mediation in interactions between cancer cells and MSCs, specific mRNA molecules and proteins transportation, and drug sequestration [112, 133].

## 1.5. Cancer-associated adipocytes (CAAs):

Cancer-associated adipocytes (CAAs) are a type of stromal cells and an important factor in TME that play a role in resistance to apoptosis, angiogenesis, metastasis, drug resistance, and cancer cell invasion [134-136]. CAAs promote cancer cell malignancy and tumor growth by secreting pro-inflammatory cytokines, hormones, adipokines, adiponectin, resistin, and leptin [109, 137, 138]. CAAs can also cause chemoresistance by secreting exosomes, for instance, exosomal miR21s derived from CAAs can inhibit apoptosis of cancer cells and promote chemoresistance by binding to apoptotic protease-activating factor 1 (APAF1) in ovarian cancer [62, 109].

In general, CAAs cause treatment resistance through the following actions:

1. Secretion of various factors: Adipose tissue acts both as a site for energy storage and as an endocrine organ, producing a wide range of exosomes, adipokines, leptin, growth factors, and adipocytokines that lead to treatment resistance [112, 139, 140].

2. Extracellular matrix remodeling: CAAs are a crucial source for ECM components, also, the property of ECM dynamic adaptation has an important role in the invasion and progression of cancer, as well as drug resistance [112]. CAAs release factors such as matrix metalloproteinases (MMPs) and collagen VI protein, resulting in the remodeling of ECM and chemotherapy resistance [141-144]. On the whole, according to several studies, lipid metabolism and CAAs have an intricate role in the regulation of cancer sensitivity to anticancer drugs [112].

3. Metabolism regulation: Since adipocytes perform energy storage, the metabolic relationship between tumor cells and adipocytes can naturally contribute to tumor progression [112]. Similar to CAFs, the "Warburg effect" and "reverse Warburg effect" could also cause drug resistance mediated by adipocytes [139]. The amount of lactate produced by adipocytes under hypoxia conditions increases significantly [145]. Adipocytes also provide lipids for cancer cells to supply their main energy, which can cause treatment resistance and tumor progression, for instance, cancer cells take up exogenous free fatty acids (FAAs) released by adipocytes and these endogenous lipid molecules enhance the rate of fatty acid  $\beta$ -oxidation (FAO), leading to extensive synthesis of aATP [112, 146].

4. Alteration in the pharmacokinetics of chemotherapy: Pharmacokinetics of chemotherapy is altered by CAAs commonly via two actions:

(a) Increase in drug clearance, (b) Alteration in drug distribution [112]. Since the concentration of active drugs during treatment plays a key role in the efficacy of cancer therapy, in adipocyte-rich microenvironments such as adipose tissue, it is observed a local decrease in cytotoxic chemotherapy activity due to decreased concentration, which could contribute to chemoresistance [112, 147].

## 1.6. Acidosis:

Acidification of the tumor microenvironment is related to metabolic adaptation and reprogramming [148]. Moreover, tumor acidosis affects metastasis, invasion, and therapeutic response and can act as a regulator of tumor progression [148]. Tumor cells have high metabolic demands that lead to the accumulation of a high amount of n the TME, and due to the disordered nature of tumor vasculature, effective removal of  $H^+$  from the extracellular environment is prevented, accordingly, the accumulation of  $H^+$  ions in the TME is intensified [148, 149]. Consequently, it causes hypoxic conditions and changes in glycolytic metabolism. In addition, in areas where oxidative conditions prevail, the accumulation of  $H^+$  causes the hydration of carbon dioxide, which meets the bioenergetic and biosynthetic needs of cancer cells, these processes happen at a high speed [148]. Low pH in TME can promote the motility of cancer cells and subsequently, influence fibroblasts and activity and polarization of macrophages by bringing about alterations in cytoskeletal dynamics [150]. In tumor regions where the lowest pH conditions prevail, the highest rate of invasion can occur and vice versa [148]. To adapt to the acidic environment, tumor cells employ all enzyme systems [106]. In general, the establishment of acidic conditions in the TME involves two events: CO2 by respiration and lactic acid formed by glycolytic metabolism [106]. Evidence shows that acidic conditions overlap both in hypoxic areas and at the tumor-stroma interface, which strongly influences tumor invasion and proliferation [151]. Moreover, according to multiple evidence, the conditions of extracellular acidosis create a suitable and beneficial environment for dormant tumor cells to support the survival of disseminated tumor cells and the formation of metastasis, and subsequently, the chemotherapy-resistant phenotype is maintained [106, 152].

# 1.7. Metabolic reprogramming:

Changes in energy metabolism are one of the specifications of tumor cells since tumor cells proliferate abnormally and accordingly, are dependent on the increased adaptation to the nutritional microenvironment, which is operated by the metabolic reprogramming [112, 153]. Due to the glucose deficiency in the tumor microenvironment, the metabolism of cancer cells shifts to aerobic glycolysis (the Warburg effect) [154]. The final product of this metabolic process is lactate, which is associated with treatment resistance [112, 155]. According to some evidence, other metabolic features such as reversed Warburg effect, glutamine metabolism, and metabolic symbiosis can cause acquired or adaptive drug resistance and challenge anti-tumor treatment [156]. Evidence suggests that disordered metabolism is involved in metastasis and malignancy [111]. Severe reduction of amino acids such as glutamine, tryptophan, and arginine and high levels of glycolysis are observed in cancer cells [111]. The metabolic features of malignant cells are regulated by tumor metabolic stress, leading to acidity, nutrient deficiency, and oxygen competition in the TME [157]. Apart from epigenetic and genetic alterations of cancer cells, the interactions between various components of TME and metabolic competition contribute to drug resistance and the growth and metastasis of tumor cells [158, 159].

# 1.8. Interstitial fluid pressure (IFP):

Abnormal lymphatic and blood vessels cause acidic pH, hypoxia, and high interstitial fluid pressure (IFP) in TME and elevated IFP is a key player in preventing adequate drug delivery to solid tumors [106]. Factors such as abnormal ECM, high cell density, disruption of lymphatic or venous drainage, and enhanced vascular permeability cause high IFP in the tumor [160]. Abnormal proliferation of cancer cells leads to mechanical compression of blood vessels and lymphatic vessels in the confined area of the microenvironment, subsequently, meager lymphatic drainage and poor blood flow are caused, leading to abnormal vascular structures and a further reduction in a number of functional lymphatic vessels [161]. As a result of excess fluid leakage from the vascular system into interstitial space or interstitium (where accumulates and swells the elastic ECM), IFP is increased, which is higher than in normal tissue [106].

The amount of drug entering the tumor and its absorption by cancer cells affects the efficacy of chemotherapy [162]. For a drug to reach cancer cells through blood vessels, it must be transferred from blood vessels to interstitial fluid and then from interstitial space to tissues [163]. Therefore, the amount of interstitial fluid pressure is important for drug delivery, and high IFP conditions can reduce drug transfer [162]. High IFP also compresses blood vessels, diverting blood away from the tumor center to the periphery, as a result, drug delivery is further reduced [162].

# 2. Immune cells:

The immune system plays a crucial role in monitoring cancer and responding to chemotherapy. Myeloidderived suppressor cells (MDSCs), tumor-associated macrophages (TAMs), dendritic cells (DCs), regulatory and effector T cells, B cells, and natural killer cells (NKs) are the main immune cells present in the TME [15, 164]. These cells can have either opposing or stimulating effects on the tumor and play a key role in tumorigenesis [15]. Immune cells also contribute to the development of chemoresistance [165]. According to several studies, the relationship between MDSCs and malignant cells has a significant impact on chemotherapy resistance and immune system suppression [112, 166, 167]. At different stages of tumor development, diverse populations of T cells are observed in the tumor microenvironment [168, 169]. An increase in T-regulatory lymphocytes (T-reg) infiltration in the tumor microenvironment is associated with chemoresistance in several types of cancer, including colorectal, lung, kidney, HNSCC, melanoma, ovarian, and glioblastoma cancer [170-176]. Evidence suggests that tumor cells may manipulate local DCs to form suppressive T cells, ultimately leading to drug resistance [177, 178]. Drug resistance and tumor progression can be caused by high infiltration of tumor-associated neutrophils (TANs) in the TME [179]. TANs can lead to acquired drug resistance in cancer due to their capacity to increase angiogenesis, increase tumor cell proliferation, and suppress the immune system [180, 181]. Moreover, TANs can reduce the efficacy of many cancer drugs such as common cytotoxic drugs and immune checkpoint inhibitors by activating various signaling pathways [168]. Although NK cells may exhibit multi-drug resistance-like activity, according to studies, this property can be inhibited by drugs such as solutol HS-15 or verapamil [168, 182-184].

