

# Enantiomeric and Monoaminergic Contributions to Methamphetamine's Bidirectional Effects on Fentanyl-Depressed Respiration in Mice

Harrison Elder<sup>1</sup>, David Walentiny<sup>1</sup>, and Patrick Beardsley<sup>2</sup>

<sup>1</sup>Virginia Commonwealth University School of Medicine

<sup>2</sup>Virginia Commonwealth University

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## Abstract

**Rationale:** Fentanyl remains the primary cause of fatal overdoses, and its co-use with methamphetamine (METH) is a growing concern. The optical isomers of METH, dextromethamphetamine (d-METH) and levomethamphetamine (l-METH), differ substantially in dose expression and thus may differentially contribute to the racemate's bidirectional effects. Furthermore, it is unknown which of METH's monoamine (MA) receptor mechanisms mediate these respiratory effects. Thus, systematic evaluation of monoamine receptor selective agents may identify treatment targets for OIRD. **Methods:** The two optical isomers of METH, d-METH and l-METH, were tested in adult male mice to determine their effects on basal and fentanyl-depressed minute volume (MVb; i.e., respiratory frequency x tidal volum) using whole-body plethysmography. Next, six selective agonists at MA receptors involved in METH's activity [phenylephrine (PNE;  $\alpha$ 1), clonidine (CLON;  $\alpha$ 2), SKF-82958 (SKF; D1), quinpirole (QPR; D2), 8-OH-DPAT (8-OH; 5HT1A), and DOI (5HT2)] were singly tested on basal MVb, and then in combination with fentanyl. **Results:** d-METH elevated MVb and l-METH decreased MVb. Under fentanyl-depressed conditions, the bidirectional effects of racemic METH were recreated by d-METH while l-METH significantly exacerbated OIRD at 1.0 and 3.0 mg/kg. MVb was dose-dependently increased by PNE and SKF and decreased by CLON and QPR. Neither 8-OH nor DOI altered basal MVb. Under fentanyl-depressed conditions, SKF transiently elevated MVb, while PNE more persistently increased it, while DOI transiently increased MVb, and 8-OH decreased MVb. **Conclusions:** d-METH and l-METH differentially contribute to the bidirectional respiratory modulation observed with the racemate and selective activation of MA receptors altered basal respiration and OIRD.

1 **Enantiomeric and Monoaminergic Contributions to Methamphetamine's Bidirectional**  
2 **Effects on Fentanyl-Depressed Respiration in Mice**

3

4 **AUTHOR NAMES:**

5 Harrison J. Elder<sup>a,b,\*</sup>, D. Matthew Walentiny<sup>b</sup>, Patrick M. Beardsley<sup>b,c</sup>

6

7 **INSTITUTIONAL AFFILIATIONS:**

8 <sup>a</sup>Now at Behavioral Pharmacology Research Unit, Department of Psychiatry and Behavioral  
9 Sciences, Johns Hopkins School of Medicine, Baltimore MD, USA

10 <sup>b</sup>Department of Pharmacology & Toxicology, Virginia Commonwealth University School of  
11 Medicine, Richmond, VA, USA

12 <sup>c</sup>Center for Biomarker Research & Precision Medicine, Virginia Commonwealth University  
13 School of Pharmacy, Richmond, VA, USA

14

15 **\*CORRESPONDENCE:**

16 Harrison J. Elder, Ph.D.

17 Behavioral Pharmacology Research Unit,

18 Department of Psychiatry and Behavioral Sciences,

19 Johns Hopkins University School of Medicine,

20 5510 Nathan Shock Dr., Baltimore, MD 21224

21 E-mail address: [helder2@jhmi.edu](mailto:helder2@jhmi.edu) (H.J.E.)

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34

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37

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39 Fentanyl; methamphetamine; isomer; respiratory depression; monoamine receptors; co-use

40

41 **ABBREVIATIONS:**

42 fentanyl (FENT); saline (SAL); minute volume (MVb); Whole-Body Plethysmography (WBP);  
43 opioid-induced respiratory depression (OIRD).

44 **ABSTRACT:**

45 **Rationale:** Fentanyl remains the primary cause of fatal overdoses, and its co-use with  
46 methamphetamine (METH) is a growing concern. We previously demonstrated that racemic  
47 METH can either enhance or mitigate opioid-induced respiratory depression (OIRD) dependent  
48 upon whether a low or high dose is administered. The optical isomers of METH,  
49 dextromethamphetamine (*d*-METH) and levomethamphetamine (*l*-METH), differ substantially in  
50 dose expression and thus may differentially contribute to the bidirectional effects of the  
51 racemate. Furthermore, it is unknown which of METH's monoamine (MA) receptor mechanisms  
52 mediate these respiratory effects. Thus, systematic evaluation of monoamine receptor selective  
53 agents may identify treatment targets for OIRD.

54 **Methods:** The two optical isomers of METH, *d*-METH and *l*-METH, were tested in adult male  
55 mice to determine their effects on basal and fentanyl-depressed minute volume (MVb; i.e.,  
56 respiratory frequency x tidal volume) using whole-body plethysmography. Next, six selective  
57 agonists at MA receptors involved in METH's activity [phenylephrine (PNE;  $\alpha_1$ ), clonidine  
58 (CLON;  $\alpha_2$ ), SKF-82958 (SKF; D<sub>1</sub>), quinpirole (QPR; D<sub>2</sub>), 8-OH-DPAT (8-OH; 5HT<sub>1A</sub>), and DOI  
59 (5HT<sub>2</sub>)] were singly tested on MVb, and then if stimulatory, in combination with fentanyl.

60 **Results:** *d*-METH elevated MVb and *l*-METH decreased MVb. Under fentanyl-depressed  
61 conditions, the bidirectional effects of racemic METH were recreated by *d*-METH while *l*-METH  
62 significantly exacerbated OIRD at 1.0 and 3.0 mg/kg. MVb was dose-dependently increased by  
63 PNE and SKF and decreased by CLON and QPR. Neither 8-OH nor DOI altered basal MVb.  
64 Under fentanyl-depressed conditions, SKF transiently elevated MVb, while PNE more  
65 persistently increased it. Interestingly, DOI transiently increased depressed MVb, while 8-OH  
66 decreased MVb further.

67 **Conclusions:** *d*-METH and *l*-METH differentially contribute to the bidirectional respiratory  
68 modulation observed with the racemate. Selective activation of MA receptors alters basal  
69 respiration and OIRD, with D<sub>1</sub> and α<sub>1</sub> receptors representing potential targets as respiratory  
70 stimulants, whereas α<sub>2</sub>, D<sub>2</sub>, and 5HT<sub>1A</sub> receptors may mediate the exacerbation of OIRD by  
71 METH.

## 72 **1. Introduction**

73 The co-use of fentanyl with stimulants, particularly methamphetamine (METH), has  
74 signaled a new emerging fourth phase of the overdose epidemic (Friedman and Shover, 2023).  
75 From 2013 to 2019, deaths involving stimulants increased 317% (from 1.2 to 5.0 per 100,000),  
76 second only to synthetic opioids over the same period. Notably, overdose due to stimulant and  
77 synthetic opioid co-use showed the largest relative increase compared to stimulants, prescription  
78 opioids, or heroin alone (Cano and Huang, 2021; Mattson et al., 2021). These observations  
79 underscore the need to evaluate how this polydrug abuse affects life-sustaining drug-affected  
80 physiological processes such as respiration.

81 Evidence published in the scientific literature by our laboratory and others (Cruickshank  
82 and Dyer, 2009; Elder et al., 2023a; Hassan et al., 2016; Mendelson et al., 2006; Richards et al.,  
83 1995) demonstrates that amphetamine-type stimulants such as methamphetamine (METH) affect  
84 respiration primarily by increasing ventilatory frequency. Conversely, previously published  
85 studies from our laboratory in mice showed that METH's effects on respiration are not entirely  
86 stimulatory, exemplified by the presence of mild, yet significant depressant effects on  
87 uncompromised "basal" respiration, which are apparent at lower doses than those that produce  
88 stimulation (Elder et al., 2023a). A similar pattern was observed when combined with fentanyl,  
89 whereby low doses of METH exacerbated opioid-induced respiratory depression (OIRD), but a

90 high dose reversed OIRD (Elder et al., 2023a). These bidirectional effects of METH on  
91 respiratory parameters should be of particular relevance to toxicity caused by nonmedical use  
92 because they are induced by doses that would be expected to produce plasma levels similar to  
93 those achieved in humans (Mendelson et al., 2006; Ortman et al., 2021; Rauhut and Bialecki,  
94 2011). Both the respiratory stimulant and depressant effects of METH have potential  
95 consequences for treating polydrug toxicity and OIRD in that pro-depressant effects may  
96 complicate resuscitation following opioid overdose, and stimulatory effects may be exploited for  
97 the development of opioid receptor-independent analeptics.

98         Ample scientific evidence exists detailing the substantial differences in pharmacology  
99 that exist between METH's two optical isomers (enantiomers), dextromethamphetamine (*d*-  
100 METH) and levomethamphetamine (*l*-METH). Specifically, METH's enantiomers differ greatly  
101 in their overall potency, selectivity for releasing primary monoamines, and pharmacokinetic  
102 parameters, with *d*-METH exhibiting substantially greater overall potency for monoamine  
103 release, selectivity for dopamine (DA) release, and relative effects on serotonin (5-  
104 Hydroxytryptamine or 5-HT) compared to *l*-METH, which acts primarily as a selective releaser  
105 of norepinephrine (NE) (Kuczenski et al., 1995; Rothman et al., 2001; Rothman and Baumann,  
106 2003). These differences in pharmacology can be seen in Table 1 which includes the inhibitory  
107 constants ( $K_i$ ) and  $EC_{50}$  values that represent the potency of the enantiomers of METH to  
108 competitively inhibit DAT, NET, and SERT, and to induce monoamine efflux *in vitro*,  
109 respectively. The large differences in pharmacology and potency between the two enantiomers  
110 lead to marked differences in physiological, subjective, and behavioral effects in both humans  
111 and animals that could be hypothesized to extend to respiratory modulation (Mendelson et al.,  
112 2006; Nishimura et al., 2017). Evaluating the respiratory effects of METH's individual

113 enantiomers not only provides a basis for understanding the monoaminergic determinants of  
114 respiratory modulation, but is also important for translational validity, as the vast majority of  
115 illicit METH consumed globally is in the form of *d*-METH hydrochloride (HCl), while other  
116 illicit amphetamines such as MDMA and amphetamine are primarily racemic (Cunningham et  
117 al., 2013; Losacker et al., 2021; Wang et al., 2015). Based on the existing evidence, it can be  
118 hypothesized that the administration of enantiopure preparations of the two individual isomers  
119 would show a separation of the bidirectional effects produced by the racemate into stimulant and  
120 depressant effects based on their relative potency to release DA, NE, and 5-HT.

