Scale matters: genome-wide signatures of local adaptation to high-resolution environmental variation in an alpine plant

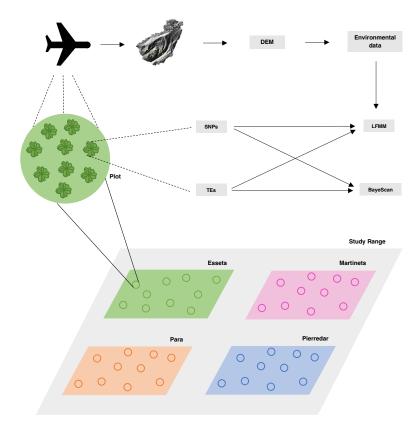
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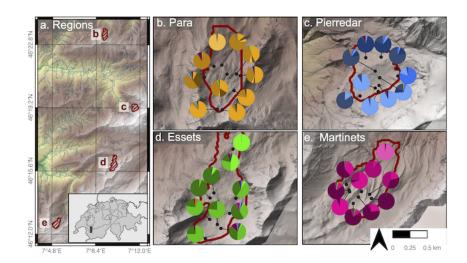
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Abstract

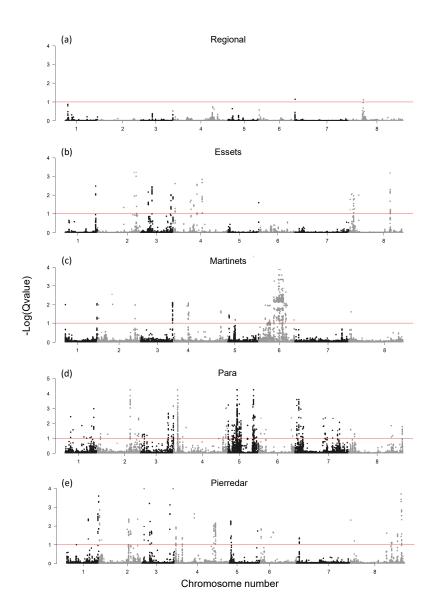
Microevolutionary processes shape adaptive responses to heterogeneous environments, where these effects vary both among and within species. However, the degree to which signatures of adaptation to environmental drivers can be detected based on spatial scale and genomic marker remains largely unknown. We studied signatures of local adaptation across different spatial extents, investigating complementary types of genomic variants-single nucleotide polymorphisms (SNPs) and polymorphic transposable elements (TEs)-in populations of the alpine model plant species Arabis alpina. We coupled high-resolution (0.5m) environmental factors, derived from remote sensing digital elevation models, with whole-genome sequenced data of 304 individuals across four populations. We demonstrate that responses of A. alpina to similar amounts of abiotic variation are largely governed by local evolutionary processes and find minimally overlapping signatures of local adaptation between SNPs and polymorphic TEs. Notably, functional annotations of high-impact genomic variants revealed several defence-related genes associated with the abiotic factors studied, which could indicate indirect selective pressure of biotic agents. Our results highlight the importance of considering different spatial extents and types of genomic polymorphisms when searching for signatures of adaptation to environmental variation. Such insights provide key information on microevolutionary processes and could guide management decisions to mitigate negative impacts of climate change on alpine plant populations.

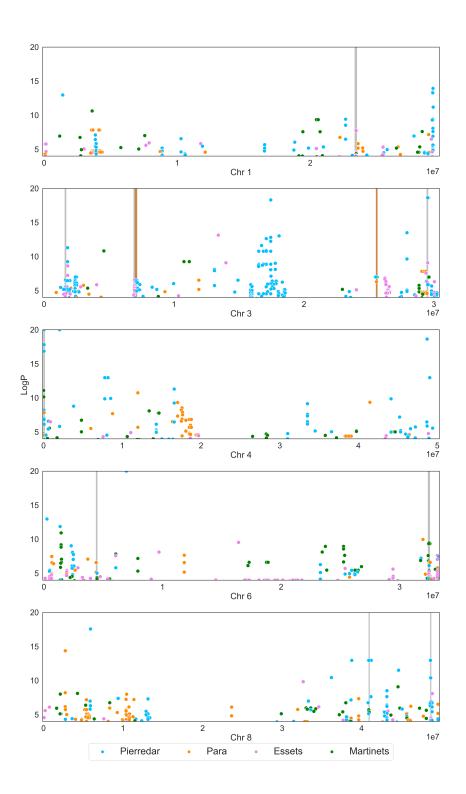




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Scale matters: genome-wide signatures of local adaptation to

2 high-resolution environmental variation in an alpine plant

3 Running title Scale effects in local adaptation

4

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27 Abstract

28 Microevolutionary processes shape adaptive responses to heterogeneous

29 environments, where these effects vary both among and within species. However, it 30 remains largely unknown to which degree signatures of adaptation to environmental 31 drivers can be detected based on the choice of spatial scale and genomic marker. We 32 studied signatures of local adaptation across different levels of spatial extents, 33 investigating complementary types of genomic variants-single nucleotide 34 polymorphisms (SNPs) and polymorphic transposable elements (TEs)-in populations 35 of the alpine model plant species Arabis alpina. We coupled environmental factors, 36 derived from remote sensing digital elevation models at very high resolution (0.5m), 37 with whole-genome sequencing data of 304 individuals across four populations. We 38 demonstrate that responses of *A. alpina* to similar amounts of abiotic variation are 39 largely governed by local evolutionary processes, and we find minimally overlapping 40 signatures of local adaptation between SNPs and polymorphic TEs. Notably, 41 functional annotations of high-impact genomic variants revealed several defence-42 related genes associated with the abiotic factors studied, which could indicate 43 indirect selective pressure of biotic agents. Our results highlight the importance of 44 considering different spatial extents and types of genomic polymorphisms when 45 searching for signatures of adaptation to environmental variation. Such insights 46 provide key information on microevolutionary processes and could guide 47 management decisions to mitigate negative impacts of climate change on alpine plant 48 populations.

49

50 **Key words:** *Arabis alpina*, genomic variation, genotype–environment associations,

51 high-resolution environmental variation, local adaptation, remote sensing

52 Introduction

53 Local adaptation enables populations to evolve traits that confer a fitness advantage 54 in their respective habitats (Kawecki & Ebert, 2004). Genomic variation may reveal 55 signatures of such adaptive processes, but identifying the underlying environmental 56 drivers and relevant spatial scale remains challenging (Savolainen et al., 2013). To 57 improve our understanding of traits under selection and their underlying functional 58 processes, one may decipher the imprints of local adaptation based on whole-59 genome patterns of variation. However, to our knowledge, fine-grained sampling of 60 individuals and environmental data, at a resolution matching genomic variation, has 61 never been used to assess environmental drivers of adaptation.

Studies investigating local adaptation often cover broad spatial extents
(commonly referred to as scales), involving continent-wide sampling for example
(Gougherty et al., 2021; Pais et al., 2017; Yeaman et al., 2016), but see (de
Villemereuil et al., 2018; Eckert et al., 2015; Fischer et al., 2013). However, adaptation
can be truly local (Rellstab et al., 2017), particularly in highly heterogeneous alpine
landscapes where environmental conditions vary over short distances (Rellstab et al.,
2020).

69 In their local study on the alpine Brassicaceae *Biscutella laevigata*, Leempoel 70 et al. (2018) highlight the importance of considering the ratio between grain (spatial 71 resolution) and extent of a study area to assess signatures of adaptation, which 72 should match the spatial context at which selection operates (Dauphin et al., 2023). 73 In rugged alpine terrain, an organism's habitat can be adequately described using 74 ecological data derived from digital elevation models (DEM; Kozak et al., 2008), which 75 correlate significantly with climatic factors measured using ground-based sensors 76 (Leempoel et al., 2015)(Pradervand et al., 2014). Such topography-derived 77 environmental descriptors provide continuous information across habitats, reflecting 78 long-term patterns of local site conditions, as compared to point-based 79 measurements acquired on-site or interpolated from local weather stations, which 80 provide short-term and often patchy environmental information. 81 Whole-genome studies of adaptation in natural plant populations mostly

82 remain restricted to model organisms like Arabidopsis thaliana (Exposito-Alonso et

83 al., 2018; Fournier-Level et al., 2011; Hancock et al., 2011) or crop wild relatives (e.g., 84 Yoder et al., 2014). Additionally, they are often based on pooled sequencing (Fischer 85 et al., 2013; Rellstab et al., 2020; Turner et al., 2010), or only consider a few 86 individuals that do not necessarily represent the local populations (Laenen et al., 87 2018). Moreover, structural variants, often involved in adaptation beyond what can 88 be captured by commonly analysed single-nucleotide polymorphisms (SNPs), have 89 been largely ignored in these studies. Transposable elements (TEs), for example, 90 represent a major fraction of many plant genomes (Bennetzen & Wang, 2014; 91 Quesneville, 2020) and are known to induce adaptive responses to abiotic stress 92 (Baduel et al., 2021; Kalendar et al., 2000), yet have remained largely overlooked in 93 studies of local adaptation. Indeed, biallelic SNPs and TEs are expected to have 94 complementary effects on the adaptive responses of plants to local conditions. For 95 example, polymorphic TEs are known to affect recombination along chromosomes, 96 such that they may promote entire blocks of linked loci to reveal signatures of 97 selection along chromosomes (Choudhury et al., 2019), while genomic variation in 98 non-synonymous SNPs may induce rather subtle changes in the respective proteins. 99 Therefore, it is expected that these marker types identify few common, but rather 100 complementary signals of local adaptation.

101 It is commonly assumed that similar environmental contrasts across sites 102 leave consistent signatures of selection in plants with a common genomic background 103 (Booker et al., 2021; Lotterhos & Whitlock, 2015) (Rellstab et al., 2020). However, for 104 populations with limited gene flow, particularly in species with mixed mating systems 105 or selfing, one may expect population-specific adaptive responses with limited 106 convergence (Rellstab et al., 2017; 2020).

107 Arabis alpina (Brassicaceae) has recently emerged as a model species for 108 ecological genomics (Wötzel et al., 2022). Studies have provided empirical evidence 109 of how local adaptation acts on the species' genome (Laenen et al., 2018; Lobréaux & 110 Miguel, 2020; e.g., Poncet et al., 2010), with notable effects on phenotypes (Chopra 111 et al., 2019; de Villemereuil et al., 2018; Vayssières et al., 2020; Wang et al., 2009). As 112 detailed in (Wötzel et al., 2022), several field studies, including reciprocal transplant 113 experiments, substantiate the potential of A. alpina to adapt on small spatial scales, 114 e.g., in response to elevational differences (Buckley et al., 2019; de Villemereuil et al.,

2018; Wingler et al., 2015). Due to this species' responsiveness to environmental
conditions, and the availability of a high-quality reference genome, it is an intriguing
organism to further investigate whether signatures of selection to fine-scale
environmental drivers are found across the whole genome.