TAMs are derived from circulating monocytes and are one of the most abundant cells in solid tumors that have a significant impact on suppressing the immune system in the TME, furthermore, TAMs play a role in chemoresistance and tumor development [111]. Among the immune cells present in the TME, macrophages have a prominent and critical role in chemoresistance due to their capabilities and high numbers. TAMs are the most abundant among immune cells in the TME, and their increased infiltration into the microenvironment leads to unfavorable outcomes in chemotherapy [112, 185, 186]. Generally, TAMs are divided into two subgroups, M1 and M2, with the M2 phenotype playing a role in promoting drug resistance and tumor progression [112, 187]. In cancer conditions, macrophages are educated by the TME, and TAMs are usually polarized from M1 to M2 [188]. Due to the interaction between TAMs and tumor cells, under the pressure of treatment, TAMs are promoted by tumor cells and differentiate into immunosuppressive M2-polarized macrophages, leading to therapeutic resistance [112]. Moreover, studies have demonstrated the type of M2 can lead to acquired drug resistance in cancer cells [189-191].

TAMs, like CAFs, release a wide range of soluble factors in the TME, including chemokines, enzymes, interleukins, exosomes, etc, to combat drug attacks, for instance, TAMs can prevent paclitaxel-induced tumor cell death by expressing cathepsin S and B [112]. Moreover, through overexpressed cytidine deaminase or CDA (an enzyme that is involved in gemcitabine degradation), TAMs can promote chemotherapy resistance in cancer [165, 192]. By secreting exosomal miR-365, TAMs can increase the metabolism of gemcitabine in cancer cells, which ultimately leads to apoptosis inhibition and tumor resistance promotion [193, 194]. TAMs upregulate Gfi-1 in tumor cells by TGF- $\beta$  secretion, which ultimately leads to reduced sensitivity of tumor cells to gemcitabine by inhibiting HMGB1 (high mobility group box 1) and CTGF (connective tissue growth factor) expression [195]. By expressing IGF, TAMs can cause resistance to chemotherapy with albuminbinding paclitaxel and gemcitabine [165, 196]. Additionally, TAMs can cause chemotherapy resistance in pancreatic cancer by inducing EMT [197]. According to evidence from prostate cancer treatment with ADT, CSCs can remodel macrophages into TAMs, subsequently, through the IL6/STAT3 signaling pathway, TAMs can increase stem-like features of CSCs and drug resistance [112, 198].

Overall, TAMs use various mechanisms to induce drug resistance, including regulating CSC properties, transforming into M2 suppressive phenotype, promoting EMT, releasing various cytokines, and suppressing immune cells [112].

## 3. DNA repair mechanisms:

Preserving the genome and transmitting a healthy genome to the next generation is an essential task for living organisms [199]. However, DNA is constantly exposed to both endogenous insults (such as intracellular free radical oxygen species (ROS), etc.) and exogenous genotoxic insults (such as ionizing radiation (IR), ultraviolet (UV) radiation, chemotherapeutic drugs, etc.), that can damage to DNA [199]. Although DNA damage is a crucial target for radiotherapy and chemotherapy, it can also lead to the development of cancer [199]. Therefore, to maintain genome integrity, living organisms rely on a complex system and multiple mechanisms to counteract DNA-damaging factors, collectively known as the DNA Damage Response (DDR) [200].

Several DNA repair pathways are utilized to counteract DNA damage, including:

- (1) Base excision repair (BER), (2) Nucleotide excision repair (NER),
- (3) Homologous recombination (HR), (4) Non-homologous end joining (NHEJ),
- (5) Mismatch repair (MMR), (6) Microhomology-mediated end joining (MMEJ),
- (7) DNA damage tolerance (DDT) (Translession synthesis (TLS), template switching (TS)),
- (8)  $O^6$ -methylguanine-DNA methyltransferase (MGMT) pathway,
- (9) Fanconi anemia (FA) pathway, (10) Single-strand annealing (SSA) [200-203]

Among these pathways, BER, MMR, NER, HR, and NHEJ are the major and essential pathways in DNA repair [200, 201].

Moreover, there are several types of DNA damage, including:

- (1) Clustered damaged sites, (2) Base damage, (3) Single-strand breaks (SSBs),
- (4) Double-strand breaks (DSBs), (5) Sugar damage, (6) DNA cross-linking [204]

DSBs are the most destructive and the most deleterious type of DNA damage for cells which can bring about cell death or carcinogenesis [204]. Among the types of DNA damage, SSBs and DSBs are prominent which can bring about genome rearrangement. The direct and indirect BER, MMR, and NER pathways repair SSBs damage, while the SSA, NHEJ, and HR pathways repair DSB damages [200, 205, 206]. Moreover, DNA adducts and replication errors are repaired by the NER and MMR pathways, respectively [200].

To control the DDR, cells utilize epigenetics and miRNAs as regulators. Epigenetic alterations in gene expression and tumor heterogeneity play a key role, as a result, epigenetic chromatin regulation can influence the mechanisms and pathways involved in the DNA repair process [207]. Histone deacetylases (HDACs) can contribute to the preparation of chromatin for DSB repair promotion via NHEJ and HR [207]. Furthermore, DNA methylation is a common epigenetic mechanism in cancer cells and gene inactivation [207]. Alterations in gene promoter methylation status of DDR components are observed in diverse cancer types including oral squamous cell carcinoma, thyroid cancer, non-small cell lung cancer (NSCLC), neck squamous cell carcinoma, gastric cancer, acute myeloid leukemia (AML), breast cancer, ovarian cancer, bladder cancer [207-218]. Moreover, the methylation status of some DDR genes can be employed as treatment response, prognostic, and diagnostic biomarkers in diverse types of cancer [207]. miRNAs function as regulators in various processes, including tumorigenesis, and post-transcriptional control of DNA repair components, in addition, miRNAs can regulate the expression levels of DNA repair genes and subsequently modulate the sensitivity of cancer cells to DNA-damaging agents [207].

Chemotherapeutic agents commonly used include Topoisomerase I inhibitors, Alkylating agents (such as cisplatin), and DNA Topoisomerase II inhibitors [219]. During chemotherapy using Camptothecin (a Topoisomerase I inhibitor), if SSB damage occurs, the BER pathway is activated, and subsequently, PARP1 and APE1 enzymes are activated, however, if DSB damage occurs, the HR and NHEJ pathways are activated, followed by the HR pathway activating AMT and CHK1 enzymes, and NHEJ activating DNA-PK enzyme [202]. When Etoposide (a Topoisomerase II inhibitor) is prescribed, DSB damage occurs, which activates the HR and NHEJ pathways [202]. The HR pathway activates ATM and CHK1 proteins, and NHEJ activates DNA-PK [202]. When Cisplatin (an Alkylating agent) is prescribed, DNA interstrand cross-link (ICL) damage (activating HR and NER pathways) and intrastrand cross-link damage (activating NER pathway) occur [202]. Then, the HR pathway activates ATM and CHK1 proteins, and the NER pathway activates XPA, XPB, and XPG proteins [202].

## 4. Cancer Stem Cells (CSCs)

One of the reasons for cancer progression and treatment failure is cancer heterogeneity [220]. Different cancer cell types in the tumor contribute to tumor heterogeneity, and among these cells, cancer stem cells (CSCs) are highly involved in the initiation and progression of cancer as well as have self-renewal and differentiation abilities [221]. Stem cells in cancers can be divided into two categories based on their function:

(1) Resident cancer stem cells that can initiate the tumor,

(2) Migratory stem cells that metastasize and form tumors in another location [221].

Due to these capabilities, CSCs play a key role in tumor initiation, drug resistance, metastasis, and cancer recurrence [222]. During successful chemotherapy, although a significant portion of tumor cells undergo apoptosis, a subset of CSCs may survive and cause cancer recurrence [223, 224].

There is a wide range of mechanisms and factors that contribute to CSCs to promote chemoresistance, including:

(1) Tumor microenvironment (Autophagy, Inflammation, Adipocyte-released factors, CAFs, Mesenchymal stem cells (MSCs), Extracellular matrix (ECM), Hypoxia, Endothelial cells (ECs), Immune cells), (2) EMT induction or activation of EMT-transcription factors,

(3) Self-renewal ability (high telomerase activity),

(4) High expression of CSCs markers (such as CD133, ALDH1, CD44, CD24+),

- (5) Quiescence / Dormancy / low proliferation rate,
- (6) Stemness genes (such as Bmi1 and Musashi (MSI)),
- (7) Epigenetic mechanisms (DNA methylation, Histone modifications),
- (8) Signaling pathways (such as Hedgehog pathways, Notch pathways, and Wnt pathways),

(9) Resistant to DNA damage-induced cell death (Promoting the DNA repair capability, Enhancing ROS scavenging, Activating the anti-apoptotic signaling pathways), (10) Metabolism alteration,

(11) Higher expression of multi-drug resistance (MDR) or detoxification proteins (Aldehyde dehydrogenase (ALDH), Drug-transporter proteins (ABCG1, ABCB1)),

(12) Non-coding RNAs (ncRNAs) [220, 225-230].