121         While the findings from experiments with amphetamine-type stimulants demonstrated  
122 potentially useful respiratory stimulant effects for the reversal of OIRD, such stimulants are  
123 limited in their clinical utility for several reasons. Specifically, amphetamines are themselves  
124 drugs of abuse that are highly addictive, cause neurotoxicity, produce respiratory stimulation at  
125 potentially unsafe doses, and can result in toxic interactions in combination with mu opioid  
126 receptor (MOR) agonists (Ashok et al., 2017; Mark et al., 2004; Volkow et al., 2001).  
127 Extrapolating from METH's primary mechanism of action as an indirect agonist of monoamine  
128 receptors, it is likely that selective activation of individual monoamine receptor targets  
129 differentially mediate its bidirectional effects on respiration. Therefore, it can be hypothesized  
130 that if individual DA, NE, and 5-HT receptor subtypes differentially contribute to the stimulant  
131 or depressant effects of METH on respiration, selective activation of those receptors would be  
132 expected to produce the stimulant or depressant effects.

133         There were two objectives of the present study. First, we wanted to evaluate whether  
134 METH's enantiomers differentially contribute to racemic METH's previously observed  
135 bidirectional effects on basal and fentanyl-depressed respiration using whole-body

136 plethysmography (WBP) methodology utilized in earlier studies by our lab, including the  
 137 aforementioned experiments with racemic METH and fentanyl (Elder et al., 2023a, 2023b).  
 138 Should the enantiomers exhibit differential modulation of respiratory parameters, it may provide  
 139 insight into the physiological targets that mediate the stimulant vs depressant components of the  
 140 racemate and allow their separation. Secondly, we wanted to assess monoamine receptor  
 141 selective agonists to determine whether they altered basal respiration. Agonists that effectively  
 142 elevated basal MVb or were devoid of significant depressant effects were consequentially  
 143 evaluated for their effects on depressed MVb in subjects pretreated with fentanyl. The results  
 144 from these studies would provide insight into which mechanism(s) of METH's pharmacology  
 145 might be involved in toxic vs. potentially therapeutic respiratory outcomes.

146

Target	<i>d</i> -Methamphetamine		<i>l</i> -Methamphetamine	
	K <sub>i</sub> (nM)	EC <sub>50</sub> (nM)	K <sub>i</sub> (nM)	EC <sub>50</sub> (nM)
DAT	114	24.5	4840	416
NET	48	12.3	234	28.5
SERT	2,137	736	14,000	4,640
DAT/NET	2.38	1.99	20.68	14.59
SERT/NET	44.52	59.84	59.83	162.8

151  
 152 **Table 1: Pharmacodynamic Profiles of *d*- and *l*-Methamphetamine for Monoamine Release**  
 153 **and Reuptake Inhibition *in vitro*.** Values given on the left for each enantiomer correspond to  
 154 reuptake inhibition potency as measured by the inhibition constant (K<sub>i</sub>) for each transporter as a  
 155 concentration in nanomolar (nM). Righthand values for each enantiomer correspond to the EC<sub>50</sub>  
 156 values for monoamine release in nanomolar (nM). Values for DAT/NET and SERT/NET rows  
 157 represent the selectivity of each individual enantiomer for reuptake inhibition or release as a ratio  
 158 of the identified receptor affinities and EC<sub>50</sub>'s, respectively. Data adapted from (Rothman and  
 159 Baumann, 2003).

Phenylephrine		Clonidine		SKF-82958		Quinpirole		8-OH-DPAT		DOI	
Target Receptor	Affinity K <sub>i</sub> (nM)	Target Receptor	Affinity K <sub>i</sub> (nM)	Target Receptor	Affinity K <sub>i</sub> (nM)						
<b>α<sub>1</sub></b>	<b>100 - 370</b>	α <sub>1</sub>	501	<b>D<sub>1</sub></b>	<b>4.56</b>	D <sub>1</sub>	1,000	<b>5HT<sub>1A</sub></b>	<b>0.65</b>	5HT <sub>1A</sub>	2,355
α <sub>2</sub>	1253 - 1467	<b>α<sub>2</sub></b>	<b>27 - 41</b>	D <sub>2</sub>	264	<b>D<sub>2</sub></b>	<b>47 - 204</b>	5HT <sub>7</sub>	39-251	<b>5HT<sub>2A</sub></b>	<b>0.79 - 14.5</b>
				D <sub>3</sub>	n.d.	<b>D<sub>3</sub></b>	<b>24.35</b>			<b>5HT<sub>2B</sub></b>	<b>26.84</b>
				D <sub>4</sub>	n.d.	<b>D<sub>4</sub></b>	<b>52.7</b>			<b>5HT<sub>2C</sub></b>	<b>3.01 - 60</b>

161

162 **Table 2: Receptor Selectivity and Binding Profiles of Selected Monoamine Agonists.** Values  
 163 given are inhibitory constants (K<sub>i</sub>) with units of nanomolar (nM) derived from ligand  
 164 displacement studies with selected agonists at related receptors. Receptor targets and K<sub>i</sub> values in  
 165 bold correspond to the target of interest. Receptor-agonist pairings for which no reliable data was  
 166 available are indicated by “n.d.”. Data adapted from the following studies (Andersen et al., 1985;  
 167 Boess and Martin, 1994; Borsini et al., 1995; Boundy et al., 1993; Boyajian and Leslie, 1987;  
 168 Campiani et al., 1998; Egan et al., 1998; Lawler et al., 1999; Lovenberg et al., 1993; Nelson et  
 169 al., 1999; Neumeyer et al., 2003; Sokoloff et al., 1990; Sprouse et al., 2004; Van Tol et al.,  
 170 1991).

171

## 172 2. Materials and Methods

### 173 2.1. Materials

174 *d*-Methamphetamine hydrochloride [(2*S*)-*N*-methyl-1-phenylpropan-2-amine]] was  
 175 provided by the National Institute on Drug Abuse (Bethesda, MD, USA) Drug Supply Program.  
 176 *l*-Methamphetamine hydrochloride (Catalogue # 13998; (2*R*)-*N*-methyl-1-phenylpropan-2-  
 177 amine; Cayman Chemical, Ann Arbor, MI, USA), Fentanyl citrate (#F3886; *N*-phenyl-*N*-[1-(2-  
 178 phenylethyl)piperidin-4-yl]propenamide; Sigma-Aldrich, Inc., St. Louis, MO, USA),  
 179 phenylephrine (#P6126; 3-[(1*R*)-1-hydroxy-2-(methylamino)ethyl]phenol; Sigma-Aldrich, Inc.),  
 180 clonidine (Catalogue #1140407; *N*-(2,6-dichlorophenyl)-4,5-dihydro-1*H*-imidazol-2-amine; U.S.  
 181 Pharmacopeia (USP, North Bethesda, MD, USA), SKF-82958 (#HY-10435A; 9-chloro-5-

182 phenyl-3-prop-2-enyl-1,2,4,5-tetrahydro-3-benzazepine-7,8-diol; MedChemExpress LLC,  
183 Monmouth Junction, NJ, USA), quinpirole (#Q102; (4aR,8aR)-5-propyl-1,4,4a,6,7,8,8a,9-  
184 octahydropyrazolo[3,4-g]quinoline; Sigma-Aldrich, Inc.), 8-OH-DPAT (#H8520; 7-  
185 (dipropylamino)-5,6,7,8-tetrahydronaphthalen-1-ol; Sigma-Aldrich, Inc.); and DOI (Catalogue  
186 #13885; 1-(4-iodo-2,5-dimethoxyphenyl)propan-2-amine; Cayman Chemical), were obtained  
187 commercially. All drugs were prepared in saline, sterilized by filtration through 0.2 µm filtration  
188 disks, and administered s.c. at a volume of 10 ml/kg body weight.

## 189 2.2. Subjects

190 Adult male mice [Swiss Webster, CFW(SW), Charles River Laboratories International,  
191 Raleigh, NC, USA] weighing approximately 25–50 g at the time of testing were housed four  
192 subjects per cage in Association for Assessment and Accreditation of Laboratory Animal Care-  
193 accredited facilities. Mice had *ad libitum* access to food (Teklad 7012 Rodent Diet; Envigo,  
194 Madison, WI, USA) and tap water. Vivaria were maintained at 22°C ± 2°C and 45%–50%  
195 humidity, with lights set to a reverse 12-h light/dark cycle (lights off at 10:00). All tests were  
196 conducted on weekdays during the dark period between 11:00 and 17:00 to ensure mice were  
197 active (i.e., not asleep). All subjects were acclimated to the vivarium for at least one week before  
198 the commencement of studies and were experimentally and drug-naive before testing. Subjects  
199 were tested once and were not used for any subsequent tests to preclude drug or testing history  
200 effects. All procedures were carried out in accordance with the National Research Council's  
201 Guide for Care and Use of Laboratory Animals (2011). This experimental protocol was approved  
202 by the Institutional Animal Care and Use Committee at Virginia Commonwealth University.

## 203 2.3. Apparatus

204 Mice were tested using whole-body plethysmograph devices (FinePointe WBP Chamber with  
205 Halcyon Technology, Data Sciences International, St. Paul, MN, USA) while unrestrained and  
206 allowed free movement in individual isolated experimental vessels. Experimental vessels (0.5 L  
207 volume with adjustable 0.5L/min room air bias flow) were housed in a laboratory illuminated by  
208 custom 660 nm-emitting T8-style ceiling-mounted light tubes each with 96, 0.2-watt Epistar  
209 2835 SMD LEDs (Shenzhen Benwei Electronics Co., Ltd., Longhua District, Shenzhen, China).  
210 This wavelength is minimally visible to mice (Peirson et al., 2018) which enabled maintenance  
211 of subjects in the dark phase of their activity cycle during testing. These testing conditions have  
212 been used previously by our lab to accurately measure the respiratory effects of both stimulant  
213 and depressant drugs in mice (Elder et al., 2023a, 2023b). For the experiments described here  
214 ambient room air was used for bias flow inputs to experimental vessels rather than the gas  
215 mixture of 5% CO<sub>2</sub>, 21% O<sub>2</sub> with balanced N<sub>2</sub> used in previous studies. Ambient room air was  
216 utilized to create normocapnic conditions for all experiments in order to maximize face validity  
217 and increase the translational capacity of results. Respiratory rate (Freq), tidal volume (TVb),  
218 and minute volume (MVb) were recorded using software (FinePointe Software Research Suite;  
219 Data Sciences International).