119 Here, we re-sequenced 304 individuals of *A. alpina*, sampled in local 120 populations with a common history, using a spatially explicit, hierarchical design 121 (individuals in plots nested within four local populations, Figures 1 and 2). We 122 computed the environmental factors at a very high resolution (0.5 m) to describe the 123 local habitat for each individual. Through associations with genomic variation, we 124 inferred drivers of local adaptation (i) at the regional level, i.e., among the four local 125 populations, and (ii) at the local level, i.e., among plots within each local population. 126 In parallel, we performed outlier detection analyses to identify possible genomic 127 signatures of local adaptation irrespective of underlying environmental drivers. We 128 focused on whether signals of adaptation between regional and local spatial extents, 129 as well as between types of genomic markers (SNPs vs. TEs), show common or 130 different patterns and functions, and to what degree patterns of local adaptation are

132 Materials and Methods

131

133 Study species and sampling

134 Arabis alpina L. has a broad ecological niche covering a large altitudinal and

driven by particular environmental factors at different spatial extents.

135 latitudinal range, making it suitable for investigating adaptation to variation across

- 136 heterogeneous environments (Wötzel et al., 2022). This perennial Brassicaceae is
- 137 found throughout the Northern Hemisphere, predominantly on calcareous bedrock.
- 138 In the European Alps, A. alpina is found across an elevation range of 400 m to 3200 m
- a.s.l. This species reproduces sexually, predominantly via selfing (Ansell et al., 2008;
- 140 Buehler et al., 2012; Laenen et al., 2018; Tedder et al., 2011), though outcrossing
- 141 populations can be found in parts of its range (e.g., Ansell et al., 2008; Tedder et al.,
- 142 2011). SNP-based estimates of individual inbreeding coefficients (F), as well as
- 143 positive *F*_{IS} values for both SNPs and polymorphic TEs, suggest substantial inbreeding

in the studied populations, likely due to extensive selfing (Choudhury et al., 2019;Rogivue et al., 2019b).

146 We sampled four local populations of A. alpina from the western Swiss Alps in 147 summer 2014 (Figure 1 and 2), from temperate alpine climates with generally 148 abundant rain and snowfall (Drake et al., 2006). Individuals were sampled 149 predominantly from calcareous bedrock in the alpine vegetation zone above 2000 m 150 a.s.l. in the regions Para (N = 69), Pierredar (N = 69), Essets (N = 70) and Martinets 151 (N = 96) (Table S1; see also Rogivue et al., 2019b). The four studied populations are 152 considered descendants from the same refugial population and share a similar 153 genomic background (Rogivue et al., 2018). Each population is situated within a 154 different catchment valley (Figure 1a), presumably preventing regular gene flow 155 between these populations. However, mating within each population can occur over 156 large distances, as insects may transport pollen up to 1 km (Buehler et al., 2012) and 157 their small, light seeds can be disseminated by wind. Within each population, we 158 selected ten plots from contrasting habitat types to encompass large ecological 159 gradients, within which we sampled the leaves of six to ten plants at least 1 m apart 160 to avoid sampling families (Figure 1 and Table S1). We stored the samples in silica gel 161 and extracted DNA as described elsewhere (Rogivue et al., 2019b). The precise 162 geographic coordinates of each individual (± 2 cm) were obtained with a differential 163 global positioning system (DGPS) receiver, using information provided by Real Time 164 Kinematik (RTK) Global Navigation Satellite System (GNSS) (Table S1).

165

166 Environmental factors

167 Using SAGA GIS (Conrad et al., 2015), we computed 13 topography-derived 168 environmental factors (Table S2) from a precise and high-resolution DEM at 0.5 m 169 pixel resolution, based on light detection and ranging (LiDAR) data obtained from the 170 regional authorities (Canton Vaud, Switzerland). These topographic factors are known 171 proxies for ecologically relevant descriptors, including elevation (and therefore 172 temperature), climate, hydrology, soil conditions, light availability and exposure 173 (Guillaume et al., 2021; Lecours et al., 2017; Leempoel et al., 2015; Wilson & Gallant, 174 2000).

175 Primary terrain attributes including slope, aspect as northness (AspC – cosine 176 of aspect) and eastness (AspS – sine of aspect), and curvature are often used in 177 species distribution models at local and regional scales, as they underlie multiple 178 biophysical processes (such as water flow, snow movement, erosion and solar 179 radiation (Guisan & Zimmermann, 2000; Lyon et al., 2008; Moore et al., 1991). Slope 180 (Slo) describes the steepness of the terrain and aspect describes its orientation. Both 181 are good proxies for solar radiation and temperature, especially when the 182 surrounding relief is of minor importance (Leempoel et al., 2015). Horizontal 183 curvature (Hcu) affects the convergence and the divergence of the water flow, while 184 vertical curvature (Vcu) describes the acceleration and the deceleration of the water 185 flow (Wilson & Gallant, 2000).

186 Over the last two decades, more elaborate secondary terrain attributes have 187 been developed as proxies to model specific biophysical features. Vector ruggedness 188 measure (VRM) describes the degree of terrain unevenness and distinguishes 189 between rocky and smooth terrain (Sappington et al., 2007). This descriptor 190 correlates with soil moisture and is a surrogate for stony soil in the western Alps 191 (Leempoel et al., 2015). Positive topographic openness (TOP) and negative 192 topographic openness (TON) express the protection of a focal point from the 193 surrounding relief (Yokoyama et al., 2002). These factors are based on the maximum 194 angle found at the zenith (TOP) or nadir (TON) from the point over a defined radius. 195 By describing the terrain convexities and concavities, these two factors reflect soil 196 drainage as well as protection from wind (Doneus, 2013).

197 Other factors have been designed to specifically model hydrological 198 processes. The downslope distance gradient (DDG) quantifies the influence of slope 199 on local water drainage (Hjerdt et al., 2004). The topographic wetness index (TWI) 200 quantifies the topographic control of hydrological processes and is formulated as the 201 logarithm of the ratio between the catchment area and the tangent of slope. It has 202 been used to predict soil pH, snow cover and ambient humidity (Beven & Kirkby, 203 1979; Böhner & Selige, 2006), where negative values were correlated with mean and 204 minimum air temperature, while positive values were correlated with the daily range 205 of air temperature (Leempoel et al., 2015).

206 Light availability and temperature can be described by secondary terrain 207 attributes that account for surrounding terrain. The sky view factor (SVF) expresses 208 the ratio of the radiation received by a planar surface to the radiation emitted by the 209 entire hemispheric environment (Böhner & Antonić, 2009), such that SVF proxies for 210 water and light availability. At a microclimatic level, SVF also describes wind 211 protection. Finally, total insolation (Ti6, direct and diffuse radiation) and direct solar 212 insolation in the growing month of June (Di6, direct radiation) account for sun angle, 213 surrounding relief and the SVF. DEM-derived solar radiation is a direct proxy for light 214 availability affecting photosynthesis rates and indirectly accounts for air and soil 215 temperature, influencing evapotranspiration, snow melt and soil moisture (Tovar-216 Pescador et al., 2006).

As environmental factors often covary, we applied Spearman correlation tests to pairs of factors and retained only one factor of the pair if Spearman's $\rho > |0.8|$. Finally, we performed a principal component analysis (PCA) on the retained 12 standardised environmental factors at the locations of the 304 sampled individuals, using the 'prcomp' function in the *stats* package in R 4.1.0 (R Development Core Team, 2021).

223

224 Genomic data

225 We used genomic data described elsewhere (Rogivue et al., 2019a, b), representing 226 SNPs and polymorphic TEs (presence/absence). In short, we sequenced each 227 individual with Illumina HiSeq2500 (ATLAS Biolabs GmbH, Berlin, Germany; 125-bp 228 paired-end reads). After filtering and mapping (BWA 0.7.12; Li & Durbin, 2010) to the 229 high-quality, chromosome-level reference genome (Jiao et al., 2017), the remaining 230 reads corresponded to an average coverage of 11.7x. We performed SNP calling with 231 FreeBayes 1.0.2 (Garrison & Marth, 2012), with 291,396 biallelic SNPs remaining at 232 the regional extent (across all four sites) after stringent filtering (excluding SNPs 233 within TE sequences; Rogivue et al., 2019b). At the local extent, the number of SNPs 234 retained were 220,214 in Essets, 113,900 in Martinets, 287,261 in Para and 160,322 235 in Pierredar (minor allele frequency < 0.025). To describe the spatial genetic structure 236 of the sampled individuals, we carried out an analysis of model-based ancestry,

implemented in the program Admixture (Alexander et al., 2009), on the retained SNPs at the regional level (N = 291,396) for K = 1-20.

For the TEs, we genotyped presence/absence of polymorphic copies of long terminal repeat retrotransposons (LTR-RTs) using TEPID (Stuart et al., 2016) and inferred their zygosity from read coverage at breakpoints to treat them similarly to SNPs. Of the 20,548 identified polymorphic TEs (Rogivue et al., 2019b), 3,874 loci with no missing data were retained for subsequent environmental association analyses and outlier detection.

245

246 Genotype–environment association analyses (GEA)

247 We performed GEA analyses (Lasky et al., 2022; Rellstab et al., 2015) at two

248 hierarchical levels (Figure 1): (i) the regional level, i.e., among the four studied

249 populations; and (ii) the local catchment level, i.e., among plots within each

250 population. We applied latent factor mixed models (LFMM; Frichot et al., 2013)

implemented in the R package *LFMM2* (Caye et al., 2019), a univariate GEA method in

which neutral genetic structure is intrinsically considered.

We used the 'snmf' function in the R package *LEA* to replace the missing observations (10%) in the SNP dataset with imputed data based on the population structure from the sNMF analysis (K = 1-10, entropy = T, and 10 repetitions). We then imputed the data with the 'impute' function in LEA, where latent factors (K) were obtained using the lowest genomic inflation factor (gif) value (method = 'mode'). Once the SNP datasets were complete, we ran the LEA function 'lfmm_ridge' for latent factors K = 1-8 for each environmental factor.