There is extensive evidence indicating that ncRNAs, including lncRNAs and miRNAs, play a key role in regulating CSC capabilities such as asymmetric cell division, cancer recurrence, tumor initiation, self-renewal, and drug resistance [220, 231-236]. Moreover, according to multiple studies, ncRNAs control the cancer progression and growth and division of cancer stem cells by regulating downstream signaling pathways and transcription factors [226, 237-240].

# **Discussion and Conclusion:**

Cancer cells employ a wide range of factors and mechanisms to resist chemotherapy. Among these factors, the triad of CSCs, exosomes, and ncRNAs are particularly prominent. CSCs not only utilize diverse intracellular and extracellular mechanisms to develop chemoresistance but also have a significant role in chemoresistance through their interactions with various components of TME and other cancer cells. Furthermore, It has also been established that CSCs play a crucial role in recurrence after successful chemotherapy. As previously mentioned, exosomes present in the microenvironment have a prominent role in cellular communications and interactions between different components of TME and cancer cells. Additionally, ncRNAs have an important role in intracellular signaling pathways and intercellular communications between CSCs and other factors. Therefore, it can be deduced that ncRNAs are likely to have a significant impact on the function of mentioned factors involved in chemotherapy resistance caused by CSCs. Moreover, CSCs can utilize their high capacity of survival and chemoresistance to stay alive after chemotherapy, as well as employ the combination of exosomes and ncRNAs to spread chemoresistance among other cancerous cells by establishing wide intercellular communications with tumor cells and the tumor microenvironment components, ultimately leading to the exacerbation of chemotherapy resistance and cancer relapse. Indeed, ncRNA can be considered the most prominent factor among other factors in the chemoresistance of CSCs. Therefore, an in-depth understanding of the interactions between CSCs, exosomes, and ncRNAs is very important and vital in achieving the desired chemotherapy results, and research in these three fields is very necessary to understand their precise function in chemotherapy resistance.

In addition, it is recommended that the use of smart nanoparticles as a drug delivery method for small molecules could be an effective approach for delivering a greater and more fruitful amount of small molecules to cancer cells, considering the distinct conditions within the tumor microenvironment (hypoxia, acidity, etc.). One of the challenges in drug delivery is the limited understanding of drug targets. Large-scale screening studies using CRISPR-Cas technology can help to improve our understanding of high-priority drug targets.

On the whole, the application of the following strategies can improve the design and therapeutic outcomes of small molecule drugs as combination therapy for chemotherapy:

1. Focusing more research on interactions between CSCs, exosomes, and ncRNAs as well as their function in chemotherapy resistance for a better and deeper understanding of these areas.

2. Identifying the factors that maintain proper function and favorable interactions between CSCs, exosomes, and ncRNAs to be considered as targets for small molecule drugs so that targeting them causes effective

disruption in the function and interactions of these three components.

3. Launching extensive screening studies using CRISPR-Cas technology to identify potential drug targets for the design of effective small molecule drugs.

4. More research on drug delivery systems based on smart nanoparticles to improve and increase their efficiency in the appropriate and sufficient delivery of small molecule drugs to the tumor microenvironment and cancer cells.

#### **Conflict of Interest:**

No conflict of interest has been expressed by the author.

# **References:**

1. Zhu, R., et al., *Current progress in cancer treatment using nanomaterials*. Frontiers in Oncology, 2022. **12**.

2. Cheng, Z., et al., Nanomaterials for cancer therapy: Current progress and perspectives. Journal of hematology & oncology, 2021.14 (1): p. 1-27.

3. Sun, G., et al., Role of small molecule targeted compounds in cancer: progress, opportunities, and challenges. Frontiers in cell and developmental biology, 2021. **9** : p. 694363.

4. Zhong, L., et al., *Small molecules in targeted cancer therapy: Advances, challenges, and future perspectives.* Signal transduction and targeted therapy, 2021. **6** (1): p. 201.

5. Tan, S., et al., *Exosomal miRNAs in tumor microenvironment*. Journal of Experimental & Clinical Cancer Research, 2020. **39** : p. 1-15.

6. Zhong, S., et al., *Targeting tumor microenvironment by small-molecule inhibitors*. Translational oncology, 2020.13 (1): p. 57-69.

7. Lucero, R., et al., *Glioma-derived miRNA-containing extracellular vesicles induce angiogenesis by repro*gramming brain endothelial cells. Cell reports, 2020. **30** (7): p. 2065-2074. e4.

8. Varricchi, G., et al., Innate effector cells in angiogenesis and lymphangiogenesis. Current opinion in immunology, 2018.53 : p. 152-160.

9. Petrova, V., et al., The hypoxic tumour microenvironment. Oncogenesis, 2018. 7 (1): p. 10.

10. Zeltz, C., et al. Cancer-associated fibroblasts in desmoplastic tumors: emerging role of integrins . in Seminars in cancer biology . 2020. Elsevier.

11. Ma, L., et al., *Tumor cell biodiversity drives microenvironmental reprogramming in liver cancer*. Cancer cell, 2019.36 (4): p. 418-430. e6.

12. Zhang, X., et al.,  $I\nu\eta\beta\pi\nu\gamma$  III3  $\kappa\nu\alpha\sigma\epsilon\gamma\nu$   $\rho$   $\beta\sigma\tau\eta\mu\psi\epsilon\lambda\sigma\delta$   $\alpha\nu\delta$   $\pi\lambda\alpha\sigma\mu\alpha$   $\epsilon\lambda\lambda$  $\rho\epsilon\mu\delta\epsilon\lambda$  $\zeta$   $\tau\eta\epsilon$   $\sigma\nu\pi\rho\epsilon\sigma\sigma\epsilon\epsilon$  $\tau\nu\mu\rho\rho\mu\kappa\rho\sigma\epsilon\nu\rho\sigma\mu\epsilon\nu\tau$   $\nu$   $\delta\epsilon\sigma\mu\sigma\lambda\alpha\sigma\tau\kappa$   $\tau\nu\mu\rho\rho\zeta$ . Journal of Controlled Release, 2019. **309** : p. 173-180.

13. Clément-Colmou, K., et al., Influence of radiotherapy fractionation schedule on the tumor vascular microenvironment in prostate and lung cancer models. Cancers, 2020. **12** (1): p. 121.

14. Armignacco, R., et al., The adipose stem cell as a novel metabolic actor in adrenocortical carcinoma progression: Evidence from an in vitro tumor microenvironment crosstalk model. Cancers, 2019.11 (12): p. 1931.

15. Lei, X., et al., Immune cells within the tumor microenvironment: Biological functions and roles in cancer immunotherapy. Cancer letters, 2020. **470** : p. 126-133.

16. Zhang, H., et al., *CAF secreted miR-522 suppresses ferroptosis and promotes acquired chemo-resistance in gastric cancer*. Molecular cancer, 2020. **19** : p. 1-17.

17. De Palma, M., D. Biziato, and T.V. Petrova, *Microenvironmental regulation of tumour angiogenesis*. Nature Reviews Cancer, 2017.17 (8): p. 457-474.

 Moriwaki, K. and M. Asahi, Augmented TME O-GlcNAcylation Promotes Tumor Proliferation through the Inhibition of p38 MAPKEffect of O-GlcNAcylation in the TME. Molecular Cancer Research, 2017.15 (9): p. 1287-1298.

19. Khalaf, K., et al., Aspects of the tumor microenvironment involved in immune resistance and drug resistance. Frontiers in immunology, 2021. **12**: p. 656364.

20. Whiteside, T.L. Exosome and mesenchymal stem cell cross-talk in the tumor microenvironment . in Seminars in immunology . 2018. Elsevier.

21. Xie, F., et al., *Extracellular vesicles in cancer immune microenvironment and cancer immunotherapy*. Advanced science, 2019.6 (24): p. 1901779.

22. Daassi, D., K.M. Mahoney, and G.J. Freeman, *The importance of exosomal PDL1 in tumour immune evasion*. Nature Reviews Immunology, 2020. **20** (4): p. 209-215.

23. Jarosz-Biej, M., et al., *Tumor microenvironment as a "game changer" in cancer radiotherapy*. International journal of molecular sciences, 2019. **20** (13): p. 3212.

24. Peng, J., et al., Intratumoral fate of functional nanoparticles in response to microenvironment factor: Implications on cancer diagnosis and therapy. Advanced Drug Delivery Reviews, 2019.143 : p. 37-67.

25. Maacha, S., et al., Extracellular vesicles-mediated intercellular communication: roles in the tumor microenvironment and anti-cancer drug resistance. Molecular cancer, 2019. **18** : p. 1-16.

26. Sullivan, R., et al., The emerging roles of extracellular vesicles as communication vehicles within the tumor microenvironment and beyond. Frontiers in Endocrinology, 2017. 8 : p. 194.

27. Van Niel, G., G. d'Angelo, and G. Raposo, *Shedding light on the cell biology of extracellular vesicles*. Nature reviews Molecular cell biology, 2018. **19** (4): p. 213-228.

28. Minciacchi, V.R., M.R. Freeman, and D. Di Vizio. Extracellular vesicles in cancer: exosomes, microvesicles and the emerging role of large oncosomes. in Seminars in cell & developmental biology. 2015. Elsevier.