#### 220 *2.4. Three-phase WBP Protocol*

221 WBP testing for all treatment conditions was carried out using the three-phase protocol  
222 described previously (Elder et al., 2023a) for assessing drug effects on basal and opioid-  
223 depressed respiration. The present study consisted of two stages, with the first dedicated to  
224 evaluating the differential effects of METH isomers on basal- and fentanyl-depressed MVb, and  
225 the second involving the systematic evaluation of monoamine agonists for their ability to affect  
226 basal and fentanyl-depressed respiration.

227 In stage one of this study, the two optical isomers (enantiomers) of METH, *d*-METH and  
228 *l*-METH, were tested under basal and fentanyl-depressed conditions following a procedure that  
229 was identical to those described in (Elder et al., 2023a) for tests with *d*-amphetamine and  
230 racemic METH, aside from the change in gas mixture. Both *d*- and *l*-METH were evaluated  
231 under both basal and depressed conditions at the same nominal doses as those used for tests of  
232 racemic METH (1.0, 3.0, 10 mg/kg) to evaluate whether the effects of either enantiomer differed  
233 from those reported originally with the racemate and to determine their individual contributions  
234 to its effects. An additional test with a higher dose of *l*-METH (30 mg/kg) was conducted to  
235 evaluate potency differences across an extended dose range. For all experiments under basal  
236 conditions, saline was administered prior to Phase II (basal conditions), followed by a dose of  
237 either *d*- or *l*-METH as the ‘test compound’ prior to the start of Phase III. Control groups  
238 received three saline injections (vehicle), with one injection administered prior to the initiation of  
239 Phases I, II, and III, respectively. Experiments under depressed conditions consisted of treatment  
240 with fentanyl (0.3 mg/kg s.c.) administered prior to Phase II. This dose of fentanyl was selected  
241 because it consistently produces MVb depression of approximately 50% from baseline at the  
242 beginning of Phase III. Additionally, 0.3 mg/kg fentanyl has been employed consistently across  
243 studies in our laboratory as the standard dose for tests on fentanyl-depressed conditions because  
244 it reproduces fentanyl’s bidirectional effects on depressed MVb.

245 Experiments with monoamine agonists in stage two consisted of an initial evaluation in  
246 which six agonists selective at different monoamine receptors involved in METH’s activity  
247 (Bolme et al., 1974; Corcoran et al., 2014; Desai et al., 2005; Eilam and Szechtman, 1989;  
248 Guenther et al., 2009; Jaster et al., 2022; Stone et al., 2014; Zarrindast et al., 2002) were  
249 evaluated using the three-phase protocol. The six monoamine agonists chosen to selectively

250 activate monoamine receptors of interest were phenylephrine (PNE;  $\alpha_1$ ), clonidine (CLON;  $\alpha_2$ ),  
251 SKF-82958 (SKF; D<sub>1</sub>), quinpirole (QPR; D<sub>2</sub>), 8-OH-DPAT (8-OH; 5HT<sub>1A</sub>), and DOI (5HT<sub>2</sub>).  
252 The affinities of each agonist at their respective receptor targets are given in Table 2. For all tests  
253 during an initial evaluation, saline was administered prior to Phase II (basal conditions),  
254 followed by a dose of a monoamine agonist as the ‘test compound’ prior to the start of Phase III.  
255 Control groups received three saline injections (vehicle), with one injection administered prior to  
256 the initiation of Phases I, II, and III, respectively. Phenylephrine (0.3, 1.0, 10 mg/kg s.c.),  
257 clonidine (0.03, 0.1, 1.0 mg/kg s.c.), SKF-82958 (0.1, 0.3, 1.0 mg/kg s.c.), quinpirole (0.3, 1.0,  
258 3.0 mg/kg s.c.), 8-OH-DPAT (0.01, 0.1, 0.3 mg/kg s.c.), and DOI (0.1, 1.0, 3.0 mg/kg s.c.) were  
259 screened under basal conditions to determine their ability to elevate eupneic MVb. Results from  
260 initial tests of basal respiration were used to identify the maximally effective dose for stimulating  
261 MVb if MVb elevation occurred. In the second evaluation stage, doses of each monoamine  
262 agonist that produced the greatest elevation of MVb under basal conditions or the highest two  
263 doses of inactive compounds were selected for subsequent testing under fentanyl-depressed  
264 conditions for their ability to modulate MVb depression. Depressed conditions in this stage  
265 consisted of treatment with fentanyl (0.3 mg/kg s.c.) administered prior to Phase II. Additional  
266 doses of monoamine agonists were evaluated under fentanyl-depressed conditions if the initial  
267 dose produced a significant reversal of depressed MVb in order to determine the dose-  
268 responsiveness of MVb elevation. Compounds that decreased basal respiration were excluded  
269 from subsequent tests under fentanyl-depressed conditions.

## 270 2.5. Statistical Analysis

271 The primary dependent measure, normalized MVb, was expressed as a percentage of baseline  
272 MVb collected during Phase I. Normalized group MVb data from time-course tests were

273 analyzed using the methodology that has been described in previous publications for determining  
274 the effect of treatment conditions (Elder et al., 2023a). Complete reversal of fentanyl respiratory  
275 depression was defined as occurring when treatment groups did not differ significantly from  
276 vehicle control at a respective time point(s) during Phase III. Partial reversal was defined as an  
277 increase in the MVb of a treatment group following the initiation of Phase III but that remained  
278 significantly lower than that of vehicle controls. Significant respiratory stimulant effects were  
279 considered to occur when a treatment group had MVb values that were significantly greater than  
280 fentanyl-treated controls at any time point in Phase III, regardless of level of reversal. Raw MVb  
281 values (ml/min) from Phase I were analyzed using one-way ANOVAs to determine if between-  
282 group differences existed at baseline, followed by a Holm-Šídák multiple comparisons test  
283 comparing all treatment groups if a significant group effect was detected. Area under the curve  
284 (AUC) calculations were conducted to summarize the overall influence of treatment on  
285 normalized MVb over the entirety of Phase III after agonist administration ( $t = 20 - 80$ ). AUC  
286 data were analyzed via one-way ANOVA, and significant treatment effects were followed by  
287 Holm-Šídák multiple comparisons tests to detect differences between individual treatment  
288 groups and vehicle or fentanyl-treated controls. All analyses were performed using software  
289 (GraphPad Prism 9 for Macintosh; GraphPad Software, San Diego, CA, USA) and statistical  
290 significance for all analyses was set at a level of  $\alpha = 0.05$ .

### 291 **3. Results**

#### 292 *3.1. Differential effects of methamphetamine enantiomers on basal and depressed respiration*

293 The dose-dependent effects of *d*-METH (1.0, 3.0, 10 mg/kg) on basal MVb in subjects  
294 who received saline prior to Phase II are shown in Figure 1A. Administration of *d*-METH  
295 significantly affected MVb [ $F(48, 444) = 10.10$ ;  $p < 0.0001$ ], producing dose-dependent

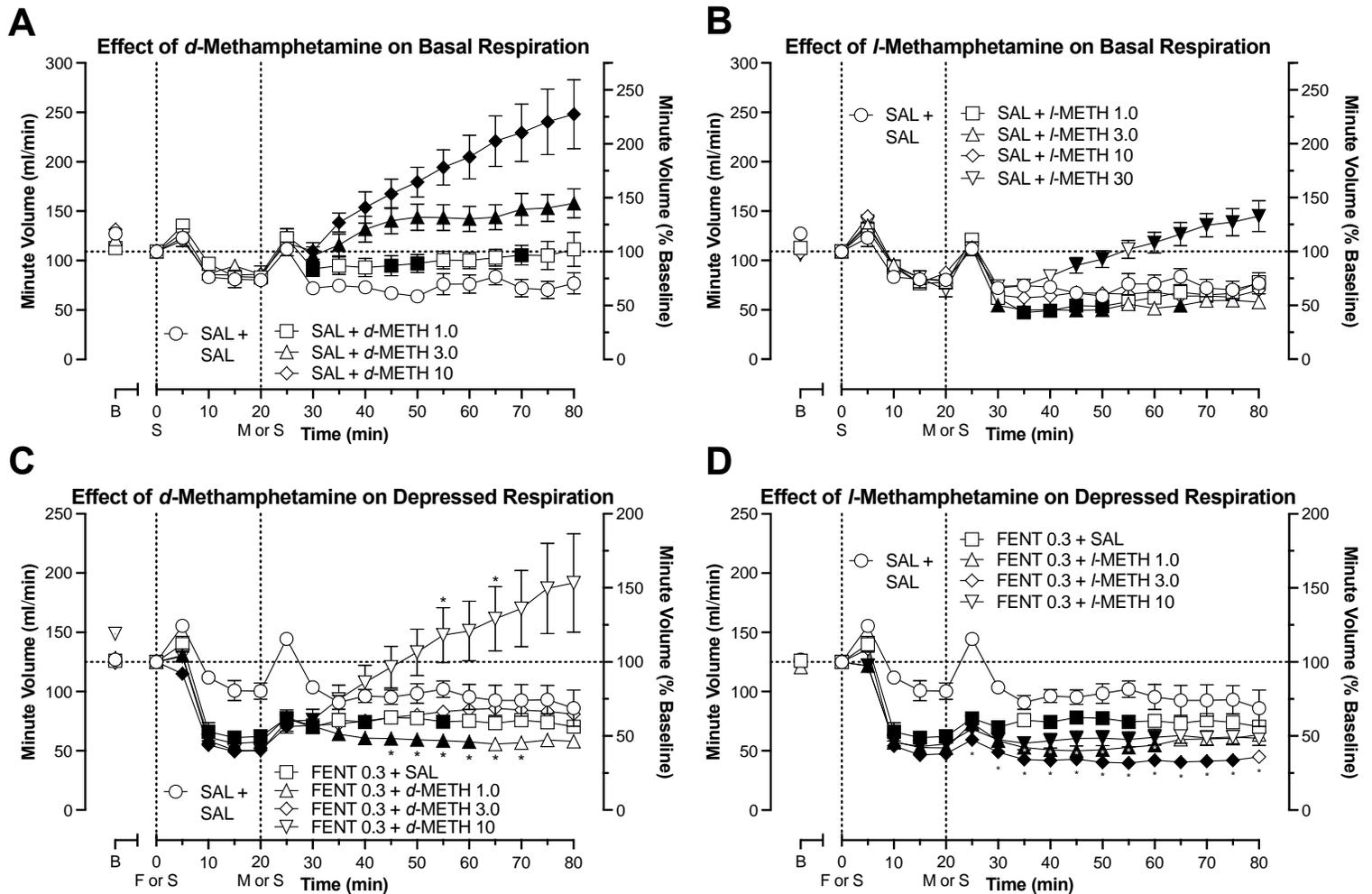
296 elevations of MVb that were significantly ( $p < 0.05$ ) greater than saline controls at one or more  
297 time points post-administration for all doses tested. All doses of *d*-METH significantly increased  
298 MVb compared to saline controls within 10 min of administration, after which MVb in subjects  
299 who received intermediate (3.0 mg/kg) and high (10 mg/kg) doses continued to increase,  
300 eventually reaching peak values at 60 min post-administration of 144.9% ( $p = 0.0020$ ) and  
301 227.5% ( $p = 0.0029$ ) of baseline, respectively. The results of experiments with *l*-METH (Figure  
302 1B) indicated that treatment significantly affected basal respiration [ $F(64, 556) = 7.580$ ;  $p <$   
303  $0.0001$ ]. Administration of an intermediate (3.0 mg/kg) dose of *l*-METH under basal conditions  
304 significantly decreased MVb to 49.91% of baseline ( $t = 30$ ,  $p = 0.0101$ ) within 10 min post-  
305 administration, and MVb values remained significantly depressed relative to saline-treated  
306 controls for 20 min until they no longer differed from controls at 35 min post-administration ( $t =$   
307  $55$ ). At the lowest dose (1.0 mg/kg) of *l*-METH basal MVb was similarly depressed within 15  
308 min of administration to 43.33% of baseline ( $t = 35$ ;  $p = 0.0080$ ) which was significantly lower  
309 than controls. Interestingly, administration of a higher dose (10 mg/kg) of *l*-METH did not  
310 significantly alter MVb at any time point after administration, neither stimulating nor depressing  
311 basal respiration. Therefore, a higher dose, 30 mg/kg, was subsequently tested and produced  
312 significant stimulation of a magnitude similar to 3.0 mg/kg *d*-METH but with a more protracted  
313 onset, as seen in the 25 min latency to exert significant effects. The results obtained from this  
314 follow-up test displayed the dose-dependent transition from depressant effects at low doses (1.0,  
315 3.0 mg/kg) to respiratory stimulant effects at high doses (30 mg/kg) that were similar to the  
316 effects of low-moderate doses of *d*-METH.