260 We defined the best number of latent factors (K) as the value with the 261 genomic inflation factor (λ , averaged over the 12 environmental factors) closest to 1 262 (Table S3), as suggested by Caye et al. (2019). We did not choose a specific K for each 263 environmental factor, as K has no specific biological meaning. Therefore, for the SNP 264 dataset at the regional level (i) we used K = 6, and at the local level (ii) we used K = 2265 for Essets, K = 3 for Martinets, K = 6 for Para, and K = 2 for Pierredar. For the TE 266 dataset, we used K = 1 for all analyses. Following Rogivue et al. (2019b), we treated 267 the two marker types (SNPs and polymorphic TEs) with different K values, reflecting

- the differential patterns of variation observed among populations (e.g., 6% of SNPs
- were shared among populations, compared with 90% of polymorphic TEs; SNPs had
- higher pairwise *F*_{ST} values (0.09–0.18) than those for polymorphic TEs (0.03–0.09)).
- 271 We ran the 'lfmm_test' function in *LFMM2* (Caye et al., 2019) to calculate the *z* scores
- and p values based on the λ and χ^2 distributions. To control for false positives, we
- applied the Benjamini-Hochberg algorithm (Benjamini & Hochberg, 1995) with a false
 discovery rate (FDR) of 1%.
- 275

276 **Outlier analysis**

To complement the GEA analyses, we used BayeScan 2.1 (Foll & Gaggiotti, 2008) to identify outlier SNPs and polymorphic TEs putatively under divergent selection, i.e., showing genetic differentiation above that expected for neutral markers. BayeScan is an *F*_{ST}-based approach that considers differences in allele frequencies between the common pool and each population. Outlier loci are those resulting from divergent selection beyond the level of what demographic processes, such as random genetic drift, may impose in terms of genetic differentiation.

- Like with LFMM, we ran BayeScan (i) at the regional level among the four populations, and (ii) at the local level among plots within each local population. We converted the SNP and TE datasets from LFMM to BayeScan format using a custom R script. We defined the prior odd at 100, with 5,000 outputted iterations, a thinning interval size of 10, 20 pilot runs of length 5,000, and a burn-in of 50,000. We visualised BayeScan results using Manhattan plots created with the R package *qqman* (Turner, 2018).
- 291

GO term enrichment analyses among levels of spatial extent, statistical methods, and marker types

294 We characterised the putative functions of candidate loci identified in the above

analyses using the same annotation as in Rogivue et al. (2019b). For simplicity, we

- 296 considered only the SNPs with a high impact on their respective protein based on
- 297 SnpEff (Cingolani et al., 2012), where high-impact SNPs were those identified as
- 298 missense, splice acceptor and splice donor variants, as well as start/stop codon loss

or gain. For similar reasons, we retained only polymorphic TEs located within 2 kb
upstream or downstream of annotated genes as possibly high-impact due to either
gene disruption or spreading of epigenetic marks lowering gene expression (Hollister
& Gaut, 2009).

We performed a gene ontology (GO) term enrichment analysis of the significantly associated SNPs and polymorphic TEs with topGO 2.44.0 (Alexa et al., 2006). We established the significance of the terms according to Fisher's exact tests (*p* value cut off at 0.01 and not adjusted for multiple testing, as recommended by the topGO authors).

We compared the annotated genes and enriched GO terms between hierarchical levels (regional vs. local extents), statistical methods (GEA vs. outlier analyses), and marker types (SNPs vs. polymorphic TEs) to identify analogous signals of adaptation. We searched for common high-impact SNPs and clusters (see below) among local populations and between levels of spatial extent, which we visualised using upset plots created with the R package *UpSetR* (Conway et al., 2017).

314

315 Clusters of loci identified as adaptive

We further compared common clusters of high-impact SNPs across the genome by defining 6228 blocks of 100 kbp, with a step of 50 kbp. We counted the number of high-impact SNPs, identified using either LFMM or BayeScan, per block and per region, where a cluster was assigned if a block or region contained at least two significant SNPs. We note that we summed all the LFMM-derived high-impact SNPs, irrespective of their associated environmental factors.

322 Results

323 Spatial genetic structure

324 The 291,396 SNPs retained at the regional level were used to determine the genetic

325 structure across the studied populations. Using the program Admixture with K = 1-20,

326 we found substantial genetic differentiation among populations despite the common

327 genomic background reported in a phylogeographic study across the Alps (Rogivue et

328 al., 2018). Admixture converged to a minimum cross-validation for 15 clusters, to

329 which 60% of individuals could be unambiguously assigned (membership coefficient

> 0.8), whereas admixed individuals were typically found in a single population. All

331 clusters were well nested within the four populations (3 in Essets, 4 in Martinets, 4 in

Para, 4 in Pierredar) and contained < 4% of the variants originating from other

333 populations (Figure 1).

334

335 Environmental factors and genotype–environment association (GEA) analysis

336 We evaluated 13 environmental factors derived from LiDAR-acquired DEMs at 0.5 m 337 resolution. This spatial resolution matches our sampling grain, i.e., individuals' sites 338 within plots (minimum distance between sampled plants > 1 m). Among these 339 environmental factors, direct insolation in June was highly positively correlated with 340 total insolation in June ($\rho = 0.99$; Table S4), so we discarded it to reduce collinearity 341 between explanatory variables in subsequent analyses. A principal component 342 analysis (PCA) on the retained 12 environmental factors indicated that the 343 environmental conditions across the four sampled populations were similar overall, 344 with the habitat conditions of sampled individuals spanning similar gradients along 345 the environmental factors considered (Figure S1). This finding substantiates the 346 reasoning that our sampling scheme provides optimal premises for investigating 347 convergent signatures of adaptation across populations (Rellstab et al., 2020) based 348 on genomic and environmental variation.

In GEA analyses (Figure S2), horizontal curvature was most prominently associated with genomic variation across all analyses, while eastness (with only a few significant associations) and slope (with none) seemed to be uninformative regarding signatures of selection (Table 1, Table S5). Notably, the environmental factors with the largest number of significant associations differed substantially among the local populations (Table S5).

The specificity of the environmental descriptors in driving local adaptation was assessed through patterns of interaction between the markers and the environmental factors. The majority of loci were associated with only one environmental factor, while up to 32.8% showed an association with multiple environmental factors in either the regional or local analyses (Figure 3a–e). The

360 spatial extent considered in the GEA analyses had a considerable effect on the 361 number and relative frequency of significant associations per environmental factor; 362 notably, each local population showed a distinct frequency distribution (Table 1, 363 Tables S5 and S6). Accordingly, only 260 high-impact SNPs and polymorphic TEs were 364 found to be significant between the regional and the four local analyses (Figure 2f). 365 These were located in 209 candidate adaptive genes shared among the two levels of 366 spatial extent (Figure S3a). Only one gene was identified in all five GEA analyses. 367 However, no ortholog of this gene is known from A. thaliana and, hence, its function 368 remains unknown.

For the two marker types, SNPs and polymorphic TEs, we found similar
percentages of loci that were significantly associated with at least one environmental
factor at the regional level: 2.46% of the SNPs and 2.48% of the polymorphic TEs.
Despite such broad variation, only one candidate gene (Aa_G559430), homologous to
the terpenoid cyclase AT3G14490 in *A. thaliana*, was highlighted by both SNPs and
polymorphic TEs.

375

Outlier analysis

377 At the regional level, BayeScan identified only two outlier SNPs, of which only one 378 was identified as high-impact. Conversley, between 15 (Essets) and 134 (Martinets) 379 high-impact SNPs significantly exceeded the neutral expectation at the local level (q 380 values < 0.1; Figure 4, Figure S4, Table 2, Table S7), though only one SNP was 381 detected across all local populations. Most of the outlier SNPs identified were tightly 382 flanked by other SNPs in linkage disequilibrium, which is known to extend across a 383 considerable physical distance (Rogivue et al., 2019b). Conversley, outlier analyses for 384 polymorphic TEs showed an inverted trend: few significant outliers were identified at 385 the local level (between zero in Essets and three in Martinets), while 108 significant 386 outliers were detected across the regional level (Table 2 and Figure S5). Comparing 387 the two statistical methods, GEA and outlier analyses, found neither common SNPs 388 nor genes at the regional level, with a few SNPs detected with both analyses at the 389 level of local populations (Table 2).

390

GO term enrichment

392 To substantiate the functional relevance of the loci putatively involved in divergent 393 selection, we performed tests for gene ontology (GO) term enrichment. We followed 394 the advice of the authors of topGO and did not apply any correction for multiple 395 testing to avoid excessive false negatives, at the cost of obtaining false positives. 396 Accordingly, the results of the enrichment analyses point towards, but do not 397 necessarily substantiate, genes of interest for being involved in adaptive processes. 398 Of the 7,180 associated SNPs identified by GEA at the regional level, we found 653 399 high-impact SNPs across all 12 environmental factors occurring within 519 annotated 400 genes (Table 1, Table S8), highlighting 42 significantly enriched GO terms (Table S9). 401 In turn, the 15 polymorphic TEs occurring within 2 kb upstream or downstream of 402 annotated genes highlighted 11 significantly enriched GO terms (Table 1, Tables S9 403 and S10). At the local level, high-impact SNPs and genes highlighted 22 significantly 404 enriched GO terms that were shared among three or four local populations (Figure 405 S3b). No gene flanking an associated polymorphic TE was identified in Essets or 406 Pierredar, whereas one was detected in Martinets and 62 in Para (Table S5). GO term 407 enrichment analyses were not considered for polymorphic TEs, as only a few linked 408 genes were identified.

409 At the regional-level, GEA analyses identified only two enriched GO terms 410 based on more than five genes comprising of at least one high-impact SNP, and these 411 were therefore considered top candidates (Table 1, highlighted in Table S9): 'defence 412 response' (GO:0006952) and 'innate immune response' (GO:0045087). The remaining 413 40 SNP-based significant GO terms and all 11 TE-based ones indicated a weaker signal 414 of selection (Table S9). Of the two outliers detected at the regional level, the SNP at 415 position 145,431 on chromosome 7 is a missense variant located in the gene 416 Aa G219310 (orthologous to AT4G38420 in A. thaliana), encoding a 'copper ion 417 binding' protein involved in the oxidation-reduction process (GO:0055114, 418 http://atgenie.org/transcript?id=AT4G38420.1). The outlier SNP located on 419 chromosome 8 is a synonymous variant situated in the gene Aa G499380 420 (orthologous to AT5G16970 in *A. thaliana*; Table S11) involved in responses to 421 oxidative stress; however, as a synonymous SNP it is likely not the locus directly

under selection, but may be linked to the causative variation (i.e., not captured
possibly for statistical reasons). No GO term analysis was possible because only one
high-impact SNP was annotated (Table 2).

425 At the local level, few GO terms were classified as top candidates: two in 426 Essets ('defence response' (GO:0006952) and 'innate immune response' 427 (GO:0045087)), one in Martinets ('protein phosphorylation' (GO:0006468)), three in 428 Para (three times 'protein phosphorylation' (GO:0006468)), and none in Pierredar 429 (Tables S5a and S9). Among the 134 identified high-impact outlier SNPs in Martinets 430 (Table S11), we found two top candidate GO terms: 'response to oomycetes' 431 (GO:0002239) and 'defence response to fungus, incompatible interaction' 432 (GO:0009817); Table S12). The three other regions presented no top candidate GO 433 terms (Table S12), despite the occurrence of several high-impact outliers (Table S11).