29. Maas, S.L., X.O. Breakefield, and A.M. Weaver, *Extracellular vesicles: unique intercellular delivery vehicles*. Trends in cell biology, 2017. **27** (3): p. 172-188.

30. Bebelman, M.P., et al., *Biogenesis and function of extracellular vesicles in cancer*. Pharmacology & therapeutics, 2018.188 : p. 1-11.

31. Tovar-Camargo, O.A., S. Toden, and A. Goel, *Exosomal microRNA biomarkers: emerging frontiers in colorectal and other human cancers*. Expert review of molecular diagnostics, 2016. **16** (5): p. 553-567.

32. Higginbotham, J.N., et al., *Amphiregulin exosomes increase cancer cell invasion*. Current Biology, 2011. **21** (9): p. 779-786.

33. Rak, J. and A. Guha, *Extracellular vesicles-vehicles that spread cancer genes*. Bioessays, 2012. **34** (6): p. 489-497.

34. Naito, Y., et al., *How cancer cells dictate their microenvironment: present roles of extracellular vesicles.* Cellular and Molecular Life Sciences, 2017. **74** : p. 697-713.

35. Fu, H., et al., *The emerging roles of exosomes in tumor-stroma interaction*. Journal of cancer research and clinical oncology, 2016. **142** : p. 1897-1907.

36. Santos, P. and F. Almeida, *Role of exosomal miRNAs and the tumor microenvironment in drug resistance*. Cells, 2020. **9** (6): p. 1450.

37. Xu, R., et al., *Extracellular vesicles in cancer—implications for future improvements in cancer care.* Nature reviews Clinical oncology, 2018. **15** (10): p. 617-638.

38. Li, I. and B.Y. Nabet, *Exosomes in the tumor microenvironment as mediators of cancer therapy resistance*. Molecular cancer, 2019.18 : p. 1-10.

39. Gangoda, L., et al., Extracellular vesicles including exosomes are mediators of signal transduction: are they protective or pathogenic? Proteomics, 2015. 15 (2-3): p. 260-271.

40. Steinbichler, T.B., et al., *Therapy resistance mediated by exosomes.* Molecular cancer, 2019. **18** (1): p. 1-11.

41. Mashouri, L., et al., *Exosomes: composition, biogenesis, and mechanisms in cancer metastasis and drug resistance.* Molecular cancer, 2019. **18** : p. 1-14.

42. Milman, N., L. Ginini, and Z. Gil, *Exosomes and their role in tumorigenesis and anticancer drug resistance*. Drug Resistance Updates, 2019. **45** : p. 1-12.

43. Anfossi, S., et al., *MicroRNAs, regulatory messengers inside and outside cancer cells.* Exosomes, Stem Cells and MicroRNA: Aging, Cancer and Age Related Disorders, 2018: p. 87-108.

44. Fattore, L., et al., *Reprogramming miRNAs global expression orchestrates development of drug resistance in BRAF mutated melanoma*.Cell Death & Differentiation, 2019. **26** (7): p. 1267-1282.

45. Malhotra, A., et al., Stabilization of miRNAs in esophageal cancer contributes to radioresistance and limits efficacy of therapy.Biochimie, 2019. **156** : p. 148-157.

46. Yan, W., et al., Cancer-cell-secreted exosomal miR-105 promotes tumour growth through the MYCdependent metabolic reprogramming of stromal cells. Nature cell biology, 2018. **20** (5): p. 597-609.

47. Zhou, C.-F., et al., Cervical squamous cell carcinoma-secreted exosomal miR-221-3p promotes lymphangiogenesis and lymphatic metastasis by targeting VASH1. Oncogene, 2019. **38** (8): p. 1256-1268.

48. Cai, L., et al., *Epstein-Barr virus-encoded microRNA BART1 induces tumour metastasis by regulating PTEN-dependent pathways in nasopharyngeal carcinoma.* Nature communications, 2015. **6** (1): p. 7353.

49. Tian, R., et al., Differential expression of miR16 in glioblastoma and glioblastoma stem cells: their correlation with proliferation, differentiation, metastasis and prognosis. Oncogene, 2017. **36** (42): p. 5861-5873.

50. Uddin, M.N., M. Li, and X. Wang, Identification of Transcriptional Markers and microRNA-mRNA Regulatory Networks in Colon Cancer by Integrative Analysis of mRNA and microRNA Expression Profiles in Colon Tumor Stroma. Cells, 2019. 8 (9): p. 1054.

51. Slack, F.J. and A.M. Chinnaiyan, *The role of non-coding RNAs in oncology*. Cell, 2019. **179** (5): p. 1033-1055.

52. Kanchan, R.K., et al., microRNAs orchestrate pathophysiology of breast cancer brain metastasis: advances in therapy. Molecular cancer, 2020. **19** (1): p. 1-16.

53. Conti, I., et al., miRNAs as influencers of cell-cell communication in tumor microenvironment. Cells, 2020. 9 (1): p. 220.

54. Cheng, W.C., et al., *RAB27B-activated secretion of stem-like tumor exosomes delivers the biomarker* microRNA-146a-5p, which promotes tumorigenesis and associates with an immunosuppressive tumor microenvironment in colorectal cancer. International Journal of Cancer, 2019. **145** (8): p. 2209-2224.

55. Patel, H., et al., Modulating secreted components of tumor microenvironment: A masterstroke in tumor therapeutics. Cancer Biology & Therapy, 2018. **19** (1): p. 3-12.

56. Wang, M., et al., *Emerging function and clinical values of exosomal microRNAs in cancer*. Molecular therapy-Nucleic acids, 2019.16 : p. 791-804.

57. Sun, Z., et al., *Effect of exosomal miRNA on cancer biology and clinical applications*. Molecular cancer, 2018. **17** : p. 1-19.

58. Pontecorvi, G., et al., Tumor-derived extracellular vesicles and microRNAs: Functional roles, diagnostic, prognostic and therapeutic options. Cytokine & Growth Factor Reviews, 2020. **51** : p. 75-83.

59. Bach, D.H., et al., The role of exosomes and miRNAs in drug-resistance of cancer cells. International journal of cancer, 2017.141 (2): p. 220-230.

60. Seo, H.A., et al., Microrna-based combinatorial cancer therapy: Effects of micrornas on the efficacy of anti-cancer therapies. Cells, 2019. 9 (1): p. 29.

61. Sharma, A., Chemoresistance in cancer cells: exosomes as potential regulators of therapeutic tumor heterogeneity. Nanomedicine, 2017. **12** (17): p. 2137-2148.

62. Au Yeung, C.L., et al., Exosomal transfer of stroma-derived miR21 confers paclitaxel resistance in ovarian cancer cells through targeting APAF1. Nature communications, 2016. 7 (1): p. 11150.

63. Chen, Y., et al., *Tumor-associated macrophages: an accomplice in solid tumor progression*. Journal of biomedical science, 2019.26 (1): p. 1-13.

64. Netea-Maier, R.T., J.W. Smit, and M.G. Netea, *Metabolic changes in tumor cells and tumor-associated macrophages: a mutual relationship.* Cancer letters, 2018. **413** : p. 102-109.

65. Sawa-Wejksza, K. and M. Kandefer-Szerszeń, *Tumor-associated macrophages as target for antitumor therapy*. Archivum immunologiae et therapiae experimentalis, 2018. **66** : p. 97-111.

66. Park, J.E., et al., *Hypoxia-induced tumor exosomes promote M2-like macrophage polarization of infiltrating myeloid cells and microRNA-mediated metabolic shift.* Oncogene, 2019. **38** (26): p. 5158-5173.

67. Cooks, T., et al., Mutant p53 cancers reprogram macrophages to tumor supporting macrophages via exosomal miR-1246. Nature communications, 2018. 9 (1): p. 771.

68. Hsieh, C.-H., S.-K. Tai, and M.-H. Yang, Snail-overexpressing cancer cells promote M2-like polarization of tumor-associated macrophages by delivering MiR-21-abundant exosomes. Neoplasia, 2018.20 (8): p. 775-788.

69. Lin, F., et al., Bladder cancer cell-secreted exosomal miR-21 activates the PI3K/AKT pathway in macrophages to promote cancer progression. International journal of oncology, 2020. 56 (1): p. 151-164.

70. Chen, X., et al., *Exosomes derived from hypoxic epithelial ovarian cancer deliver microRNA-940 to induce macrophage M2 polarization.* Oncology Reports, 2017. **38** (1): p. 522-528.

71. Chen, X., et al., *Exosomes derived from hypoxic epithelial ovarian cancer cells deliver microRNAs to macrophages and elicit a tumor-promoted phenotype.* Cancer letters, 2018. **435** : p. 80-91.

72. Nieto, M.A., *Epithelial plasticity: a common theme in embryonic and cancer cells.* Science, 2013. **342** (6159): p. 1234850.