317           The effects of *d*-METH on respiration that was depressed by the administration of  
318 fentanyl (0.3 mg/kg) are shown in Figure 1C. In fentanyl-pretreated mice, there was a significant

319 effect of *d*-METH over time [ $F(64, 560) = 9.741$ ;  $p < 0.0001$ ] on respiration. In contrast to the  
320 dose-dependent stimulation of MVb observed with *d*-METH under basal conditions,  
321 administration of *d*-METH to subjects that were pretreated with fentanyl (0.3 mg/kg) produced  
322 bidirectional, dose-dependent effects following a similar pattern as was observed with racemic  
323 METH. Specifically, *d*-METH at the lowest dose tested (1.0 mg/kg) had pro-depressant effects,  
324 the highest dose (10 mg/kg) had pronounced stimulating effects, and the intermediate dose (3.0  
325 mg/kg) had no significant effect on fentanyl-depressed MVb. The pro-depressant effects of 1.0  
326 mg/kg *d*-METH were characterized by a 30-min increase in the duration of significant  
327 depression ( $t = 35 - 60$ ) along with MVb values that were significantly lower than fentanyl-  
328 treated controls from 25 – 50 min ( $t = 45 - 70$ ) post-administration. The respiratory stimulant  
329 properties of 10 mg/kg *d*-METH became apparent at 15 min post-administration ( $t = 35$ ) when  
330 MVb values rose slightly above saline-treated controls (74.82 vs 72.74% of baseline,  
331 respectively), constituting a complete reversal of fentanyl-induced depression. Subsequently,  
332 MVb values continued to rise throughout the remainder of Phase III in subjects who received 10  
333 mg/kg *d*-METH, finally reaching a peak of 153.3% of baseline at the final observation point ( $t =$   
334 80), however this increase did not reach statistical significance compared to saline-treated  
335 controls.

336 The effects of *l*-METH (1.0, 3.0, 10 mg/kg) on fentanyl-depressed respiration are shown  
337 in Figure 1D. Analysis of MVb data showed a significant effect of *l*-METH treatment over time  
338 [ $F(64, 560) = 4.534$ ;  $p < 0.0001$ ] on fentanyl-depressed respiration. The effects of *l*-METH  
339 administration under depressed conditions were consistent with results observed under basal  
340 conditions. Specifically, all doses of *l*-METH had pro-depressant effects on MVb that varied  
341 according to an inverted-U-shaped dose-response and were greatest after administration of the

342 intermediate 3.0 mg/kg dose. The pro-depressant effects of *l*-METH on fentanyl-depressed  
343 respiration were characterized by increased duration and magnitude of MVb depression  
344 beginning 10 – 15 min after administration. MVb in subjects who received 3.0 mg/kg *l*-METH  
345 20 min after 0.3 mg/kg fentanyl varied between 31.81 – 39.15% of baseline after onset (t = 30 –  
346 80) and remained significantly depressed compared to saline controls until the penultimate time  
347 point in phase III (t = 75). The highest dose of 10 mg/kg *l*-METH had the least pro-depressant  
348 effect on MVb that were characterized by nonsignificant reductions of 8 – 15% of baseline after  
349 onset (t = 30 min) compared with fentanyl-treated controls at all time points except for one in  
350 which MVb depression was significant (t = 35, p = 0.0259).



351 **Figure 1: Effect of *d*- and *l*-methamphetamine on basal and fentanyl-depressed minute**  
 352 **volume.** A) Dose- and time-effects of *d*-methamphetamine (*d*-METH) and B) *l*-  
 353 methamphetamine (*l*-METH) on basal minute volume following saline (SAL) pretreatment. C)  
 354 Dose- and time-effects of *d*-METH and D) *l*-METH on depressed minute volume following  
 355 pretreatment with 0.3 mg/kg fentanyl (FENT). Left ordinate: mean raw MVb (ml/min) indexing  
 356 values of symbols only during baseline (B) of Phase I. Right ordinate: normalized (percent  
 357 baseline) MVb indexing values of symbols during the 80-min test session following Phase I  
 358 baseline. These symbols indicate mean MVb expressed as a percentage of baseline MVb of 8  
 359 mice per treatment group. Filled symbols indicate a significant ( $p \leq 0.05$ ) difference from SAL +

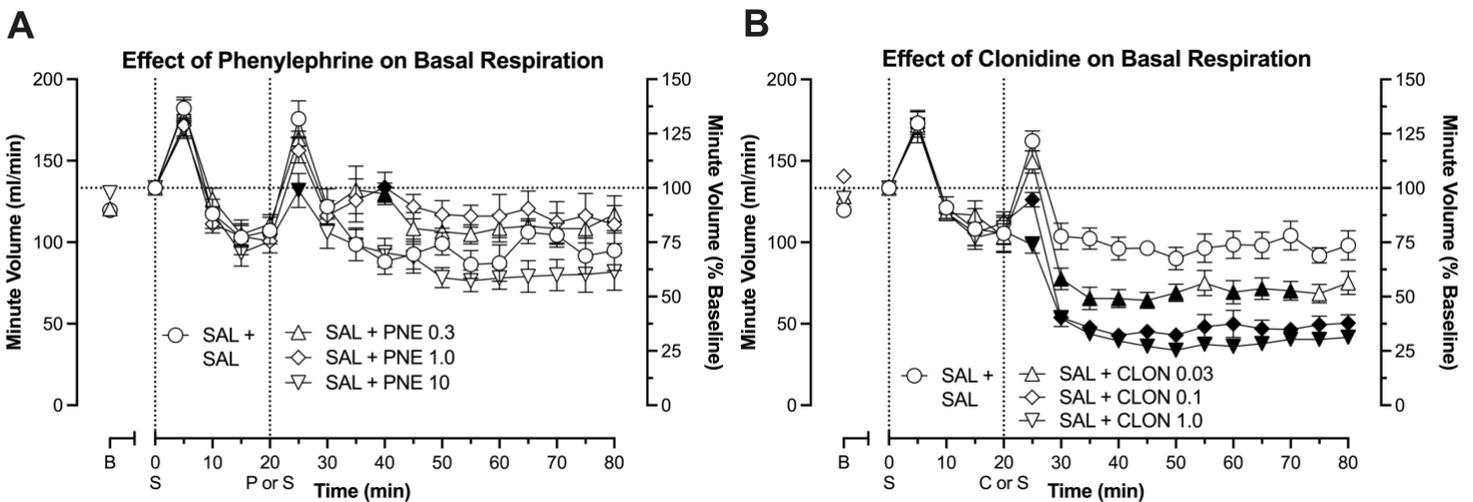
360 SAL treated controls at individual time points. Additional \* symbols above (FENT 0.3 + *d*-  
361 METH 10), below (FENT 0.3 + *d*-METH 1.0; FENT 0.3 + *l*-METH 3.0) or within (FENT 0.3 +  
362 *l*-METH 1.0) specific time points indicate a significant difference at that time point between  
363 individual treatment groups and FENT 0.3 + SAL controls of  $p \leq 0.05$  according to Holm-Šídák  
364 post-hoc comparisons. Abscissa labels: M = *d*- or *l*-METH injection, S = saline injection, F =  
365 fentanyl injection. N = 8 per group. No significant differences were detected at baseline across  
366 experimental conditions when raw MVb values were compared via one-way ANOVA for  
367 experiments presented in panel A) [F(3, 28) = 1.294;  $p = 0.2958$ ], panel C) [F(4, 35) = 1.584;  $p =$   
368 0.2002], or panel D) [F(4, 35) = 0.2669;  $p = 0.8973$ ]. Significant differences in raw MVb means  
369 were detected at baseline for experimental conditions presented in panel B when compared via  
370 one-way ANOVA [F(4, 35) = 2.697;  $p = 0.0465$ ], but no significant differences were detected  
371 between individual groups by subsequent Holm-Šídák multiple comparisons tests.

### 372 3.2. Testing monoamine receptor agonists for their effects on respiration

373 The results from basal experiments with the selective  $\alpha_1$ -receptor agonist phenylephrine  
374 and the selective  $\alpha_2$ -receptor agonist clonidine are shown in Figure 2A and 2B, respectively.  
375 Phenylephrine treatment significantly affected basal MVb over time [F(48, 448) = 1.847;  $p =$   
376 0.0008] that varied as a function of dose according to an inverted-U-shaped relationship.  
377 Administration of low (0.3 mg/kg) and intermediate (1.0 mg/kg) doses of phenylephrine  
378 stimulated respiration elevating MVb values above those of controls by 15 min post-  
379 administration. However, increases in MVb induced by phenylephrine were small in magnitude  
380 and only significantly differed from controls at a single time point ( $t = 40$ ) after administration of  
381 0.3 mg/kg ( $p = 0.0043$ ) and 1.0 mg/kg ( $p = 0.0037$ ). At the highest dose of 10 mg/kg,  
382 phenylephrine's effects on MVb depressed MVb values throughout phase III, although MVb

383 depression was only significant at a single time point ( $t = 25$ ;  $p = 0.0353$ ) compared with  
384 controls.