434

435 **Common clusters of adaptive loci**

436 We identified 12 common blocks (genomic regions spanning 100 kbp, with a step of 437 50 kbp) of high-impact SNPs detected by GEA analyses, most of which were shared 438 between at least two local populations (Figure 5). Only one genomic block was 439 common among three populations, and two (consecutive) genomic blocks were 440 shared between all four populations. Based on the outlier loci identified with 441 BayeScan, we found three common blocks, two of which were consecutive on 442 chromosome 3, which were common to the same two populations (Essets and 443 Pierredar) and in common with one population based on LFMM (Pierredar). The 444 remaining block was shared between Para and Pierredar. These 15 common blocks 445 comprised 211 significant associations in GEA analyses for 94 SNPs.

446 **Discussion**

- 447 Investigating the genes and functions that are under selection and identifying the
- 448 factors driving selection are essential for understanding local adaptation, particularly
- in view of global change. The high-quality and high-resolution genomic and
- 450 environmental data now available provide increasing insights into the evolution of
- 451 species and how environmental conditions can shape evolution. By assessing genomic

452 variation at the whole-genome level at both single nucleotides (SNPs) and TE markers 453 for 304 individuals, and by characterising each individual's micro-habitat, we found 454 that environmental factors describing soil water availability and solar radiation are 455 important drivers of local adaptation in A. alpina. While we detected loci showing 456 signals of local adaptation in the genic (high-impact SNPs and polymorphic TEs) and 457 non-genic (polymorphic TEs near to genes) genome regions, the detected signals 458 differed substantially among the four local populations, with inconsistencies across 459 the regional and local spatial extents. These findings highlight the specificity of local 460 adaptation for populations, despite these populations sharing a common genomic 461 background and experiencing similar environmental conditions (Fig. S1; Rogivue et 462 al., 2018). Such truly local responses to abiotic environmental cues may reflect in part 463 the high frequency of selfing observed in Alpine populations of A. alpina (Buehler et 464 al., 2012; Tedder et al., 2011), which could reinforce genomic imprints of local 465 adaptation once they have become established (Trickovic & Glémin, 2022). This 466 finding provides evidence that convergent signatures of selection, even within closely 467 situated populations of a given species, may only be expected under particularly 468 strong selective pressure and, thus, are rarely realised in populations of mostly selfing 469 individuals, as investigated here.

470

471 Singular signals of genomic adaptation across levels of spatial extent

472 In our hierarchical study design, we identified few gene candidates for local 473 adaptation common between the regional and local analyses (Figure 3f). While the 474 literature about the spatial extent of adaptation is still limited, two recent studies on 475 forest trees also showed that adaptation at different levels of spatial extent appears 476 complementary (Brousseau et al., 2021; von Takach et al., 2021). These authors 477 concluded that such scale issues are important to consider for understanding 478 genomic signatures of local adaptation, which is in line with our findings. The very 479 local adaptive response of populations reported here was already highlighted by 480 Rellstab et al. (2017) in a study on the strictly outcrossing Arabidopsis halleri. There, 481 only 31% of previously identified candidate adaptive SNPs were confirmed in an

independent set of populations, underlining the truly local nature of adaptation inresponse to a highly heterogeneous environment.

484 The low degree of overlapping results between the two marker types, SNPs 485 and polymorphic TEs, implies that their respective variation reflects adaptive 486 responses involving complementary candidate genes and gene functions. We cannot exclude potential effects of using different bioinformatic tools to characterize each 487 488 marker type, steming from fundamental differences between the types of 489 polymorphisms. For example, SNP-derived estimates of population-specific 490 inbreeding coefficients *F*_{IS} were lower than respective values calculated from 491 polymorphic TEs (Rogivue et al., 2019b). By ignoring sequence variation, we 492 underestimate heterozygosity in TEs, resulting in inflated F_{IS} values. Nevertheless, our 493 findings highlight the relevance of incorporating structural polymorphisms, such as 494 TEs and copy number variants, when studying complex evolutionary responses to 495 changing environments. Similar findings have been reported previously for 496 Brassicaceae (Niu et al., 2019; Quadrana et al., 2016), emphasising the need to 497 consider TE variation when investigating adaptation to global warming (Rey et al., 498 2016). The present study is, to our knowledge, the first to show that SNPs within 499 genes and polymorphic TE copies indicate complementary candidates of local 500 adaptation.

501 Unsurprisingly, the two statistical methods used (GEA and outlier detection) 502 revealed complementary outcomes and did not point to the same candidate genes. 503 To some degree, this inconsistency may be attributed to the smaller number of 504 candidates found in the outlier analysis (Table 2). Such a result may be associated 505 with sampling individuals in plots of contrasting environments, rather than along 506 abiotic gradients (Richardson et al., 2014). Additionally, the fundamental differences 507 between the two methodological approaches (statistical, environment-related GEA, 508 population genomic outlier detection) suggest that one should expect 509 complementary, rather than overlapping, signals of adaptation. Our results indeed 510 integrateeffects of selection across various abiotic and (unaccounted for) biotic 511 factors. Accordingly, conditional neutrality (Mee & Yeaman, 2019) might might at 512 least partly explain the small number of outliers detected that contrasts with the 513 numerous loci associated with environmental heterogeneity. Relatively high genetic

- 514 load was indeed detected within those expending alpine populations (Zeitler et al.,
- 515 2023), and corresponding loci may be conditionally deleterious to a large extent,
- 516 hence showing limited allelic differentiation despite significant genotype-by-
- 517 environment interactions. As simulations show that GEA analyses have a high power
- 518 of detection under scenarios of weak selection and low dispersal (Forester et al.,
- 519 2016), such approaches may be sensitive enough to identify conditionally neutral loci
- 520 (Lasky et al., 2022; Yoder & Tiffin, 2018).
- 521

Selective factors are distinct among SNPs and polymorphic TEs, and are predominantly consistent between levels of spatial extent

524 In our study, the type of genomic marker investigated considerably affected the

525 relative importance of the associated environmental factors (Table 1, Figure 3).

526 Horizontal curvature, a proxy for drainage and hydrological processes, detected the

527 most candidate SNPs. Conversely, GEAs using polymorphic TEs highlighted that the

528 factor with the largest number of significant candidates was total insolation in June

529 (or the highly correlated direct insolation in June), a factor representing solar

530 radiation. Although mechanistic underpinnings remain out of scope, this finding again

531 highlights the complementarity of SNPs and polymorphic TEs in revealing responses

532 to different environmental cues.

533 Across the two hierarchical levels of spatial extent, the order of importance of 534 the environmental factors differed slightly, while the top environmental factors 535 largely remained consistent (Table 1 and Table S5). This result suggests that the 536 studied populations inhabit similar environments and that our study design was 537 appropriate for detecting potentially convergent signatures of adaptation (Figure S1). 538 Nevertheless, environmental contrasts such as (indirectly accounted for) differences 539 in biotic interactions as well as restricted gene flow, and hence large effects of 540 genetic drift, likely contributed to overruling selective effects to impose a common 541 signal in the genomes of *A. alpina* individuals.

542

543 Defence-related functions are key in heterogeneous alpine environments

544 In alpine environments, plants must deal with harsh, highly variable conditions to 545 survive. In our study, high-impact polymorphisms in genes related to defence 546 responses are overrepresented among candidates of local adaptation (Table S9). 547 Likewise, loci identified by the outlier analyses also highlighted defence-related 548 functions under selection, though outlier loci did not directly link to any 549 environmental drivers. Biotic interactions are usually considered to play a minor role 550 when compared to abiotic environmental factors for plants at high elevations 551 (Zvereva & Kozlov, 2022). Nevertheless, despite our focus on potential abiotic drivers 552 of local adaptation, our study indicated that genes with biotic-related biological 553 functions are key in adaptive responses in alpine plants. Therefore, we advocate also 554 including biotic drivers in future environmental association analyses to better 555 represent potential environmental drivers of adaptation.

556 The two biological functions under selection at the regional level, i.e., 'defence 557 response' and 'innate immune response', were also found at the local level in Essets. 558 No other common biological function was found among the two levels of spatial extent, 559 again stressing the notion that local adaptation is truly local. Moreover, adaptive 560 functions previously described in studies on A. alpina, e.g., related to survival, 561 flowering and fecundity (de Villemereuil et al., 2018; Toräng et al., 2015; Wötzel et al., 562 2022), were not found in our analyses. This discrepancy might be due to differences in 563 the spatial extent and resolution of the studies, the complementary methodological 564 approaches, and the different types of environmental factors considered. Beyond this, 565 high levels of selfing encountered in Alpine populations of A. alpina may amplify 566 population-specific adaptive responses, though such an effect depends on various 567 conditions such as patterns of dominance, levels of selective pressure, and migration 568 rate (Trickovic & Glémin, 2022).

In conclusion, by combining whole-genome re-sequencing data with highresolution descriptions of environmental factors, our study pointed to several genes and biological functions potentially under selection. These findings pave the ground for a better understanding of adaptive processes and could guide further

573 investigations of *A. alpina* and related (alpine) species. Further, our findings suggest

574 that signatures of local adaptation are dependent on the spatial extent of the study, 575 the type of genomic markers employed, and the statistical methods used. The 576 common genomic blocks indicative of imprints of adaptation (Figure 5) refer to 577 regions in the genome that are under sufficiently strong selection to overcome the 578 presumed high level of genetic drift induced by demography and life-history traits of 579 A. alpina in our study regions. Taken together, our results suggest that 580 microevolutionary processes may likely result in non-overlapping signatures of local 581 adaptation across spatial scales.