73. Gavert, N. and A. Ben-Ze'ev, *Epithelial-mesenchymal transition and the invasive potential of tumors*. Trends in molecular medicine, 2008. **14** (5): p. 199-209.

74. Du, B. and J.S. Shim, Targeting epithelial-mesenchymal transition (EMT) to overcome drug resistance in cancer. Molecules, 2016. **21** (7): p. 965.

75. Bigagli, E., et al., Transcriptomic characterization, chemosensitivity and regulatory effects of exosomes in spontaneous EMT/MET transitions of breast cancer cells. Cancer Genomics & Proteomics, 2019. 16 (3): p. 163-173.

76. Hardin, H., et al., *Thyroid cancer stem-like cell exosomes: regulation of EMT via transfer of lncRNAs.* Laboratory Investigation, 2018. **98** (9): p. 1133-1142.

77. Xiao, D., et al., Melanoma cell-derived exosomes promote epithelial-mesenchymal transition in primary melanocytes through paracrine/autocrine signaling in the tumor microenvironment. Cancer letters, 2016. **376** (2): p. 318-327.

78. Mizushima, N. and M. Komatsu, Autophagy: renovation of cells and tissues. Cell, 2011. 147 (4): p. 728-741.

79. Huang, F., B.-R. Wang, and Y.-G. Wang, Role of autophagy in tumorigenesis, metastasis, targeted therapy and drug resistance of hepatocellular carcinoma. World journal of gastroenterology, 2018.24 (41): p. 4643.

80. Folkerts, H., et al., *The multifaceted role of autophagy in cancer and the microenvironment*. Medicinal research reviews, 2019.**39** (2): p. 517-560.

81. Janji, B., G. Berchem, and S. Chouaib, *Targeting autophagy in the tumor microenvironment: new challenges and opportunities for regulating tumor immunity.* Frontiers in Immunology, 2018. **9** : p. 887.

82. Gewirtz, D.A., *Cytoprotective and nonprotective autophagy in cancer therapy*. 2013, Taylor & Francis. p. 1263-1265.

83. Dutta, S., et al., Interactions between exosomes from breast cancer cells and primary mammary epithelial cells leads to generation of reactive oxygen species which induce DNA damage response, stabilization of p53 and autophagy in epithelial cells. PloS one, 2014.9 (5): p. e97580.

84. Liu, D.X., et al., Exosomes derived from HBV-associated liver cancer promote chemoresistance by upregulating chaperone-mediated autophagy. Oncology Letters, 2019. **17** (1): p. 323-331.

85. Qu, Y., et al., Exosomes derived from miR-181-5p-modified adipose-derived mesenchymal stem cells prevent liver fibrosis via autophagy activation. Journal of cellular and molecular medicine, 2017.21 (10): p. 2491-2502.

86. Chen, J., et al., *Micro RNA-30a ameliorates hepatic fibrosis by inhibiting Beclin1-mediated autophagy.* Journal of Cellular and Molecular Medicine, 2017. **21** (12): p. 3679-3692.

87. Yuwen, D., et al., Prognostic role of circulating exosomal miR-425-3p for the response of NSCLC to platinum-based chemotherapy.Cancer Epidemiology, Biomarkers & Prevention, 2019. 28 (1): p. 163-173.

88. Pathania, A.S. and K.B. Challagundla, *Exosomal long non-coding RNAs: emerging players in the tumor microenvironment*. Molecular Therapy-Nucleic Acids, 2021. **23** : p. 1371-1383.

89. Zhang, Z., et al., Exosomal transfer of long non-coding RNA SBF2-AS1 enhances chemoresistance to temozolomide in glioblastoma. Journal of Experimental & Clinical Cancer Research, 2019.38 (1): p. 1-16.

90. Masoud, G.N. and W. Li,  $HI\Phi$ -1a  $\pi a \tau \eta \omega a \psi$ :  $\rho o \lambda \epsilon$ ,  $\rho \epsilon \gamma \upsilon \lambda a \tau i o \nu \tau \epsilon \rho \epsilon \nu \tau i o \nu \phi o \rho \varsigma a \nu \varsigma \epsilon \rho \tau \eta \epsilon \rho a \pi \psi$ . Acta Pharmaceutica Sinica B, 2015.5 (5): p. 378-389.

91. Takahashi, K., et al., *Modulation of hypoxia-signaling pathways by extracellular linc-RoR*. Journal of cell science, 2014.127 (7): p. 1585-1594.

92. Xue, M., et al., *Hypoxic exosomes facilitate bladder tumor growth and development through transferring long non-coding RNA-UCA1*. Molecular cancer, 2017. **16** : p. 1-13.

93. Sun, K., et al., Paradoxical roles of autophagy in different stages of tumorigenesis: protector for normal or cancer cells. Cell & bioscience, 2013. 3 (1): p. 1-8.

94. Ding, L., et al., A novel stromal lncRNA signature reprograms fibroblasts to promote the growth of oral squamous cell carcinoma via LncRNA-CAF/interleukin-33. Carcinogenesis, 2018. **39** (3): p. 397-406.

95. Warburg, O., F. Wind, and E. Negelein, *The metabolism of tumors in the body*. The Journal of general physiology, 1927.8 (6): p. 519.

96. Liberti, M.V. and J.W. Locasale, *The Warburg effect: how does it benefit cancer cells?* Trends in biochemical sciences, 2016.41 (3): p. 211-218.

97. Papa, S., P.M. Choy, and C. Bubici, *The ERK and JNK pathways in the regulation of metabolic reprogramming*. Oncogene, 2019.38 (13): p. 2223-2240.

98. Baghban, R., et al., *Tumor microenvironment complexity and therapeutic implications at a glance*. Cell Communication and Signaling, 2020. **18** : p. 1-19.

99. Liu, T., et al., *Cancer-associated fibroblasts build and secure the tumor microenvironment*. Frontiers in cell and developmental biology, 2019. **7** : p. 60.

100. Walker, C., E. Mojares, and A. del Río Hernández, *Role of extracellular matrix in development and cancer progression*. International journal of molecular sciences, 2018. **19** (10): p. 3028.

101. Lu, P., V.M. Weaver, and Z. Werb, *The extracellular matrix: a dynamic niche in cancer progression*. Journal of cell biology, 2012.**196** (4): p. 395-406.

102. Senthebane, D.A., et al., The role of tumor microenvironment in chemoresistance: to survive, keep your enemies closer. International journal of molecular sciences, 2017. **18** (7): p. 1586.

103. Page-McCaw, A., A.J. Ewald, and Z. Werb, *Matrix metalloproteinases and the regulation of tissue remodelling*. Nature reviews Molecular cell biology, 2007. 8 (3): p. 221-233.

104. Cox, T.R. and J.T. Erler, *Remodeling and homeostasis of the extracellular matrix: implications for fibrotic diseases and cancer*. Disease models & mechanisms, 2011. 4 (2): p. 165-178.

105. Theocharis, A.D., et al., *Extracellular matrix structure*. Advanced drug delivery reviews, 2016. **97** : p. 4-27.

106. Wei, R., et al., Cellular and extracellular components in tumor microenvironment and their application in early diagnosis of cancers. Analytical Cellular Pathology, 2020. **2020**.

107. Clause, K.C. and T.H. Barker, *Extracellular matrix signaling in morphogenesis and repair*. Current opinion in biotechnology, 2013.24 (5): p. 830-833.

108. Tan, T.-T. and L.M. Coussens, *Humoral immunity, inflammation and cancer.* Current opinion in immunology, 2007. **19** (2): p. 209-216.

109. Liu, H., H. Zhao, and Y. Sun. Tumor microenvironment and cellular senescence: Understanding therapeutic resistance and harnessing strategies. in Seminars in Cancer Biology. 2022. Elsevier.

110. Schuster, R., et al., *The inflammatory speech of fibroblasts*. Immunological reviews, 2021. **302** (1): p. 126-146.

111. Wu, P., et al., Adaptive mechanisms of tumor therapy resistance driven by tumor microenvironment. Frontiers in cell and developmental biology, 2021. **9** : p. 641469.

112. Ni, Y., et al., The role of tumor-stroma interactions in drug resistance within tumor microenvironment. Frontiers in Cell and Developmental Biology, 2021. **9** : p. 637675.

113. Morin, P.J., *Drug resistance and the microenvironment: nature and nurture.* Drug Resistance Updates, 2003. 6 (4): p. 169-172.

114. Chen, Y., et al., Therapeutic remodeling of the tumor microenvironment enhances nanoparticle delivery. Advanced Science, 2019. 6 (5): p. 1802070.

115. Jo, Y., et al., Chemoresistance of cancer cells: requirements of tumor microenvironment-mimicking in vitro models in anti-cancer drug development. Theranostics, 2018. 8 (19): p. 5259.

116. Xu, S., et al., *The role of collagen in cancer: from bench to bedside.* Journal of translational medicine, 2019. **17** : p. 1-22.

117. Natarajan, S., et al., Collagen remodeling in the hypoxic tumor-mesothelial niche promotes ovarian cancer metastasis. Cancer research, 2019. **79** (9): p. 2271-2284.