385 Under basal conditions, clonidine treatment significantly [ $F(48, 448) = 5.550$ ;  $p <$   
386  $0.0001$ ] affected MVb over time, producing potent and long-lasting depressant effects at all  
387 doses tested (0.03, 0.1, 1.0 mg/kg). The depression of basal MVb following administration of  
388 clonidine was rapid and sustained, with decreased onset latency and increased duration of  
389 depression as the dose increased. Both intermediate (0.1 mg/kg) and high (1.0 mg/kg) doses of  
390 clonidine significantly depressed MVb relative to controls within 5 min and remained  
391 significantly depressed thereafter for the entirety of phase III. The magnitude of MVb depression  
392 by clonidine was similarly dose-dependent, with maximum depression of 48.3%, 32.1%, and  
393 25.3% of baseline occurring after administration of 0.03, 0.1, and 1.0 mg/kg, respectively.



394 **Figure 2: Effects of selective  $\alpha_1$  and  $\alpha_2$  adrenergic receptor agonists on basal respiration.**

395 Panel A) Dose- and time-effects of phenylephrine (PNE) and B) clonidine (CLON) on basal  
396 minute volume following saline (SAL) pretreatment. Filled symbols indicate a significant ( $p \leq$   
397  $0.05$ ) difference from SAL + SAL treated controls at individual time points. Abscissa labels: P =

398 phenylephrine injection, C = clonidine injection, S = saline injection. N = 8 per group. No  
399 significant differences were detected at baseline across experimental conditions when raw MVb  
400 values were compared via one-way ANOVA for groups in panel A) [ $F(3, 28) = 0.7559$ ;  $p =$   
401  $0.5283$ ] or B) [ $F(3, 28) = 1.325$ ;  $p = 0.2859$ ]. All other details are the same as in Figure 1.

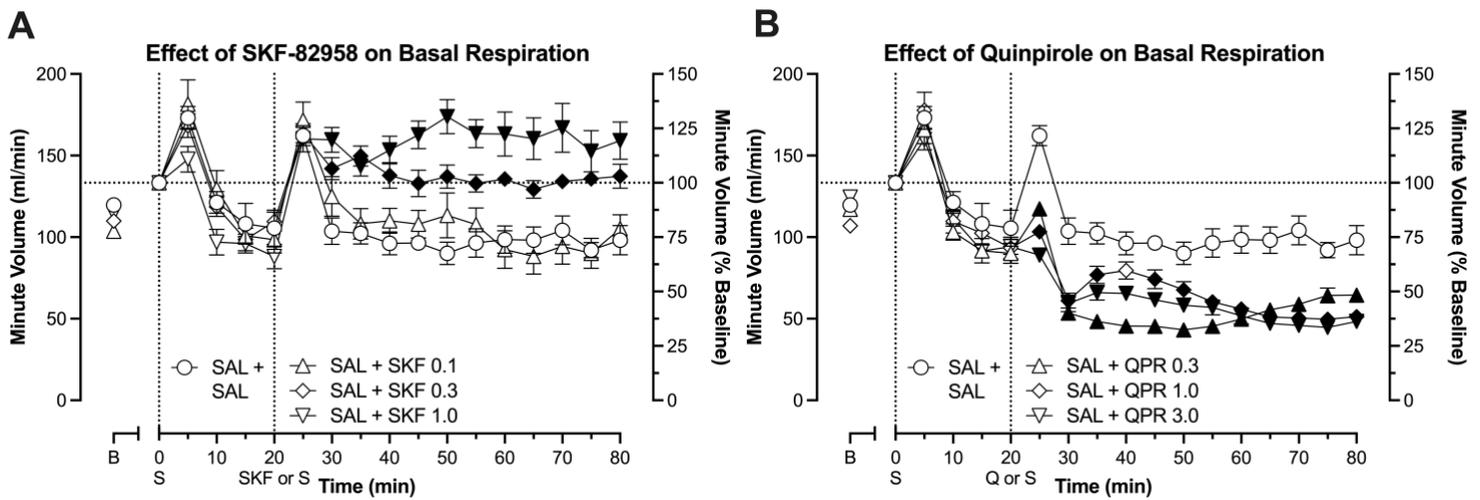
402         The results from tests under basal conditions with the selective D<sub>1</sub>-receptor agonist SKF-  
403 82958 and the selective D<sub>2</sub>-receptor agonist quinpirole are shown in Figures 3A and 3B,  
404 respectively. SKF-82958 treatment had significant effects on basal MVb over time [ $F(48, 448) =$   
405  $8.112$ ;  $p < 0.0001$ ] that were dose-dependent. Administration of intermediate (0.3 mg/kg) and  
406 high (1.0 mg/kg) doses of SKF-82958 significantly stimulated respiration, elevating MVb values  
407 above those of controls within 10 min post-administration, after which they remained  
408 significantly elevated for the rest of the session. The magnitude of MVb elevation by  
409 intermediate and high doses of SKF was dose-dependent, achieving maximum MVb values of  
410 122.8% and 130.4% of baseline after onset ( $t \geq 25$ ), respectively. At the lowest dose of 0.1  
411 mg/kg, SKF-82958's effects on MVb were nonsignificant compared with controls, and produced  
412 only modest increases ( $< 10\%$ ) in MVb throughout phase III.

413         Under basal conditions, quinpirole treatment significantly [ $F(48, 448) = 5.861$ ;  $p <$   
414  $0.0001$ ] affected MVb over time, producing complex and sustained depressant effects at all doses  
415 tested (0.03, 1.0, 3.0 mg/kg). The depression of basal MVb following administration of  
416 quinpirole was rapid and significant at all doses. Quinpirole's effects on MVb were characterized  
417 by a substantial initial decrease immediately after administration, followed by effect profiles that  
418 varied with dose. Both intermediate (1.0 mg/kg) and high (3.0 mg/kg) doses of quinpirole  
419 displayed a complex modulation of respiration over the course of phase III characterized by a  
420 period of rapid recovery and then gradual reduction in MVb. In comparison, the lowest dose (0.3

421 mg/kg) displayed a more typical pattern of depression and gradual recovery. Maximum  
 422 depression to 32.42%, 37.28%, and 33.47% of baseline occurred after administration of 0.3, 1.0,  
 423 and 3.0 mg/kg, respectively. Time to peak depression differed between the lowest (30 min) and  
 424 higher two doses (55 min).

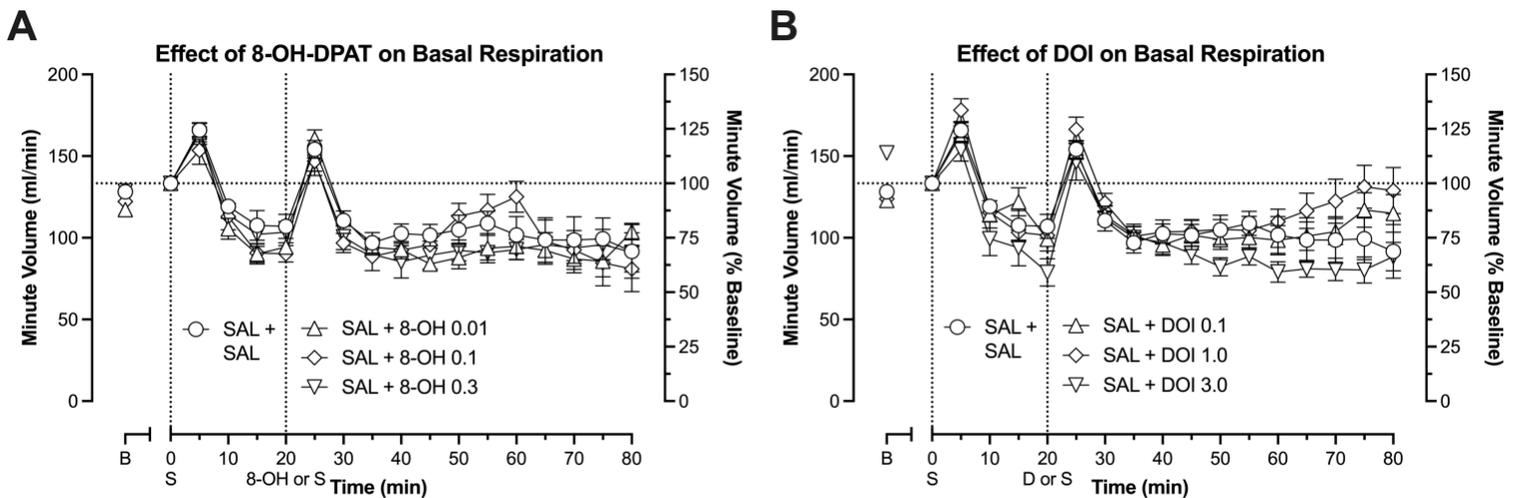
425 **Figure 3: Effects of selective dopamine D<sub>1</sub> and D<sub>2</sub> receptor agonists on basal respiration.**

426 Panel A) Dose- and time-effects of SKF-82958 (SKF) and B) quinpirole (QPR) on basal minute



427 volume following saline (SAL) pretreatment. Filled symbols indicate a significant ( $p \leq 0.05$ )  
 428 difference from SAL + SAL treated controls at individual time points. Abscissa labels: SKF =  
 429 SKF-82958 injection, Q = quinpirole injection, S = saline injection. N = 8 per group. No  
 430 significant differences were detected at baseline across experimental conditions when raw MVb  
 431 values were compared via one-way ANOVA for groups in panel A) [ $F(3, 28) = 1.199$ ;  $p =$   
 432  $0.3281$ ] or B) [ $F(3, 28) = 1.921$ ;  $p = 0.1491$ ]. All other details are the same as in Figure 1.

433 The effects of selective 5HT<sub>1a</sub>-receptor agonist 8-OH-DPAT and the selective 5HT<sub>2</sub>-  
 434 receptor agonist DOI are shown in Figure 4A and 4B, respectively. No significant effect of 8-  
 435 OH-DPAT treatment on basal MVb over time was detected [F(48, 448) = 1.269; p = 0.1147].  
 436 Minor fluctuations in MVb occurred over the course of phase III, with low and high doses of 8-  
 437 OH-DPAT tending to lower MVb relative to controls, but post-hoc analyses of differences in  
 438 MVb by time point could not be completed due to the lack of significant interaction of treatment  
 439 and time. Conversely, a significant interaction of time and treatment [F(48, 448) = 1.452; p =  
 440 0.03] was detected following administration of DOI (0.1, 1.0, 3.0 mg/kg). Treatment with DOI at  
 441 intermediate (1.0 mg/kg) and high (3.0 mg/kg) doses affected MVb gradually over phase III and  
 442 peaked towards the end of the session. DOI at 1.0 mg/kg nonsignificantly increased MVb  
 443 relative to controls between 40- and 60-min post-administration (t = 60 – 80), while a dose of 3.0  
 444 mg/kg tended to decrease MVb to a nonsignificant degree beginning at 25 min post-  
 445 administration (t = 45).