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594 **References**

- Alexa, A., Rahnenführer, J., & Lengauer, T. (2006). Improved scoring of functional
 groups from gene expression data by decorrelating GO graph structure. *Bioinformatics*, 22(13), 1600–1607. doi:10.1093/bioinformatics/bt1140
- Alexander, D. H., Novembre, J., & Lange, K. (2009). Fast model-based estimation of
 ancestry in unrelated individuals. *Genome Research*, 19(9), 1655–1664.
 doi:10.1101/gr.094052.109
- Ansell, S. W., Grundmann, M., Russell, S. J., Schneider, H., & Vogel, J. C. (2008).
 Genetic discontinuity, breeding-system change and population history of *Arabis alpina* in the Italian Peninsula and adjacent Alps. *Molecular Ecology*, 17(9), 2245–
 2257. doi:10.1111/j.1365-294X.2008.03739.x
- Baduel, P., Leduque, B., Ignace, A., Gy, I., Gil, J., Loudet, O., Colot, V., & Quadrana,
 L. (2021). Genetic and environmental modulation of transposition shapes the
 evolutionary potential of *Arabidopsis thaliana*. *Genome Biology*, 22, 26.
 doi:10.1186/s13059-021-02348-5
- Benjamini, Y., & Hochberg, Y. (1995). Controlling for the false discovery rate a
 practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society Series B Statistical Methodology*, 57(1), 289–300. doi:10.1111/j.2517612 6161.1995.tb02031.x
- Bennetzen, J. L., & Wang, H. (2014). The contributions of transposable elements to
 the structure, function, and evolution of plant genomes. *Annual Review of Plant Biology*, 65, 505–530. doi:10.1146/annurev-arplant-050213-035811
- Beven, K., & Kirkby, M. J. (1979). A physically based, variable contributing area
 model of basin hydrology. *Hydrological Sciences*, 24(1), 43–69.
 doi:10.1080/02626667909491834
- Böhner, J., & Antonić, O. (2009). Land-surface parameters specific to topoclimatology. In T. Hengl & H. I. Reuter (Eds.), *Geomorphometry: Concepts, Software, Applications* (pp. 195–126): Elsevier.
- Böhner, J., & Selige, T. (2006). Spatial prediction of soil attributes using terrain
 analysis and climate regionalisation. *Göttinger Geographische Abhandlungen, 115*,
 13–28.
- Booker, T. R., Yeaman, S., & Whitlock, M. C. (2021). Global adaptation complicates
 the interpretation of genome scans for local adaptation. *Evolution Letters*, 5(1), 4–
 doi:10.1002/evl3.208
- 628 Brousseau, L., Fine, P. V. A., Dreyer, E., Vendramin, G. G., & Scotti, I. (2021).
- 629 Genomic and phenotypic divergence unveil microgeographic adaptation in the
 630 Amazonian hyperdominant tree *Eperua falcata* Aubl. (Fabaceae). *Molecular*
- 631 *Ecology*, *30*, 1136–1154. doi:10.1111/mec.15595
- Buckley, J., Widmer, A., Mescher, M. C., & De Moraes, C. M. (2019). Variation in
 growth and defence traits among plant populations at different elevations:
- 634 Implications for adaptation to climate change. *Journal of Ecology*, 107(5), 2478–
- 635 2492. doi:10.1111/1365-2745.13171
- Buehler, D., Graf, R., Holderegger, R., & Gugerli, F. (2012). Contemporary gene flow
 and mating system of *Arabis alpina* in a Central European alpine landscape. *Annals of Botany*, 109(7), 1359–1367. doi:10.1093/aob/mcs066
- 639 Caye, K., Jumentier, B., Lepeule, J., & François, O. (2019). LFMM 2.0: Latent factor
- 640 models for confounder adjustment in genome and epigenome-wide association
- 641 studies. *Molecular Biology and Evolution*, *36*(4), 852–860.
- 642 doi:10.1093/molbev/msz008

- 643 Chopra, D., Mapar, M., Stephan, L., Albani, M. C., Deneer, A., Coupland, G.,
- 644 Willing, E.-M., Schellmann, S., Schneeberger, K., Fleck, C., Schrader, A., &
- 645 Hülskamp, M. (2019). Genetic and molecular analysis of trichome development in
- 646 Arabis alpina. Proceedings of the National Academy of Sciences of the United
- 647 States of America, 116(Genetic and molecular analysis of trichome development in
- 648 Arabis alpina), 12078–12083. doi:10.1073/pnas.1819440116
- 649 Choudhury, R. A., Rogivue, A., Gugerli, F., & Parisod, C. (2019). Impact of
 650 polymorphic transposable elements on linkage disequilibrium along chromosomes.
 651 Molecular Ecology, 28(6), 1550–1562. doi:10.1111/mec.15014
- 652 Cingolani, P., Platts, A., Wang, L. L., Coon, M., Nguyen, T., Wang, L., Land, S. J.,
- Lu, X., & Ruden, D. M. (2012). A program for annotating and predicting the
 effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of *Drosophila melanogaster* strain w(1118); iso-2; iso-3. *Fly*, 6(2), 80–92.
 doi:10.4161/fly.19695
- 657 Conrad, O., Bechtel, B., Bock, M., Dietrich, H., Fischer, E., Gerlitz, L., Wehberg, J.,
 658 Wichmann, V., & Bohner, J. (2015). System for automated geoscientific analyses
 659 (SAGA) v. 2.1.4. *Geoscientific Model Development*, 8(7), 1991–2007.
- 660 doi:10.5194/gmd-8-1991-2015
- 661 Conway, J. R., Lex, A., & Gehlenborg, N. (2017). UpSetR: an R package for the
 662 visualization of intersecting sets and their properties. *Bioinformatics*, 33(18),
 663 2938–2940. doi:10.1093/bioinformatics/btx364
- Dauphin, B., Rellstab, C., Wüest, R. O., Karger, D. N., Holderegger, R., Gugerli, F.,
 & Manel, S. (2023). Re-thinking the environment in landscape genomics. *Trends in Ecology & Evolution*, 38(3), 261–274. doi:10.1016/j.tree.2022.10.010
- de Villemereuil, P., Mouterde, M., Gaggiotti, O. E., & Till-Bottraud, I. (2018).
 Patterns of phenotypic plasticity and local adaptation in the wide elevation range of
 the alpine plant *Arabis alpina*. *Journal of Ecology*, *106*(5), 1952–1971.
 doi:10.1111/1365-2745.12955
- Doneus, M. (2013). Openness as visualization technique for interpretative mapping of
 airborne lidar derived digital terrain models. *Remote Sensing*, 5(12), 6427–6442.
 doi:10.3390/rs5126427
- 674 Drake, J. M., Randin, C. F., & Guisan, A. (2006). Modelling ecological niches with
 675 support vector machines. *Journal of Applied Ecology*, 43(3), 424–432.
 676 doi:10.1111/j.1365-2664.2006.01141.x
- Eckert, A. J., Maloney, P. E., Vogler, D. R., Jensen, C. E., Delfino Mix, A., & Neale,
 D. B. (2015). Local adaptation at fine spatial scales: an example from sugar pine
- 679 (*Pinus lambertiana*, Pinaceae) *Tree Genetics & Genomes*, 11(3), 42.
- 680 doi:10.1007/s11295-015-0863-0
- Exposito-Alonso, M., Vasseur, F., Ding, W., Burbano, H. A. A., & Weigel, D. (2018).
 Genomic basis and evolutionary potential for extreme drought adaptation in *Arabidopsis thaliana. Nature Ecology & Evolution, 2*, 352–358.
- 684 doi:10.1038/s41559-017-0423-0
- Fischer, M. C., Rellstab, C., Tedder, A., Zoller, S., Gugerli, F., Shimizu, K. K.,
 Holderegger, R., & Widmer, A. (2013). Population genomic footprints of selection
 and associations with climate in natural populations of *Arabidopsis halleri* from
 the Alps. *Molecular Ecology*, 22(22), 5594–5607. doi:10.1111/mec.12521
- the Alps. *Molecular Ecology*, 22(22), 5594–5607. doi:10.1111/mec.12521
- Foll, M., & Gaggiotti, O. (2008). A genome-scan method to identify selected loci
 appropriate for both dominant and codominant markers: a Bayesian perspective.
- appropriate for both dominant and codominant markers: a Bayesian perspective.
 Genetics, 180, 977–993.
- 22

692 Forester, B. R., Jones, M. R., Joost, S., Landguth, E. L., & Lasky, J. R. (2016). 693 Detecting spatial genetic signatures of local adaptation in heterogeneous 694 landscapes. Molecular Ecology, 25(1), 104-120. doi:10.1111/mec.13476 695 Fournier-Level, A., Korte, A., Cooper, M. D., Nordborg, M., Schmitt, J., & Wilczek, 696 A. M. (2011). A map of local adaptation in Arabidopsis thaliana. Science, 334(6052), 86-89. doi:10.1126/science.1209271 697 698 Frichot, E., Schoville, S. D., Bouchard, G., & François, O. (2013). Testing for 699 associations between loci and environmental gradients using latent factor mixed 700 models. Molecular Biology and Evolution, 30(7), 1687–1699. 701 Garrison, E., & Marth, G. (2012). Haplotype-based variant detection from short-read 702 sequencing. arXiv. 703 Gougherty, A. V., Keller, S. R., & Fitzpatrick, M. C. (2021). Maladaptation, 704 migration and extirpation fuel climate change risk in a forest tree species. *Nature* 705 Climate Change, 11, 166-171. doi:10.1038/s41558-020-00968-6 706 Guillaume, A. S., Leempoel, K., Rogivue, A., Rochat, E., Kasser, M., Gugerli, F., 707 Parisod, C., & Joost, S. (2021). Multiscale very-high resolution topographic 708 models in Alpine ecology: pros and cons of airborne LiDAR and drone-based 709 stereo-photogrammetry technologies. Remote Sensing, 13. doi:10.3390/rs13081588 710 Guisan, A., & Zimmermann, N. (2000). Predictive habitat distribution models in 711 ecology. Ecological Modelling, 135(2-3), 147-186. 712 Hancock, A. M., Brachi, B., Faure, N., Horton, M. W., Jarymowycz, L. B., Sperone, 713 F. G., Toomajian, C., Roux, F., & Bergelson, J. (2011). Adaptation to climate 714 across the Arabidopsis thaliana genome. Science, 334(6052), 83-86. 715 Hjerdt, K., McDonnell, J., Seibert, J., & Rodhe, A. (2004). A new topographic index 716 to quantify downslope controls on local drainage. Water Resources Research, 717 40(5), 199-207. doi:10.1029/2004WR003130 718 Hollister, J. D., & Gaut, B. S. (2009). Epigenetic silencing of transposable elements: 719 A trade-off between reduced transposition and deleterious effects on neighboring 720 gene expression. Genome Research, 19(1419-1428). doi:10.1101/gr.091678.109 721 Jiao, W.-B., Garcia Accinelli, G., Hartwig, B., Kiefer, C., Baker, D., Severing, E., 722 Willing, E.-M., Piednoël, M., Woetzel, S., Madrid-Herrero, E., Huettel, B., 723 Hürmann, U., Reinhard, R., Koch, M. A., Swan, D., Clavijo, B., Coupland, G., & 724 Schneeberger, K. (2017). Improving and correcting the contiguity of long-read 725 genome assemblies of three plant species using optical mapping and chromosome 726 conformation capture data. Genome Research, 27, 778-786. 727 doi:10.1101/gr.213652.116 728 Kalendar, R., Tanskanen, J., Immonen, S., Nevo, E., & Schulman, A. H. (2000). 729 Genome evolution of wild barley (Hordeum spontaneum) by BARE-1 730 retrotransposon dynamics in response to sharp microclimatic divergence. Proceedings of the National Academy of Sciences of the United States of America. 731 732 97(12), 6603-6607. doi:10.1073/pnas.110587497 733 Kawecki, T. J., & Ebert, D. (2004). Conceptual issues in local adaptation. Ecology 734 Letters, 7(12), 1225–1241. doi:10.1111/j.1461-0248.2004.00684.x 735 Kozak, K. H., Graham, C. H., & Wiens, J. J. (2008). Integrating GIS-based 736 environmental data into evolutionary biology. Trends in Ecology & Evolution, 737 23(3), 141–148. 738 Laenen, B., Tedder, A., Nowak, M. D., Toräng, P., Wunder, J., Wötzel, S., Steige, K. 739 A., Kourmpetisd, Y., Odong, T., Drouzas, A. D., Bink, M. C. A. M., Ågren, J., Coupland, G., & Slotte, T. (2018). Demography and mating system shape the 740 741 genomewide impact of purifying selection in Arabis alpina. Proceedings of the