118. Naito, Y., et al., Micro RNA-143 regulates collagen type III expression in stromal fibroblasts of scirrhous type gastric cancer.Cancer science, 2014. **105** (2): p. 228-235.

119. Mu, W., S. Rana, and M. Zoller, *Host matrix modulation by tumor exosomes promotes motility and invasiveness*. Neoplasia, 2013.15 (8): p. 875-IN4.

120. Mathot, P., et al., *DNA methylation signal has a major role in the response of human breast cancer cells to the microenvironment*.Oncogenesis, 2017. **6** (10): p. e390-e390.

121. Roodhart, J.M., et al., Mesenchymal stem cells induce resistance to chemotherapy through the release of platinum-induced fatty acids. Cancer cell, 2011. **20** (3): p. 370-383.

122. Han, Z., et al., Mesenchymal stem cells contribute to the chemoresistance of hepatocellular carcinoma cells in inflammatory environment by inducing autophagy. Cell & bioscience, 2014.4 (1): p. 1-11.

123. Ji, R., et al., Exosomes derived from human mesenchymal stem cells confer drug resistance in gastric cancer. Cell cycle, 2015.14 (15): p. 2473-2483.

124. Xu, H., et al., Tumor-derived mesenchymal-stem-cell-secreted IL-6 enhances resistance to cisplatin via the STAT3 pathway in breast cancer. Oncology Letters, 2018. **15** (6): p. 9142-9150.

125. Lu, M., et al., Notoginsenoside R1 counteracts mesenchymal stem cell-evoked oncogenesis and doxorubicin resistance in osteosarcoma cells by blocking IL-6 secretion-induced JAK2/STAT3 signaling.Investigational New Drugs, 2021. **39**: p. 416-425.

126. Raghavan, S., et al., Carcinoma-associated mesenchymal stem cells promote chemoresistance in ovarian cancer stem cells via PDGF signaling. Cancers, 2020. 12 (8): p. 2063.

127. Wang, S., et al., The CXCR4 antagonist, AMD3100, reverses mesenchymal stem cell-mediated drug resistance in relapsed/refractory acute lymphoblastic leukemia. OncoTargets and therapy, 2020.13 : p. 6583.

128. Ridge, S.M., F.J. Sullivan, and S.A. Glynn, *Mesenchymal stem cells: key players in cancer progression*. Molecular cancer, 2017.16 (1): p. 1-10.

129. Wang, J., et al., Cell adhesion-mediated mitochondria transfer contributes to mesenchymal stem cellinduced chemoresistance on T cell acute lymphoblastic leukemia cells. Journal of Hematology & Oncology, 2018. **11** (1): p. 1-13.

130. Shi, Y., et al., *Tumour-associated mesenchymal stem/stromal cells: emerging therapeutic targets*. Nature reviews Drug discovery, 2017. **16** (1): p. 35-52.

131. Scherzed, A., et al., *BMSC enhance the survival of paclitaxel treated squamous cell carcinoma cells in vitro*. Cancer Biology & Therapy, 2011. **11** (3): p. 349-357.

132. Luo, J., et al., Infiltrating bone marrow mesenchymal stem cells increase prostate cancer stem cell population and metastatic ability via secreting cytokines to suppress androgen receptor signaling. Oncogene, 2014. **33** (21): p. 2768-2778.

133. Adamo, A., et al., Role of mesenchymal stromal cell-derived extracellular vesicles in tumour microenvironment. Biochimica et Biophysica Acta (BBA)-Reviews on Cancer, 2019. **1871** (1): p. 192-198. 134. Wu, Q., et al., Unraveling adipocytes and Cancer links: is there a role for senescence? Frontiers in cell and developmental biology, 2020. 8 : p. 282.

135. Cao, Y., *Adipocyte and lipid metabolism in cancer drug resistance*. The Journal of clinical investigation, 2019.**129** (8): p. 3006-3017.

136. Nieman, K.M., et al., Adipose tissue and adipocytes support tumorigenesis and metastasis. Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids, 2013. **1831** (10): p. 1533-1541.

137. Deng, T., et al., *Obesity, inflammation, and cancer.* Annual Review of Pathology: Mechanisms of Disease, 2016. **11** : p. 421-449.

138. Elaraby, E., et al., Natural Killer Cell Dysfunction in Obese Patients with Breast Cancer: A Review of a Triad and Its Implications. Journal of Immunology Research, 2021. 2021.

139. Duong, M.N., et al., The fat and the bad: Mature adipocytes, key actors in tumor progression and resistance. Oncotarget, 2017.8 (34): p. 57622.

140. Yu, W., et al., Adipocytes secreted leptin is a pro-tumor factor for survival of multiple myeloma under chemotherapy. Oncotarget, 2016. 7 (52): p. 86075.

141. Satoh, M., et al., Modulation of resistance to anticancer drugs by inhibition of metallothionein synthesis. Cancer research, 1994. **54** (20): p. 5255-5257.

142. Choi, J., Y.J. Cha, and J.S. Koo, Adipocyte biology in breast cancer: From silent bystander to active facilitator. Progress in lipid research, 2018. **69** : p. 11-20.

143. Iyengar, P., et al., Adipocyte-derived collagen VI affects early mammary tumor progression in vivo, demonstrating a critical interaction in the tumor/stroma microenvironment. The Journal of clinical investigation, 2005. **115** (5): p. 1163-1176.

144. Sherman-Baust, C.A., et al., Remodeling of the extracellular matrix through overexpression of collagen VI contributes to cisplatin resistance in ovarian cancer cells. Cancer cell, 2003. **3** (4): p. 377-386.

145. Perez de Heredia, F., I.S. Wood, and P. Trayhurn, *Hypoxia stimulates lactate release and modulates monocarboxylate transporter (MCT1, MCT2, and MCT4) expression in human adipocytes.* Pflugers Archiv-European Journal of Physiology, 2010. **459** : p. 509-518.

146. Yang, E., et al., *Exosome-mediated metabolic reprogramming: the emerging role in tumor microenvironment remodeling and its influence on cancer progression.* Signal transduction and targeted therapy, 2020.5 (1): p. 242.

147. Sheng, X., et al., Adipocytes sequester and metabolize the chemotherapeutic daunorubicin. Molecular Cancer Research, 2017.15 (12): p. 1704-1713.

148. Arneth, B., Tumor microenvironment. Medicina, 2019.56 (1): p. 15.

149. Hulikova, A. and P. Swietach, *Rapid CO2 permeation across biological membranes: implications for CO2 venting from tissue.* The FASEB Journal, 2014. **28** (7): p. 2762-2774.

150. Vaupel, P. and A. Mayer, *Hypoxia in tumors: pathogenesis-related classification, characterization of hypoxia subtypes, and associated biological and clinical implications.* Oxygen transport to tissue XXXVI, 2014: p. 19-24.

151. Rohani, N., et al., Acidification of tumor at stromal boundaries drives transcriptome alterations associated with aggressive phenotypes. Cancer research, 2019. **79** (8): p. 1952-1966.

152. Peppicelli, S., et al., *The acidic microenvironment as a possible niche of dormant tumor cells*. Cellular and molecular life sciences, 2017. **74** : p. 2761-2771.

153. Hanahan, D. and R.A. Weinberg, *Hallmarks of cancer: the next generation*. cell, 2011. **144** (5): p. 646-674.

154. Siska, P.J., et al., The immunological Warburg effect: Can a metabolic-tumor-stroma score (MeTS) guide cancer immunotherapy?Immunological reviews, 2020. **295** (1): p. 187-202.

155. Apicella, M., et al., Increased lactate secretion by cancer cells sustains non-cell-autonomous adaptive resistance to MET and EGFR targeted therapies. Cell metabolism, 2018. **28** (6): p. 848-865. e6.

156. Yoshida, G.J., Metabolic reprogramming: the emerging concept and associated therapeutic strategies. Journal of experimental & clinical cancer research, 2015. **34** : p. 1-10.

157. Martinez-Outschoorn, U.E., et al., *Cancer metabolism: a therapeutic perspective*. Nature reviews Clinical oncology, 2017.14 (1): p. 11-31.

158. Gupta, S., A. Roy, and B.S. Dwarakanath, *Metabolic cooperation and competition in the tumor microenvironment: implications for therapy.* Frontiers in oncology, 2017. 7: p. 68.

159. Wang, Q., et al., Role of tumor microenvironment in cancer progression and therapeutic strategy. Cancer Medicine, 2023.

160. Libutti, S.K., L. Tamarkin, and N. Nilubol, *Targeting the invincible barrier for drug delivery in solid cancers: interstitial fluid pressure.* Oncotarget, 2018. 9 (87): p. 35723.

161. Yu, T., et al., *High interstitial fluid pressure promotes tumor progression through inducing lymphatic metastasis-related protein expressions in oral squamous cell carcinoma.* Clinical and Translational Oncology, 2014. **16** : p. 539-547.