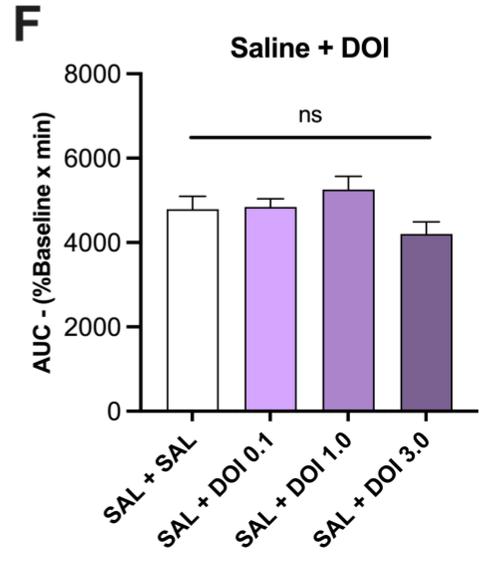
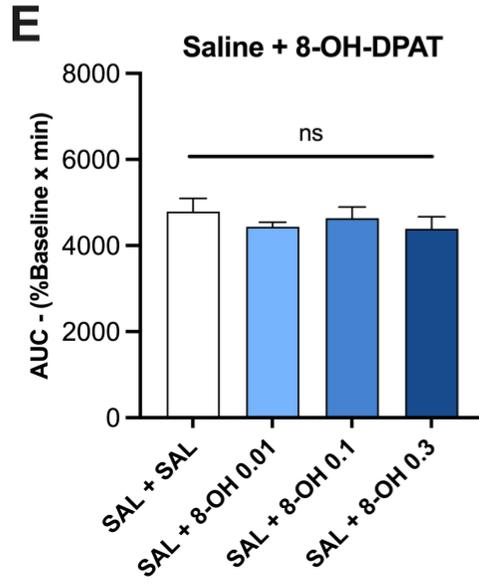
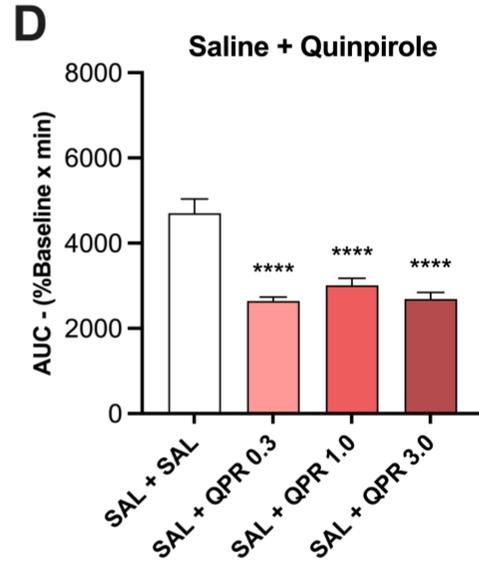
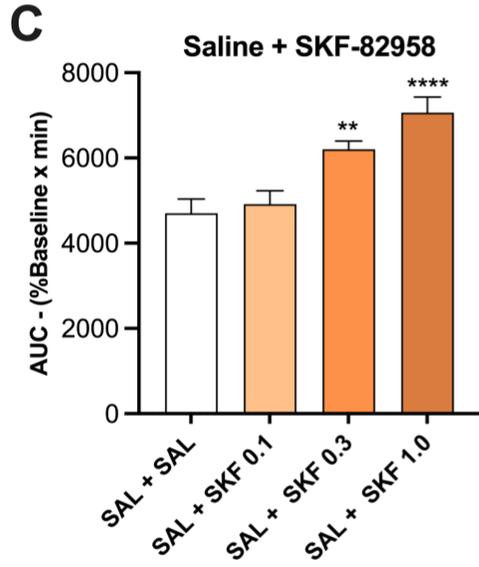
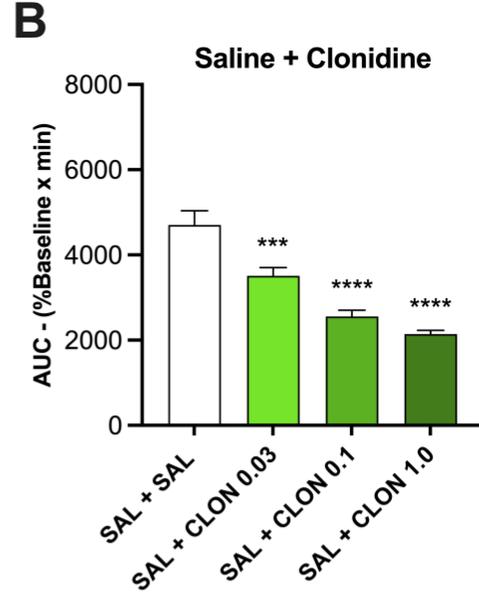
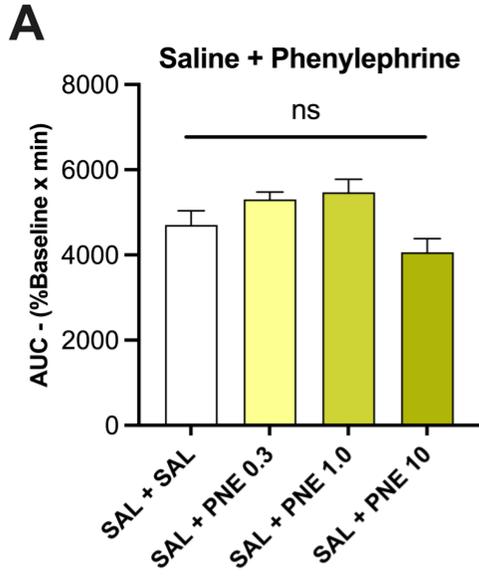


446 **Figure 4: Effects of selective 5HT<sub>1a</sub> and 5HT<sub>2</sub> serotonin receptor agonists on basal**  
 447 **respiration.** Panel A) Dose- and time-effects of 8-OH-DPAT (8-OH) and B) DOI (DOI) on  
 448 basal minute volume following saline (SAL) pretreatment. Filled symbols indicate a significant (p

449  $\leq 0.05$ ) difference from SAL + SAL treated controls at individual time points. Abscissa labels: 8-  
450 OH = 8-OH-DPAT injection, D = DOI injection, S = saline injection. N = 8 per group. No  
451 significant differences were detected at baseline across experimental conditions when raw MVb  
452 values were compared via one-way ANOVA for groups in panel A) [ $F(3, 28) = 1.180$ ;  $p =$   
453  $0.3350$ ] or B) [ $F(3, 28) = 2.694$ ;  $p = 0.0651$ ]. All other details are the same as in Figure 1.

454         The results of post-hoc analyses of monoamine agonist treatment effects on the basal area  
455 under the curve (AUC) of normalized MVb across time throughout phase III are shown in Figure  
456 5. AUC analysis of the effect of adrenergic agonist treatments demonstrated that phenylephrine  
457 significantly affected MVb AUC [ $F(3, 28) = 4.840$ ;  $p = 0.0077$ ] increasing AUC (non-  
458 significantly) relative to saline controls in an inverted-U-shaped dose-response relationship  
459 (Figure 5A). Conversely, clonidine had pronounced dose-dependent depressant effects on MVb  
460 [ $F(3, 28) = 29.23$ ;  $p < 0.0001$ ] over the course of phase III (Figure 5B) exemplified by significant  
461 reductions in AUC relative to controls at all doses ( $p \leq 0.0004$ ). Figures 5C and 5D show that  
462 treatment with the selective dopaminergic D<sub>1</sub>-like receptor agonist SKF-82958 and the D<sub>2</sub>-like  
463 receptor agonist quinpirole affected AUC in a subtype-specific manner similar to the adrenergic  
464 agonists, whereby SKF-82958 dose-dependently elevated [ $F(3, 28) = 13.16$ ;  $p < 0.0001$ ], and  
465 quinpirole decreased [ $F(3, 28) = 22.25$ ;  $p < 0.0001$ ] AUC relative to controls. Maximal effects of  
466 SKF-82958 on AUC were seen after treatment with the highest dose (1.0 mg/kg), which  
467 significantly increased AUC ( $p < 0.0001$ ) relative to controls. Quinpirole consistently and  
468 significantly decreased AUC ( $p < 0.0001$ ) relative to controls. As expected, neither 8-OH-DPAT  
469 [ $F(3, 28) = 0.5466$ ,  $p = 0.6545$ ] (Figure 5E) nor DOI [ $F(3, 28) = 2.418$ ,  $p = 0.0872$ ] (Figure 5F)  
470 had main effects on AUC nor were any significant changes detected compared to respective  
471 saline-treated controls at any dose tested. However, the nonsignificant trends present in basal

472 time course data were apparent, with DOI tending to increase slightly, and 8-OH-DPAT tending  
473 to decrease slightly, basal AUC relative to controls.



475 **Figure 5: Area Under the Curve summary analysis of the effects on Minute Volume during**  
476 **phase III by treatment.** Panel A) Dose-effects of phenylephrine (PNE); B) clonidine (CLON);  
477 C) SKF-82958 (SKF); D) quinpirole (QPR); E) 8-OH-DPAT (8-OH); and F) DOI on area under  
478 the curve (AUC) of normalized minute volume x time in saline (SAL) pretreated subjects during  
479 phase III (60 min). Abscissa labels correspond to injections given at t = 0 and t = 20, with saline  
480 identified as SAL and numbers corresponding to the dose administered in mg/kg. AUC is given  
481 on the ordinate as the product of % baseline x minutes (min). \*\*, \*\*\*, \*\*\*\* above bars indicate a  
482 significant difference between individual treatment groups and SAL + SAL controls of  $p \leq 0.01$ ;  
483 0.001; 0.0001, respectively, while “ns” above bars indicates nonsignificant differences when  
484 analyzed via a one-way ANOVA followed by Holm-Šidák post-hoc comparisons.