742 National Academy of Sciences of the United States of America, 115(4), 816–821. 743 doi:10.1073/pnas.1707492115 744 Lasky, J. R., Josephs, E. B., & Morris, G. P. (2022). Genotype-environment 745 associations to reveal the molecular basis of environmental adaptation. The Plant 746 Cell, 2023(35), 1. doi:10.1093/plcell/koac267 Lecours, V., Devillers, R., Simms, A. E., Lucieer, V. L., & Brown, C. J. (2017). 747 748 Towards a framework for terrain attribute selection in environmental studies. 749 Environmental Modeling & Software, 89, 19–30. 750 doi:10.1016/j.envsoft.2016.11.027 751 Leempoel, K., Parisod, C., Geiser, C., Daprà, L., Vittoz, P., & Joost, S. (2015). Very 752 high resolution digital elevation models: are multi-scale derived variables 753 ecologically relevant? Methods in Ecology and Evolution, 6(12), 1373–1383. 754 doi:10.1111/2041-210X.12427 755 Leempoel, K., Parisod, C., Geiser, C., & Joost, S. (2018). Multiscale landscape 756 genomic models to detect signatures of selection in the alpine plant *Biscutella* 757 laevigata. Ecology and Evolution, 8, 1794–1806. doi:10.1002/ece3.3778 758 Li, H., & Durbin, R. (2010). Fast and accurate long-read alignment with Burrows-759 Wheeler transform. *Bioinformatics*. 26(5), 589–595. doi:10.1093/bioinformatics/btp698 760 761 Lobréaux, S., & Miquel, C. (2020). Identification of Arabis alpina genomic regions 762 associated with climatic variables along an elevation gradient through whole 763 genome scan. Genomics, 112(1), 729-735. doi:10.1016/j.ygeno.2019.05.008 764 Lotterhos, K. E., & Whitlock, M. C. (2015). The relative power of genome scans to 765 detect local adaptation depends on sampling design and statistical method. 766 Molecular Ecology, 24(5), 1031–1046. doi:10.1111/mec.13100 767 Lyon, S. W., Troch, P. A., Broxton, P. D., Molotch, N. P., & Brooks, P. D. (2008). 768 Monitoring the timing of snowmelt and the initiation of streamflow using a 769 distributed network of temperature/light sensors. *Ecohydrology*, 1(3), 215–224. 770 doi:10.1002/eco.18 771 Mee, J. A., & Yeaman, S. (2019). Unpacking conditional neutrality: genomic 772 signatures of selection on conditionally beneficial and conditionally deleterious 773 mutations. The American Naturalist, 194(4), 529-540. doi:10.1086/702314 774 Moore, I. D., Grayson, R., & Ladson, A. (1991). Digital terrain modelling: a review of 775 hydrological, geomorphological, and biological applications. Hydrological 776 Processes, 5(1), 3-30. doi:10.1002/hyp.3360050103 Niu, X.-M., Xu, Y.-C., Li, Z.-W., Bian, Y.-T., Hou, X.-H., Chen, J.-F., Zou, Y.-P., 777 778 Jiang, J., Wu, O., Ge, S., Balasubramanian, S., & Guo, Y.-L. (2019). Transposable 779 elements drive rapid phenotypic variation in Capsella rubella. Proceedings of the 780 National Academy of Sciences of the United States of America, 116(14), 6908– 781 6913. doi:10.1073/pnas.1811498116 782 Pais, A. L., Whetten, R. W., & Xiang, O.-Y. (2017). Ecological genomics of local 783 adaptation in Cornus florida L. by genotyping by sequencing. Ecology and 784 Evolution, 7(1), 451-465. doi:10.1002/ece3.2623 785 Poncet, B., Herrmann, D., Gugerli, F., Taberlet, P., Holderegger, R., Gielly, L., 786 Rioux, D., Thuiller, W., Aubert, S., & Manel, S. (2010). Tracking genes of 787 ecological relevance using a genome scan: application to Arabis alpina. Molecular Ecology, 19(14), 2896–2907. doi:10.1111/j.1365-294X.2010.04696.x 788 789 Pradervand, J.-N., Dubuis, A., Pellissier, L., Guisan, A., & Randin, C. F. (2014). Very 790 high resolution environmental predictors in species distribution models: moving 791 beyond topography? Progress in Physical Geography, 38, 79–96.

792	Quadrana, L., Bortolini Silveira, A., Mayhew, G. F., LeBlanc, C., Martienssen, R. A.,
793	Jeddeloh, J. A., & Colot, V. (2016). The Arabidopsis thaliana mobilome and its
794	impact at the species level. eLIFE, 5, 25. doi:10.7554/eLife.15716
795	Quesneville, H. (2020). Twenty years of transposable element analysis in the
796	Arabidopsis thaliana genome. Mobile DNA, 11, 13. doi:10.1186/s13100-020-
797	00223-x
798	R Development Core Team. (2021). R: A language and environment for statistical
799	computing. Vienna: R Foundation for Statistical Computing. Retrieved from
800	http://www.R-project.org
801	Rellstab, C., Fischer, M. C., Zoller, S., Graf, R., Tedder, A., Shimizu, K. K., Widmer,
802	A., Holderegger, R., & Gugerli, F. (2017). Local adaptation (mostly) remains local:
803	reassessing environmental associations of climate-related candidate SNPs in
804	Arabidopsis halleri. Heredity, 118(1), 193–201. doi:10.1038/hdy.2016.82
805	Rellstab, C., Gugerli, F., Eckert, A. J., Hancock, A. M., & Holderegger, R. (2015). A
806	practical guide to environmental association analysis in landscape genomics.
807	<i>Molecular Ecology</i> , 24(17), 4348–4370. doi:10.1111/mec.13322
808	Rellstab, C., Zoller, S., Sailer, C., Tedder, A., Gugerli, F., Shimizu, K. K.,
809	Holderegger, R., Widmer, A., & Fischer, M. C. (2020). Genomic signatures of
810	convergent adaptation to Alpine environments in three Brassicaceae species.
811	<i>Molecular Ecology</i> , 29(22), 4350–4365. doi:10.1111/MEC.15648
812	Rey, O., Danchin, E., Mirouze, M., Loot, C., & Blanchet, S. (2016). Adaptation to
813	global change: a transposable element-epigenetics perspective. Trends in Ecology
814	& Evolution, 31(7), 514–526. doi:10.1016/j.tree.2016.03.013
815	Richardson, J. L., Urban, M. C., Bolnick, D. I., & Skelly, D. K. (2014).
816	Microgeographic adaptation and the spatial scale of evolution. <i>Trends in Ecology</i>
817	& Evolution, 29(3), 165–176.
818	Rogivue, A., Choudhury, R. R., Zoller, S., Joost, S., Felber, F., Kasser, M., Parisod,
819	C., & Gugerli, F. (2019a). Data from: Genome-wide variation in nucleotides and
820	retrotransposons in alpine populations of Arabis alpina (Brassicaceae). Retrieved
821	from: <u>https://datadryad.org/resource/doi:10.5061/dryad.58g217k</u>
822 823	Rogivue, A., Choudhury, R. R., Zoller, S., Joost, S., Felber, F., Kasser, M., Parisod,
823 824	C., & Gugerli, F. (2019b). Genome-wide variation in nucleotides and
824 825	retrotransposons in alpine populations of <i>Arabis alpina</i> (Brassicaceae). <i>Molecular Ecology Resources, 19</i> (3), 773–787. doi:10.1111/1755-0998.12991
825	Rogivue, A., Graf, R., Parisod, C., Holderegger, R., & Gugerli, F. (2018). The
820	phylogeographic structure of <i>Arabis alpina</i> in the Alps shows consistent patterns
827	across different types of molecular markers and of geographic scales. <i>Alpine</i>
829	<i>Botany, 128</i> (1), 35–45. doi:10.1007/s00035-017-0196-8
830	Sappington, J. M., Longshore, K. M., & Thompson, D. B. (2007). Quantifying
831	landscape ruggedness for animal habitat analysis: a case study using bighorn sheep
832	in the Mojave Desert. <i>Journal of Wildlife Management, 71</i> , 1419–1426.
833	Savolainen, O., Lascoux, M., & Merilä, J. (2013). Ecological genomics of local
834	adaptation. Nature Reviews Genetics, 14, 807–820.
835	Stuart, T., Eichten, S. R., Cahn, J., Karpievitch, Y. V., Borevitz, J. O., & Lister, R.
836	(2016). Population scale mapping of transposable element diversity reveals links to
837	gene regulation and epigenomic variation. <i>eLIFE</i> , 5, 27. doi:10.7554/eLife.2077
838	Tedder, A., Ansell, S. W., Lao, X., Vogel, J. C., & Mable, B. K. (2011). Sporophytic
839	self-incompatibility genes and mating system variation in Arabis. Annals of
840	Botany, 108(4), 699–713. doi:10.1093/aob/mcr157