162. Haider, T., et al., *Recent advances in tumor microenvironment associated therapeutic strategies and evaluation models.* Materials Science and Engineering: C, 2020. **116** : p. 111229.

163. Tredan, O., et al., *Drug resistance and the solid tumor microenvironment*. Journal of the National Cancer Institute, 2007.99 (19): p. 1441-1454.

164. Chen, F., et al., New horizons in tumor microenvironment biology: challenges and opportunities. BMC medicine, 2015.13 (1): p. 1-14.

165. Wang, S., et al., Tumor microenvironment in chemoresistance, metastasis and immunotherapy of pancreatic cancer. American journal of cancer research, 2020. **10** (7): p. 1937.

166. Romano, A., et al., *PMN-MDSC and arginase are increased in myeloma and may contribute to resistance to therapy.* Expert review of molecular diagnostics, 2018. **18** (7): p. 675-683.

167. Calcinotto, A., et al., *IL-23 secreted by myeloid cells drives castration-resistant prostate cancer*. Nature, 2018.559 (7714): p. 363-369.

168. Dzobo, K., D.A. Senthebane, and C. Dandara, *The tumor microenvironment in tumorigenesis and therapy resistance revisited*.Cancers, 2023. **15** (2): p. 376.

169. Maimela, N.R., S. Liu, and Y. Zhang, *Fates of CD8+ T cells in tumor microenvironment*. Computational and structural biotechnology journal, 2019. **17** : p. 1-13.

170. Bamias, A., et al., δρρελατιον οφ NK T-λικε  $\Delta 3+ \Delta 56+ \varsigma ελλς$  ανδ  $\Delta 4+ \Delta 25+ (ηι)$  ρεγυλατορψ T ςελλς ωιτη ΈΓΦ ανδ TNΦa ιν ασςιτες φρομ αδανςεδ οαριαν ςανςερ: Ασσοςιατιον ωιτη πλατινυμ ρεσιστανςε ανδ προγνοσις ιν πατιεντς ρεςειινγ φιρστ-λινε, πλατινυμ-βασεδ ςηεμοτηεραπψ. Gynecologic oncology, 2008. **108** (2): p. 421-427.

171. Long, Y., et al., Dysregulation of glutamate transport enhances treg function that promotes VEGF blockade resistance in glioblastoma. Cancer research, 2020. **80** (3): p. 499-509.

172. Imbert, C., et al., Resistance of melanoma to immune checkpoint inhibitors is overcome by targeting the sphingosine kinase-1. Nature communications, 2020. **11** (1): p. 437.

173. Li, C., et al., *Foxp3 overexpression decreases sensitivity to chemotherapy in mouse Lewis lung cancer cells.* Molecular medicine reports, 2012. 6 (5): p. 977-982.

174. Wang, D., et al.,  $\delta \lambda \rho \epsilon \varsigma \tau a \lambda \varsigma a \nu \varsigma \epsilon \rho \varsigma \epsilon \lambda \lambda - \delta \epsilon \rho \iota \epsilon \delta \ ^{\infty}\Lambda 20 \rho \epsilon \varsigma \rho \upsilon \tau \varsigma \rho \epsilon \gamma \upsilon \lambda a \tau o \rho \psi T \varsigma \epsilon \lambda \lambda \varsigma \tau o \pi \rho \rho \mu \sigma \tau \epsilon \varsigma \eta \epsilon \mu o \rho \epsilon - \sigma \sigma \tau a \nu \varsigma \epsilon \iota a \Phi O = O 1 \ ^{\infty} E B \Pi B / N \Phi - \kappa B \sigma \eta \nu a \lambda \iota \nu \gamma$ . Journal for immunotherapy of cancer, 2019.7 (1): p. 1-15.

175. Schuler, P.J., et al., Effects of Adjuvant Chemoradiotherapy on the Frequency and Function of Regulatory T Cells in Patients with Head and Neck CancerCD4+ CD39+ Treg and CRT. Clinical cancer research, 2013. **19** (23): p. 6585-6596.

176. Liu, X.-D., et al., Resistance to Antiangiogenic Therapy Is Associated with an Immunosuppressive Tumor Microenvironment in Metastatic Renal Cell CarcinomaAntiangiogenic Therapy Increases PD-L1 Expression. Cancer immunology research, 2015. **3** (9): p. 1017-1029.

177. Aspord, C., et al., Breast cancer instructs dendritic cells to prime interleukin 13-secreting CD4+T cells that facilitate tumor development. The Journal of experimental medicine, 2007.204 (5): p. 1037-1047.

178. Vicari, A.P., C. Caux, and G. Trinchieri. *Tumour escape from immune surveillance through dendritic cell inactivation*. in *Seminars in cancer biology*. 2002. Elsevier.

179. Incio, J., et al., Obesity-induced inflammation and desmoplasia promote pancreatic cancer progression and resistance to chemotherapy. Cancer discovery, 2016. 6 (8): p. 852-869.

180. Shaul, M.E. and Z.G. Fridlender, *Tumour-associated neutrophils in patients with cancer*. Nature reviews Clinical oncology, 2019. **16** (10): p. 601-620.

181. Bui, T.M., L.K. Yalom, and R. Sumagin, *Tumor-associated neutrophils: orchestrating cancer pathobiology and therapeutic resistance.* Expert opinion on therapeutic targets, 2021.25 (7): p. 573-583.

182. Chong, A.S.-F., et al., Diverse multidrug-resistance-modification agents inhibit cytolytic activity of natural killer cells. Cancer Immunology, Immunotherapy, 1993.36 : p. 133-139.

183. Savas, B., et al., *P-glycoprotein-mediated multidrug resistance and cytotoxic effector cells*. Nat Immun, 1992.11 (4): p. 177-92.

184. Takahashi, M., et al., Role of P-glycoprotein in human natural killer-like cell line-mediated cytotoxicity. Experimental Cell Research, 1999. **253** (2): p. 396-402.

185. Sugimura, K., et al., *High infiltration of tumor-associated macrophages is associated with a poor response to chemotherapy and poor prognosis of patients undergoing neoadjuvant chemotherapy for esophageal cancer.* Journal of surgical oncology, 2015. **111** (6): p. 752-759.

186. Li, X., et al., *Harnessing tumor-associated macrophages as aids for cancer immunotherapy*. Molecular Cancer, 2019. **18** (1): p. 1-16.

187. Liu, Y. and X. Cao, *The origin and function of tumor-associated macrophages*. Cellular & molecular immunology, 2015.12 (1): p. 1-4.

188. Moradi-Chaleshtori, M., et al., In vitro and in vivo evaluation of anti-tumoral effect of M1 phenotype induction in macrophages by miR-130 and miR-33 containing exosomes. Cancer Immunology, Immunotherapy, 2021. **70**: p. 1323-1339.

189. Huang, W.-C., et al., Cisplatin resistant lung cancer cells promoted M2 polarization of tumor-associated macrophages via the Src/CD155/MIF functional pathway. Journal of Experimental & Clinical Cancer Research, 2019. **38** (1): p. 1-17.

190. Liu, H., et al., Jagged1 promotes aromatase inhibitor resistance by modulating tumor-associated macrophage differentiation in breast cancer patients. Breast Cancer Research and Treatment, 2017.166 : p. 95-107.

191. Yu, S., et al., Αςτιατεδ ΗΙΦ1α οφ τυμορ ςελλς προμοτες ςηεμορεσιστανςε δεελοπμεντ ια ρεςρυιτινγ ΓΔΦ15προδυςινγ τυμορ-ασσοςιατεδ μαςροπηαγες ιν γαστρις ςανςερ. Cancer Immunology, Immunotherapy, 2020. 69
: p. 1973-1987.

192. Amit, M. and Z. Gil, Macrophages increase the resistance of pancreatic adenocarcinoma cells to gemcitabine by upregulating cytidine deaminase. Oncoimmunology, 2013. 2 (12): p. e27231.

193. Weizman, N., et al., Macrophages mediate gemcitabine resistance of pancreatic adenocarcinoma by upregulating cytidine deaminase. Oncogene, 2014. **33** (29): p. 3812-3819.

194. Binenbaum, Y., et al., Transfer of miRNA in Macrophage-Derived Exosomes Induces Drug Resistance in Pancreatic AdenocarcinomaExosomes Induce Gemcitabine Resistance in Pancreatic Cancer. Cancer research, 2018. **78** (18): p. 5287-5299.

195. Xian, G., et al.,  $\Sigma$ iµaστατιν αττενυατες µaςροπηαγε-µεδιατεδ γεµςιταβινε ρεσιστανςε οφ πανςρεατις δυςταλ αδενος αρςινομα βψ ρεγυλατιν τηε  $T\Gamma \Phi$ -β1/ $\Gamma \varphi$ i-1 aξις. Cancer letters, 2017. **385** : p. 65-74.

196. Ireland, L., et al., Chemoresistance in Pancreatic Cancer Is Driven by Stroma-Derived Insulin-Like Growth FactorsStroma-Derived IGFs Enhance Chemoresistance in PDAC. Cancer research, 2016.76 (23): p. 6851-6863.