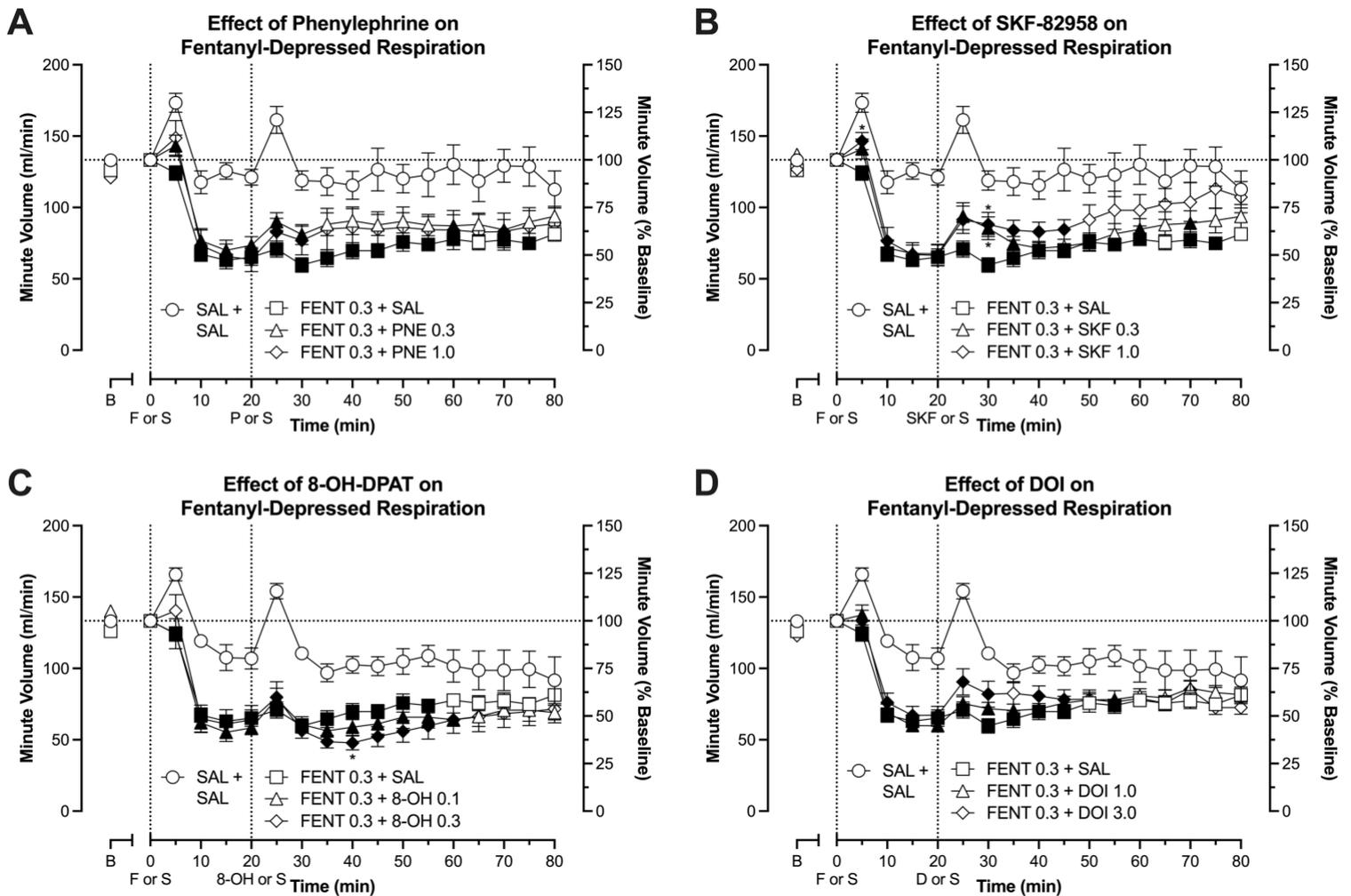
### 485 *3.3. Evaluation of active monoamine agonist effects on fentanyl-depressed respiration*

486 Based on the results obtained under basal conditions, four monoamine agonists were  
487 selected for further tests under fentanyl-depressed conditions based on either: 1) their ability to  
488 elevate basal MVb at one or more doses (phenylephrine and SKF-82958); or 2) lack of  
489 significant depression of basal MVb in conjunction with published evidence supporting efficacy  
490 of either target receptor activation or selected agonist under opioid-depressed conditions (8-OH-  
491 DPAT and DOI) (Corcoran et al., 2014; Guenther et al., 2009; Lalley et al., 1995; Onimaru et al.,  
492 1998; Stettner et al., 2008). The results of experiments with selected agonists under fentanyl-  
493 depressed conditions are shown in Figure 6. Administration of phenylephrine at two doses (0.3,  
494 1.0 mg/kg) following pretreatment with fentanyl (0.3 mg/kg) significantly affected MVb, with a  
495 main effect of treatment condition x time (Figure 6A;  $[F(48, 448) = 2.807; p < 0.0001]$ ). Subjects  
496 that received fentanyl (0.3 mg/kg) had significantly ( $p < 0.0001$ ) depressed MVb values relative  
497 to saline-treated controls that were between 47.6 and 55.2% of baseline at the time of

498 phenylephrine administration. Both doses of phenylephrine completely reversed MVb depression  
499 within 15 min ( $t = 35$ ) of administration, at which point phenylephrine-treated groups had MVb  
500 values of 66.2 and 63.7% of baseline, respectively. The results of experiments with two doses of  
501 SKF-82958 (0.3, 1.0 mg/kg) under fentanyl-depressed conditions are shown in Figure 6B.  
502 Analysis showed a significant main effect of treatment condition  $\times$  time [ $F(48, 448) = 3.333$ ;  $p <$   
503  $0.0001$ ] on MVb. Administration of both 0.3 mg/kg and 1.0 mg/kg SKF-82958 to fentanyl-  
504 pretreated subjects significantly increased MVb relative to fentanyl-treated controls within 10  
505 min ( $t = 30$ ) but failed to achieve complete reversal. MVb values in subjects treated with 1.0  
506 mg/kg SKF-82958 remained nonsignificantly greater than fentanyl-treated controls for the  
507 duration of phase III and were no longer significantly depressed relative to saline-treated controls  
508 after 30 min ( $t = 50$ ).

509         The results of experiments with the serotonin receptor agonists 8-OH-DPAT and DOI,  
510 which lacked significant effects on basal respiration, are shown in Figures 6C and 6D,  
511 respectively. There was a main effect of treatment  $\times$  time on mean MVb values across treatment  
512 groups in the analysis of fentanyl + 8-OH-DPAT results [ $F(48, 448) = 3.742$ ;  $p < 0.0001$ ].  
513 Following administration of the higher dose (0.3 mg/kg) of 8-OH-DPAT, fentanyl-induced MVb  
514 depression was significantly worsened by 20 min ( $t = 40$ ), and MVb values remained lower  
515 (nonsignificantly) than fentanyl-treated controls for an additional 20 min ( $t = 60$ ). Conversely,  
516 the results presented in Figure 6D show that administration of DOI to fentanyl-depressed  
517 subjects slightly elevated MVb values between 5- and 25-min post-administration ( $t = 25 - 45$ ).  
518 Two-way ANOVA demonstrated a main effect of treatment [ $F(48, 448) = 4.133$ ;  $p < 0.0001$ ] on  
519 MVb. Subsequent post-hoc comparisons indicated that treatment with 3.0 mg/kg DOI increased

520 MVb sufficiently to achieve complete reversal at 15 min post-administration (t = 35) that  
 521 subsided by the next 5-min bin.



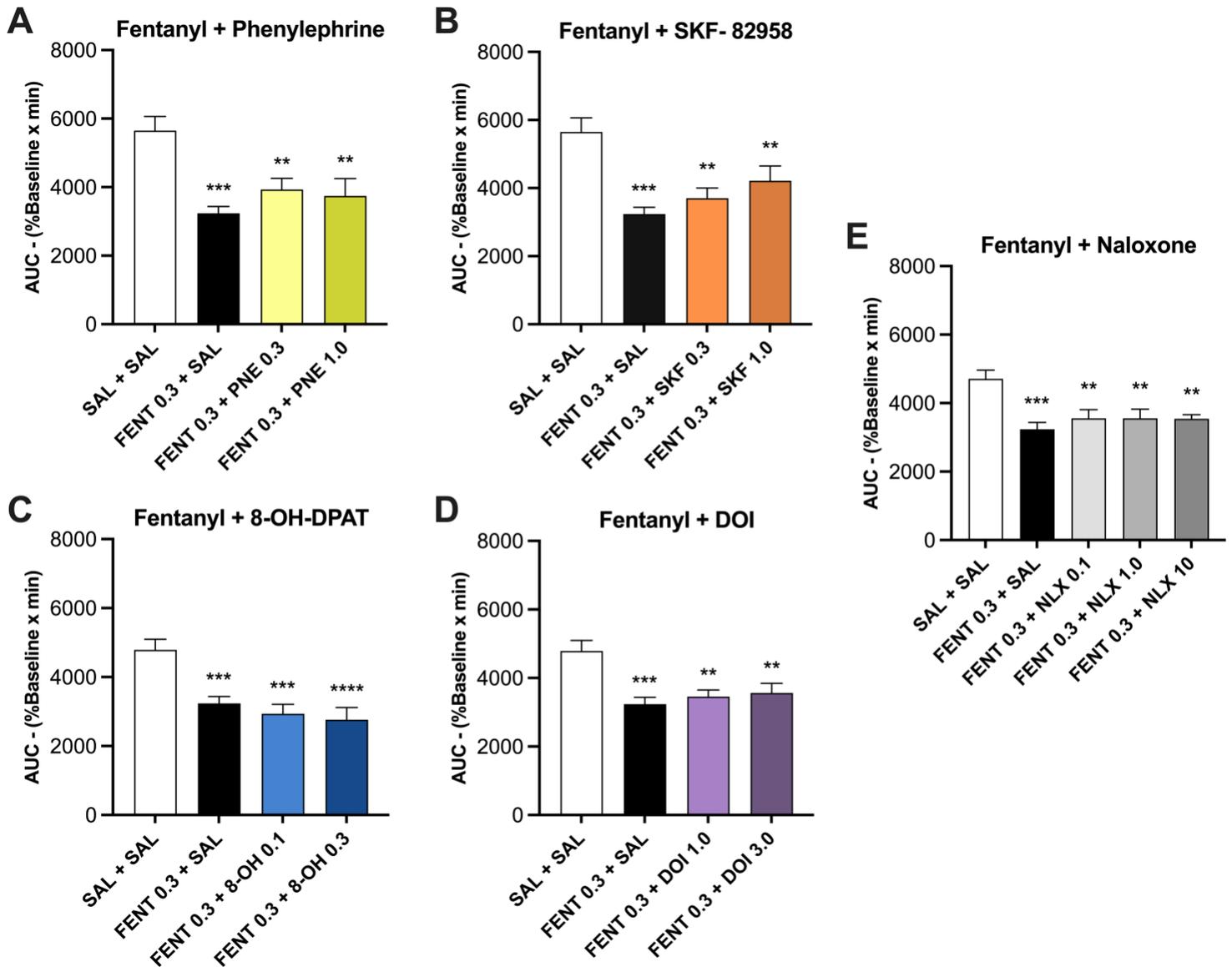
522 **Figure 6: Effects of selected monoamine agonists on fentanyl-depressed respiration.** Dose-  
 523 and time-effects of A) phenylephrine (PNE), B) SKF-82958 (SKF), C) 8-OH-DPAT (8-OH), and  
 524 D) DOI (DOI) on minute volume (MVb) depressed by pretreatment with 0.3 mg/kg fentanyl  
 525 (FENT). Abscissa labels: S = saline injection, F = fentanyl injection, PNE = phenylephrine  
 526 injection, SKF = SKF-82958 injection, 8-OH = 8-OH-DPAT injection, D = DOI injection. N = 8  
 527 per group. Additional \* symbols above or below specific points indicate a significant difference