- Toräng, P., Wunder, J., Obeso, R. J., Herzog, M., Coupland, G., & Ågren, J. (2015).
 Large-scale adaptive differentiation in the alpine perennial herb *Arabis alpina*. *New Phytologist. 206*, 459–470. doi:10.1111/nph.13176
- Tovar-Pescador, J., Pozo-Vázquez, D., Ruiz-Arias, J., Batlles, J., López, G., & Bosch,
 J. (2006). On the use of the digital elevation model to estimate the solar radiation
 in areas of complex topography. *Meteorological Applications*, 13(3), 279–287.
 doi:10.1017/S1350482706002258
- Trickovic, B., & Glémin, S. (2022). Establishment of local adaptation in partly self fertilizing population. *Genetics*, 220. doi:10.1093/genetics/iyab201
- Turner, S. D. (2018). qqman: an R package for visualizing GWAS results using Q-Q
 and manhattan plots. *Journal of Open Source Software*, *3*, 2.
 doi:10.21105/joss.00731
- Turner, T. L., Bourne, E. C., Von Wettberg, E. J., Hu, T. T., & Nuzhdin, S. V. (2010).
 Population resequencing reveals local adaptation of *Arabidopsis lyrata* to
 serpentine soils. *Nature Genetics*, 42(3), 260–263.
- Vayssières, A., Mishra, P., Roggen, A., Neumann, U., Ljung, K., & Albani, M. C.
 (2020). Vernalization shapes shoot architecture and ensures the maintenance of
 dormant buds in the perennial *Arabis alpina*. *New Phytologist*, 227(1), 99–115.
 doi:10.1111/nph.16470
- von Takach, B., Ahrens, C. W., Lindenmayer, D. B., & Banks, S. C. (2021). Scaledependent signatures of local adaptation in a foundation tree species. *Molecular Ecology*, 30(10), 2248–2261. doi:10.1111/mec.15894
- Wang, R., Farrona, S., Vincent, C., Joecker, A., Schoof, H., Turck, F., AlonsoBlanco, C., Coupland, G., & Albani, M. C. (2009). *PEP1* regulates perennial
 flowering in *Arabis alpina*. *Nature*, 459(7245), 423–427. doi:10.1038/nature07988
- Wilson, J. P., & Gallant, J. C. (Eds.). (2000). Terrain analysis: principles and *applications*. New York: Wiley.
- Wingler, A., Juvany, M., Cuthbert, C., & Munné-Bosch, S. (2015). Adaptation to
 altitude affects the senescence response to chilling in the perennial plant *Arabis alpina. Journal of Experimental Botany*, 66(1), 355–367. doi:10.1093/jxb/eru426
- Wötzel, S., Andrello, M., Albani, M. C., Koch, M. A., Coupland, G., & Gugerli, F.
 (2022). Arabis alpina: a perennial model plant for ecological genomics and lifehistory evolution. Molecular Ecology Resources, 22(2), 468–486.
- 874 doi:10.1111/1755-0998.13490
- 875 Yeaman, S., Hodgins, K. A., Lotterhos, K. E., Suren, H., Nadeau, S., Degner, J. C.,
- 876 Nurkovski, K. A., Smets, P., Wang, T., Gray, L. K., Liepe, K. J., Hamann, A.,
- 877 Holliday, J. A., Whitlock, M. C., Rieseberg, L. H., & Aitken, S. N. (2016).
- 878 Convergent local adaptation to climate in distantly related conifers. *Science*, 879 353(6306) 1431 1433 doi:10.1126/science.asf7812
- 879 *353*(6306), 1431–1433. doi:10.1126/science.aaf7812
- Yoder, J. B., Stanton-Geddes, J., Zhou, P., Briskine, R., Young, N. D., & Tiffin, P.
 (2014). Genomic signature of adaptation to climate in *Medicago truncatula*.
- 882 *Genetics*, 196(4), 1263–1275. doi:10.1534/genetics.113.159319
- Yoder, J. B., & Tiffin, P. (2018). Effects of gene action, marker density, and timing of
 selection on the performance of landscape genomic scans of local adaptation. *Journal of Heredity*, 109(1), 16–28. doi:10.1093/jhered/esx042
- 886 Yokoyama, R., Shirasawa, M., & Pike, R. J. (2002). Visualizing topography by
- 887 openness: a new application of image processing to digital elevation models.
- 888 Photogrammetric Engineering and Remote Sensing, 68(3), 257–266.

- 889 Zeitler, L., Parisod, C., & Gilbert, K. J. (2023). Purging due to self-ferilization does
- 890 not prevent accumulation of expansion load. *PLoS Genetics*, 19, e1010883.
- 891 doi:10.1371/journal.pgen.1010883
- Zvereva, E. L., & Kozlov, M. V. (2022). Meta-analysis of elevational changes in the
 intensity of trophic interactions: Similarities and dissimilarities with latitudinal
- 894 patterns. *Ecology Letters*, 25(9), 2076–2087. doi:10.1111/ele.14090

896 Data Accessibility and Benefit-Sharing

- 897 All data and custom-made codes are available in the DRYAD repository at
- 898 http://doi.org/...., reference number
- A Benefit-Sharing statement does not apply, given that all samples originated from
- 900 the country where the research was conducted.
- 901

902 Author contributions

- A.R., K.L., R.R.C., F.F., M.K., S.J., C.P. and F.G. initiated and conceived the project. AR
- 904 performed the experiments. A.R., K.L., A.S.G. and R.R.C. analysed the data. A.R., K.L.,
- 905 R.R.C., A.S.G., M.K., S.J. and C.P. contributed analysis tools. A.R., K.L. and F.G. wrote
- 906 the initial manuscript, with contributions from all co-authors.

907 Conflict of interest

908 The authors declare that they do not have any conflict of interest.

910 Tables and Figures

911 TABLE 1 Results of environmental association analyses in *Arabis alpina* at the

912 regional level (latent factor mixed model, LFMM). The number (#) of significant

913 single-nucleotide polymorphisms (SNPs), high-impact SNPs (i.e., missense, splice

914 acceptor and splice donor variants, as well as start/stop codon loss or gain) and

915 polymorphic transposable elements (TEs) within 2 kbp of a gene, as well as the

916 number of associated genes and significantly enriched gene ontology (GO) terms

917 associated with each environmental factor, with less (†) or more (‡) than five genes

918 significant. The three environmental factors most frequently associated with SNPs

919 and polymorphic TEs are highlighted in bold and italics, respectively.

920

	# associ	ated loci		# associa genes	ted	# significantly enriched GO terms †		
Environmental factor	SNPs	High- impact SNPs	TEs	From SNPs	From TEs	From SNPs	From TEs	
Total insolation in June	0	0	42	0	9	0	5	
Sky view factor	243	31	0	17	0	5	0	
Topographic wetness index	52	1	0	1	0	0	0	
Neg. topographic openness	151	6	0	6	0	0	0	
Pos. topographic openness	82	21	13	9	4	7	6	
Vector ruggedness measure	1767	179	3	148	0	5	0	
Downslope distance gradient	224	16	0	14	0	5	0	
Horizontal curvature	3355	286	36	236	0	8	0	
Vertical curvature	924	78	0	62	0	6 + 2‡	0	
Northness	382	35	0	26	0	4	0	
Eastness	0	0	2	0	2	0	0	
Slope	0	0	0	0	0	0	0	
Total	7180	653	96	519	15	42	11	

921 *† p <* 0.01, < 5 genes significant

922 *‡ p* < 0.01, > 5 genes significant

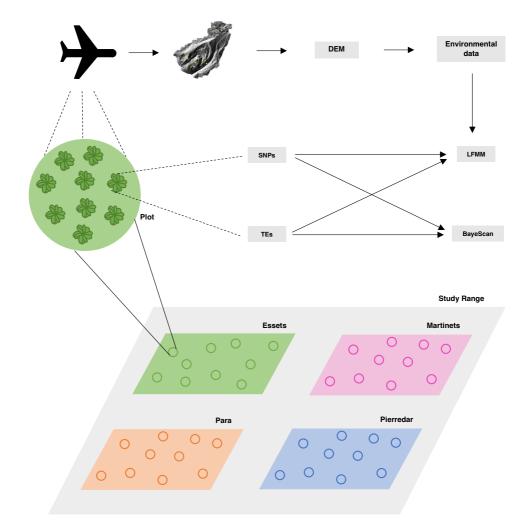
923 TABLE 2 Number of outlier loci detected by BayeScan in *Arabis alpina*. The number of significant single-nucleotide polymorphisms (SNPs) and 924 polymorphic transposable elements (TEs) at the regional level and for each of the four local populations, the number of high-impact SNPs and 925 TEs, the number of genes containing SNPs or TEs, and the number of significantly enriched gene ontology (GO) terms for SNPs and TEs, with 926 less (†) or more (‡) than five genes significant.

927

		# outliers (q value < 0.1)		# high-impact		# genes		# enriched GO terms †		# common with LFMM		# common gene with LFMM	
Spatial extent	Local population	SNPs	TEs	SNPs	TEs	SNPs	TEs	SNPs	TEs	SNPs	TEs	SNPs	TEs
Regional		2	108	1	2	1	0	0	0	0	6	0	1
Local	Essets	176	0	15	0	14	0	4	0	0	0	0	0
	Martinets	2032	3	134	0	134	0	5 + 2‡	0	26	0	24	0
	Para	1193	2	100	0	100	0	6	0	1	0	2	0
	Pierredar	599	1	27	0	27	0	5	0	2	0	3	0

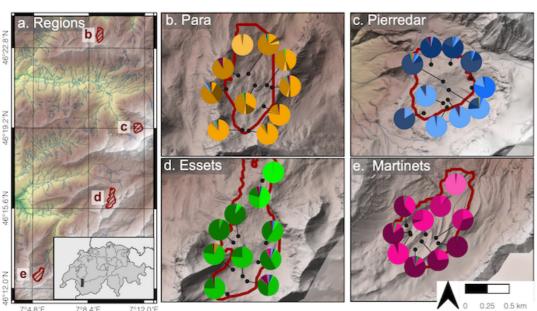
928 *t p* < 0.01, < 5 genes significant

929 **‡** *p* < 0.01, > 5 genes significant



930

931 FIGURE 1 Schematic illustration of the study design and analyses. Four local 932 populations were selected (illustrated by rectangles), each comprising ten plots 933 (circles) with six to ten plants each. Individual plants were sampled and sequenced to 934 identify single-nucleotide polymorphisms (SNPs) and polymorphic transposable 935 elements (TEs). Environmental data used to characterise the habitat of each 936 individual were computed from a regional high-resolution digital elevation model 937 (DEM) at 0.5 m pixel resolution, based on light detection and ranging (LiDAR). Two 938 types of analyses were performed to search for genomic signatures of selection: (i) 939 genotype-environment association (GEA) analyses using a latent factor mixed model 940 (LFMM; Frichot et al., 2013) and (ii) outlier analyses using BayeScan (Foll & Gaggiotti, 941 2008). Both types of analyses were performed at two hierarchical levels differing in 942 spatial extent: (i) at the regional level among the four local populations and (ii) at the 943 local level among plots within each population.