197. Kuwada, K., et al., The epithelial-to-mesenchymal transition induced by tumor-associated macrophages confers chemoresistance in peritoneally disseminated pancreatic cancer. Journal of Experimental & Clinical Cancer Research, 2018. **37**: p. 1-10.

198. Huang, H., et al., Reciprocal Network between Cancer Stem-Like Cells and Macrophages Facilitates the Progression and Androgen Deprivation Therapy Resistance of Prostate CancerCSC-Mø Facilitates the Progression and ADT Resistance of PCa. Clinical cancer research, 2018.24 (18): p. 4612-4626.

199. Torgovnick, A. and B. Schumacher, *DNA repair mechanisms in cancer development and therapy*. Frontiers in genetics, 2015.6 : p. 157.

200. Wang, M., S. Chen, and D. Ao, Targeting DNA repair pathway in cancer: Mechanisms and clinical application. MedComm, 2021.2 (4): p. 654-691.

201. Kiwerska, K. and K. Szyfter, DNA repair in cancer initiation, progression, and therapy—a double-edged sword. Journal of applied genetics, 2019. **60** (3-4): p. 329-334.

202. Goldstein, M. and M.B. Kastan, *The DNA damage response: implications for tumor responses to radiation and chemotherapy.* Annual review of medicine, 2015. **66** : p. 129-143.

203. Abbotts, R., N. Thompson, and S. Madhusudan, *DNA repair in cancer: emerging targets for personalized therapy*. Cancer management and research, 2014: p. 77-92.

204. Huang, R. and P.-K. Zhou, DNA damage repair: Historical perspectives, mechanistic pathways and clinical translation for targeted cancer therapy. Signal Transduction and Targeted Therapy, 2021.6 (1): p. 254.

205. Blasiak, J., *Single-strand annealing in cancer*.International Journal of Molecular Sciences, 2021. **22** (4): p. 2167.

206. Jackson, S.P. and J. Bartek, *The DNA-damage response in human biology and disease*. Nature, 2009. **461** (7267): p. 1071-1078.

207. Jurkovicova, D., et al., DNA Damage Response in Cancer Therapy and Resistance: Challenges and Opportunities. International Journal of Molecular Sciences, 2022. 23 (23): p. 14672.

208. Czerninski, R., et al., Promoter hypermethylation of mismatch repair genes, hMLH1 and hMSH2 in oral squamous cell carcinoma. Oral diseases, 2009. 15 (3): p. 206-213.

209. Guan, H., et al., Hypermethylation of the DNA mismatch repair gene hMLH1 and its association with lymph node metastasis and T1799A BRAF mutation in patients with papillary thyroid cancer. Cancer, 2008.113 (2): p. 247-255.

210. Wang, Y.-C., et al., Inactivation of hMLH1 and hMSH2 by promoter methylation in primary non-small cell lung tumors and matched sputum samples. The Journal of clinical investigation, 2003.111 (6): p. 887-895.

211. Lee, M.-N., et al., *Epigenetic inactivation of the chromosomal stability control genes BRCA1, BRCA2, and XRCC5 in non-small cell lung cancer.* Clinical cancer research, 2007.13 (3): p. 832-838.

212. Liu, K., et al., Promoter hypermethylation: an important epigenetic mechanism for hMLH1 gene inactivation in head and neck squamous cell carcinoma. Otolaryngology-Head and Neck Surgery, 2002.126 (5): p. 548-553.

213. Fleisher, A.S., et al., Hypermethylation of the hMLH1 gene promoter is associated with microsatellite instability in early human gastric neoplasia. Oncogene, 2001. **20** (3): p. 329-335.

214. Bernal, C., et al., DNA methylation profile in diffuse type gastric cancer: evidence for hypermethylation of the BRCA1 promoter region in early-onset gastric carcinogenesis. Biological research, 2008. **41** (3): p. 303-315.

215. Seedhouse, C., E. Das-Gupta, and N. Russell, Methylation of the hMLH1 promoter and its association with microsatellite instability in acute myeloid leukemia. Leukemia, 2003. 17 (1): p. 83-88.

216. Zhang, L. and X. Long, Association of BRCA1 promoter methylation with sporadic breast cancers: Evidence from 40 studies. Scientific reports, 2015. 5 (1): p. 17869.

217. Gras, E., et al., Loss of heterozygosity on chromosome 13q12-q14, BRCA-2 mutations and lack of BRCA-2 promoter hypermethylation in sporadic epithelial ovarian tumors. Cancer, 2001.92 (4): p. 787-795.

218. Yu, J., et al., A novel set of DNA methylation markers in urine sediments for sensitive/specific detection of bladder cancer. Clinical cancer research, 2007. **13** (24): p. 7296-7304.

219. Fojo, T., Cancer, DNA repair mechanisms, and resistance to chemotherapy . 2001, Oxford University Press. p. 1434-1436.

220. Phi, L.T.H., et al., Cancer stem cells (CSCs) in drug resistance and their therapeutic implications in cancer treatment. Stem cells international, 2018. **2018**.

221. Papaccio, F., et al., Concise review: cancer cells, cancer stem cells, and mesenchymal stem cells: influence in cancer development. Stem cells translational medicine, 2017. 6 (12): p. 2115-2125.

222. Aramini, B., et al., Dissecting tumor growth: the role of cancer stem cells in drug resistance and recurrence. Cancers, 2022.14 (4): p. 976.

223. Pattabiraman, D.R. and R.A. Weinberg, *Tackling the cancer stem cells—what challenges do they pose?* Nature reviews Drug discovery, 2014. **13** (7): p. 497-512.

224. Yang, Z.-J. and R.J. Wechsler-Reya, *Hit'em where they live: targeting the cancer stem cell niche*. Cancer cell, 2007.11 (1): p. 3-5.

225. Gaggianesi, M., et al., Messing up the cancer stem cell chemoresistance mechanisms supported by tumor microenvironment. Frontiers in Oncology, 2021: p. 2847.

226. Li, Y., et al., *Drug resistance and Cancer stem cells*. Cell Communication and Signaling, 2021. **19** (1): p. 1-11.

227. Prieto-Vila, M., et al., *Drug resistance driven by cancer stem cells and their niche*. International journal of molecular sciences, 2017. **18** (12): p. 2574.

228. Barbato, L., et al., Cancer stem cells and targeting strategies. Cells, 2019. 8 (8): p. 926.

229. Liang, L. and A.M. Kaufmann, *The Significance of Cancer Stem Cells and Epithelial–Mesenchymal Transition in Metastasis and Anti-Cancer Therapy.* International Journal of Molecular Sciences, 2023.24 (3): p. 2555.

230. Agliano, A., A. Calvo, and C. Box. The challenge of targeting cancer stem cells to halt metastasis . in Seminars in Cancer Biology . 2017. Elsevier.

231. Knoll, S., S. Emmrich, and B.M. Putzer, *The E2F1-miRNA cancer progression network*. MicroRNA Cancer Regulation: Advanced Concepts, Bioinformatics and Systems Biology Tools, 2013: p. 135-147.

232. Dar, A.A., et al., miRNA-205 suppresses melanoma cell proliferation and induces senescence via regulation of E2F1 protein. Journal of Biological Chemistry, 2011. 286 (19): p. 16606-16614.

233. Gregory, P.A., et al., The miR-200 family and miR-205 regulate epithelial to mesenchymal transition by targeting ZEB1 and SIP1. Nature cell biology, 2008. **10** (5): p. 593-601.

234. Godlewski, J., et al., Targeting of the Bmi-1 oncogene/stem cell renewal factor by microRNA-128 inhibits glioma proliferation and self-renewal. Cancer research, 2008. 68 (22): p. 9125-9130.

235. Deng, J., et al., Long non-coding RNA HOTAIR regulates the proliferation, self-renewal capacity, tumor formation and migration of the cancer stem-like cell (CSC) subpopulation enriched from breast cancer cells. PLoS One, 2017. **12** (1): p. e0170860.

236. O'Brien, C.A., et al., *ID1 and ID3 regulate the self-renewal capacity of human colon cancer-initiating cells through p21.* Cancer cell, 2012. **21** (6): p. 777-792.

237. Nahand, J.S., et al., *microRNAs: new prognostic, diagnostic, and therapeutic biomarkers in cervical cancer.* Journal of cellular physiology, 2019. **234** (10): p. 17064-17099.

238. Naeli, P., et al., Circular RNAs and gastrointestinal cancers: epigenetic regulators with a prognostic and therapeutic role.Critical reviews in oncology/hematology, 2020. 145 : p. 102854.

239. Yan, H. and P. Bu, Non-coding RNAs in cancer stem cells.Cancer letters, 2018. 421 : p. 121-126.

240. Davis, B.N. and A. Hata, *Regulation of MicroRNA Biogenesis: A miRiad of mechanisms*. Cell Communication and Signaling, 2009.7: p. 1-22.