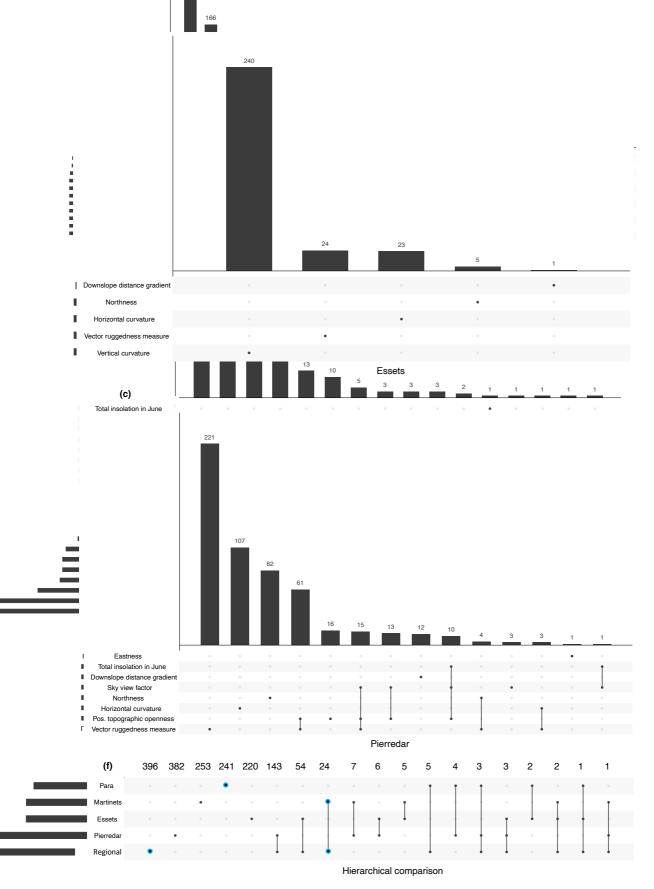
945 FIGURE 2 Locations of the four local populations of Arabis alpina and genetic

946 assignment. (a) The study region situated in the western Swiss Alps, with four local

populations (b) Para, (c) Pierredar, (d) Essets and (e) Martinets. Pie charts in (b)–(e) 947

948 represent the proportions of assignment probabilities for each plot (black dot) within

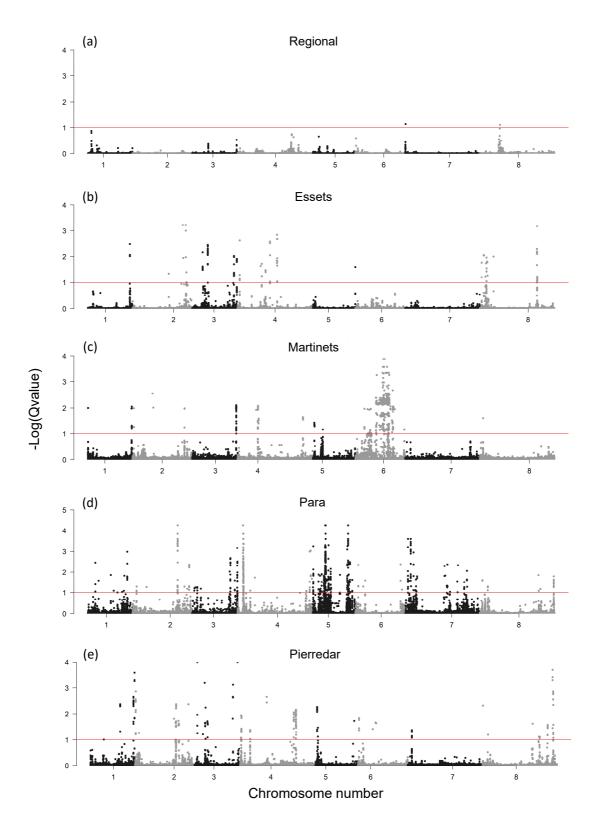
- 949 each studied region (delimited by a red line).
- 950



952 FIGURE 3 Common associations between environmental factors and hierarchical

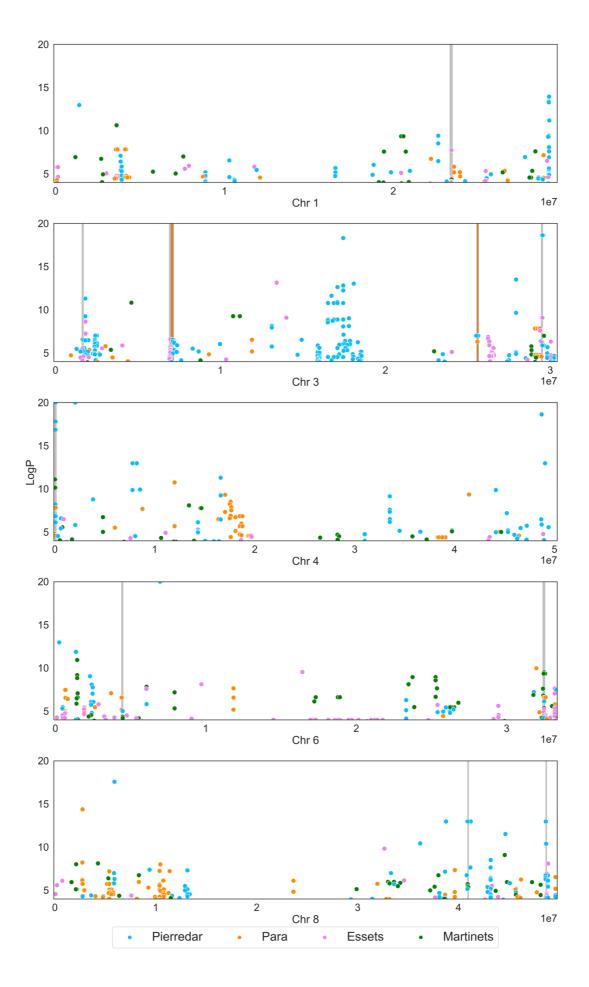
953 levels. Combination and magnitude of intersections of the significantly associated

- 954 single-nucleotide polymorphisms (SNPs, high-impact) and polymorphic transposable
- 955 elements (TEs) from the latent factor mixed models (LFMM) comparing
- 956 environmental factors within each analysis. (a) Regional level, (b)–(e) local level, and
- 957 (f) between hierarchical levels. The number of loci involved in each category is given
- 958 above the corresponding plot.



960 FIGURE 4 Manhattan plots showing BayeScan outliers. The analysis is based on the 961 genome scan of single-nucleotide polymorphisms (SNPs) in Arabis alpina for the two 962 levels of spatial extent: (a)–(d) local and (e) regional. Alternating black/grey dots 963 mark detected SNPs within each of eight chromosomes (numbers); red lines 964

- represent the false discovery rate (FDR, 10%). Values of genetic differentiation F_{ST} as
- 965 a function of the log(q value) can be found in Table S7.



- 967 FIGURE 5 Genomic regions indicating signatures of adaptation. Significant high-
- 968 impact single-nucleotide polymorphisms (SNPs) in Arabis alpina, identified in
- 969 genotype–environment association (GEA) analyses, with common chromosomal
- 970 blocks. Panels show the log *p*-value of all high-impact SNPs identified per local
- 971 population (denoted with different colours) for each of the five (out of eight)
- 972 chromosomes (CHR) of *A. alpina* for which common blocks could be found. Blocks
- 973 are denoted if detected in at least two populations, with vertical lines in grey for GEA
- 974 analyses and in brown for outlier analyses. Numbers in x-axis denote length of
- 975 chromosome in Mbp.

976 Supporting Information

977	Supportin information is available for this paper currently as a zip archive, except for
978	Figure S2 shared via a link to Google Doc (see below).
979	
980	Table S1 Geographical coordinates and values of the 13 environmental factors for
981	each sampled individual of Arabis alpina in the four local populations.
982	
983	Table 2 Definitions of the 13 environmental factors.
984	
985	Table S3 The genomic inflation factor λ , calculated using latent factor mixed models
986	(LFMMs), for single-nucleotide polymorphisms (SNPs) and polymorphic transposable
987	elements (TEs) in Arabis alpina.
988	
989	Table S4 Spearman's $ ho$ between pairs of 13 environmental factors, measured in the
990	four local populations of Arabis alpina.
991	
992	Table S5 Summary results of the latent factor mixed models (LFMMs) for the single-
993	nucleotide polymorphisms (SNPs) and the polymorphic transposable elements (TEs)
994	in Arabis alpina.
995	
996	Table S6 Associated single-nucleotide polymorphisms (SNPs) and polymorphic
997	transposable elements (TEs) in Arabis alpina, based on the latent factor mixed
998	models (LFMMs). (a) SNPs regional, (b) TEs regional, (c) SNPs local and (d) TEs local.
999	
1000	Table S7 Results of the outlier detection analyses, using BayeScan, based on single-
1001	nucleotide polymorphisms (SNPs) in Arabis alpina at the regional and local levels.
1002	
1003	Table S8 Annotated single-nucleotide polymorphisms (SNPs) in Arabis alpina at the
1004	regional and local levels, based on the results of latent factor mixed models
1005	(LFMMs).
1006	

1007	Table S9 Results of the gene ontology (GO) enrichment analysis for latent factor
1008	mixed models (LFMMs) with single-nucleotide polymorphisms (SNPs) and
1009	polymorphic transposable elements (TEs) in Arabis alpina at the regional and local
1010	levels.
1011	
1012	Table S10 Annotated associated polymorphic transposable elements (TEs) in Arabis
1013	alpina at the regional and local levels, based on the latent factor mixed models
1014	(LFMMs).
1015	
1016	Table S11 Annotated single-nucleotide polymorphisms (SNPs) in Arabis alpina at the
1017	regional and local levels, based on outlier detection analyses using BayeScan.
1018	
1019	Table S12 Gene ontology (GO) terms in Arabis alpina at the regional and local levels,
1020	based on the results of outlier detection analyses of single-nucleotide
1021	polymorphisms (SNPs) using BayeScan.
1022	
1023	
1024	Figure S1 Principal component analysis (PCA) including the 304 individuals of Arabis
1025	alpina from the four local populations, based on the twelve retained environmental
1026	factors.
1027	
1028	Figure S2 Manhattan plots of the latent factor mixed model (LFMM) analyses for
1029	Arabis alpina.
1030	https://drive.google.com/file/d/1 VbJxP9xHOdYLqtuMdzN3rcrosPwjqeg/view?usp=s
1031	hare link
1032	
1033	Figure S3 Shared genes and gene ontology terms between levels of spatial extent in
1034	Arabis alpina.
1035	
1036	Figure S4 Plots of the results of the outlier detection analyses (using BayeScan),
1037	based on the genome scan of single-nucleotide polymorphisms (SNPs) in Arabis

- 1038 *alpina*: (a) regional analysis and (b)–(e) local analyses. (b) Essets, (c) Martinets, (d)
- 1039 Para and (e) Pierredar.
- 1040
- 1041 Figure S5 Manhattan plots of the results of the outlier detection analysis (using
- 1042 BayeScan), based on the genome scan of polymorphic transposable elements (TEs) in
- 1043 Arabis alpina.