528 at that time point between individual treatment groups and FENT 0.3 + SAL controls of  $p \leq 0.05$   
529 according to Holm-Šídák post-hoc comparisons. No significant differences were detected at  
530 baseline across experimental conditions when raw MVb values were compared via one-way  
531 ANOVA for groups in panel A) [ $F(3, 28) = 0.8281$ ;  $p = 0.4896$ ], panel B) [ $F(3, 28) = 0.6345$ ;  $p =$   
532  $0.599$ ], panel C) [ $F(3, 28) = 1.116$ ;  $p = 0.3592$ ], or panel D) [ $F(3, 28) = 0.6462$ ;  $p = 0.5919$ ]. All  
533 other details are the same as in Figure 1.

#### 534 *3.4. Summary analysis of monoamine agonist effects on fentanyl-depressed respiration*

535 The results of the post-hoc area under the curve (AUC) analysis of normalized MVb x  
536 Time during phase III for agonist experiments under fentanyl-depressed conditions are shown in  
537 Figure 7. Analysis of AUC data for fentanyl and phenylephrine treatment groups via one-way  
538 ANOVA demonstrated a main effect of treatment [ $F(3, 28) = 7.637$ ;  $p = 0.0007$ ] on AUC, and  
539 subsequent post-hoc comparisons confirmed that pretreatment with fentanyl (0.3 mg/kg)  
540 decreased AUC significantly ( $p = 0.0003$ ) relative to saline-treated controls. Administration of  
541 phenylephrine (0.3, 1.0 mg/kg) nonsignificantly increased AUC relative to fentanyl-treated  
542 controls, and all treatment groups who received fentanyl had significantly lower AUCs than  
543 saline controls (Figure 7A). Similarly, administration of SKF-82958 (0.3, 1.0 mg/kg) to fentanyl-  
544 pretreated subjects significantly affected AUC [ $F(3, 28) = 8.933$ ;  $p = 0.0003$ ] over the course of  
545 phase III (Figure 7B). Treatment with SKF-82958 dose-dependently increased AUC in fentanyl-  
546 pretreated subjects, but post-hoc comparisons indicated that the increases in AUC conferred by  
547 SKF-82958 were not significant relative to fentanyl-treated controls. Figure 7C shows the effects  
548 of 8-OH-DPAT (0.1, 0.3, mg/kg) on AUC in fentanyl-depressed subjects. 8-OH-DPAT  
549 significantly affected AUC with a main effect of treatment [ $F(3, 28) = 10.29$ ;  $p < 0.0001$ ],  
550 characterized by dose-dependent reductions in AUC during phase III. However, post-hoc

551 comparisons indicated that 8-OH-DPAT-mediated decreases in AUC were nonsignificant  
552 relative to fentanyl-treated controls. The effects of treatment with DOI on AUC are shown in  
553 Figure 7D. Analysis of AUC data from groups that received DOI (1.0 and 3.0 mg/kg) following  
554 fentanyl pretreatment demonstrated a significant main effect of treatment [ $F(3, 28) = 8.933$ ;  $p =$   
555  $0.0006$ ], characterized by small, dose-dependent increases in AUC. However, as with previous  
556 agonist treatments, neither dose of DOI significantly increased AUC relative to fentanyl-treated  
557 controls, and all fentanyl-pretreated groups had significantly diminished AUCs than saline-  
558 treated controls regardless of DOI condition. Finally, Figure 7E shows AUCs for naloxone  
559 reversal that were generated from a secondary analysis of data collected in previously published  
560 experiments on the reversal of fentanyl-induced respiratory depression (Elder et al., 2023a) to  
561 provide an active and commonly used comparator treatment. Analysis of data from treatment  
562 groups that received fentanyl (0.3 mg/kg) prior to naloxone (0.1, 1.0, 10 mg/kg) at the start of  
563 phase III showed a significant main effect of treatment on AUC [ $F(4, 35) = 6.309$ ;  $p = 0.0006$ ]  
564 after naloxone administration. However, post-hoc comparisons demonstrate that despite  
565 achieving rapid and complete reversal of fentanyl-induced depression, naloxone did not  
566 significantly increase MVb AUC over 60 minutes relative to fentanyl-treated controls.



567 **Figure 7: Area Under the Curve summary analysis of the effect on fentanyl-depressed**  
 568 **minute volume during phase III by treatment.** Dose-effects of A) phenylephrine (PNE), C)  
 569 SKF-82958 (SKF), D) 8-OH-DPAT (8-OH), E) DOI (DOI), and E) naloxone (NLX) on area  
 570 under the curve (AUC) of normalized minute volume x time during phase III (60 min) in subjects  
 571 pretreated with 0.3 mg/kg fentanyl (FENT). Abscissa labels correspond to injections given at  
 572 time t = 0 + time t = 20, with saline identified as SAL, fentanyl as FENT, and numbers  
 573 corresponding to the dose administered in mg/kg. AUC is given on the ordinate as the product of

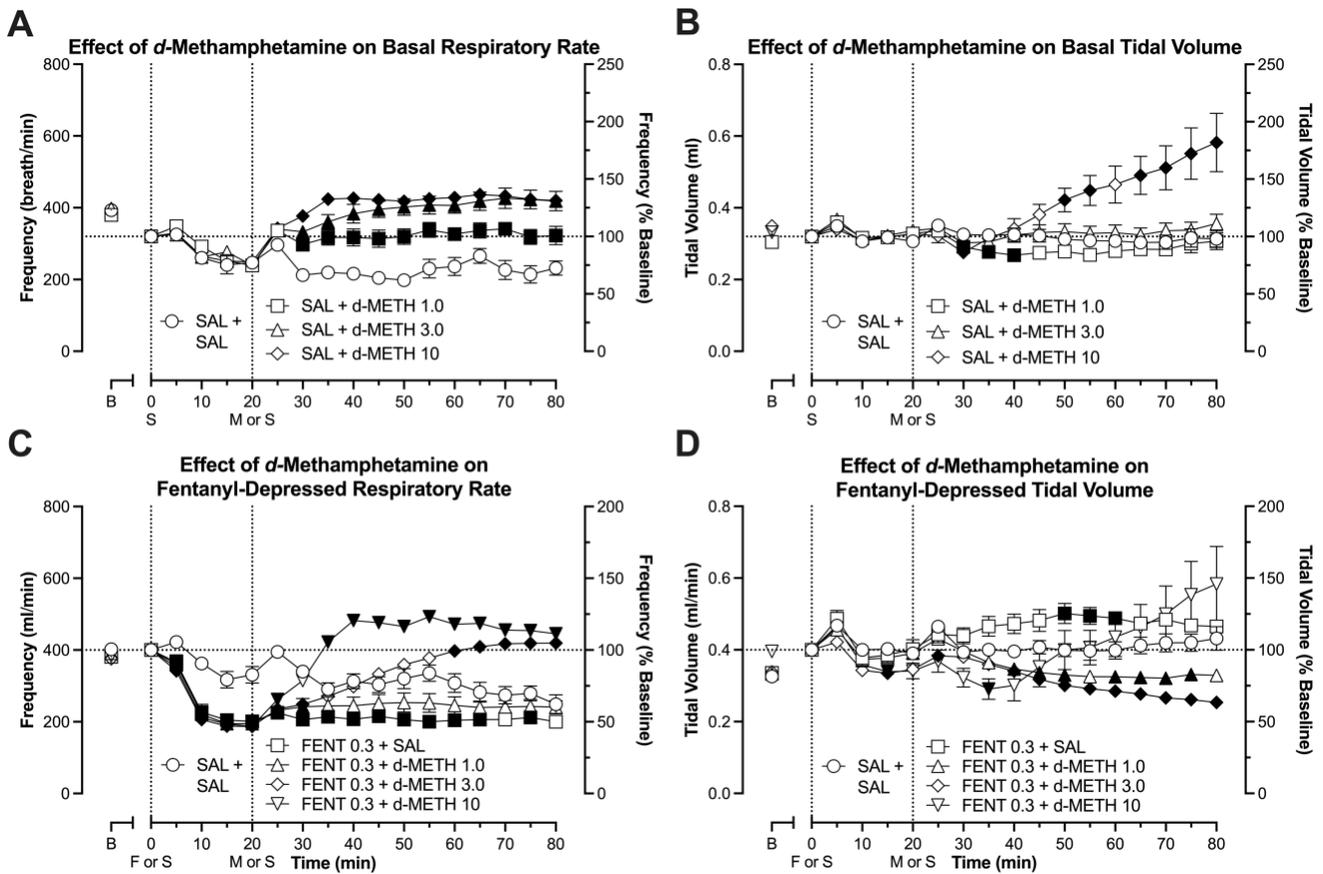
574 % baseline x minutes (min). \*\*; \*\*\*; \*\*\*\* above bars indicate a significant difference between  
575 individual treatment groups and SAL + SAL controls of  $p \leq 0.01$ ; 0.001; 0.0001, respectively,  
576 when analyzed via a one-way ANOVA. Data for AUC analysis of FENT 0.3 + NLX (0.1, 1.0,  
577 10 mg/kg) was obtained from experiments reported in Elder et al., 2023a.

### 578 3.5. Treatment Effects on Frequency and Tidal Volume

579 The dose-dependent effects of *d*-METH (1.0, 3.0, 10 mg/kg) on basal Freq in subjects  
580 who received saline prior to Phase II are shown in Figure 8A. Administration of *d*-METH  
581 significantly affected Freq [ $F(48, 444) = 8.664$ ;  $p < 0.0001$ ], producing dose-dependent  
582 elevations of Freq that were significantly ( $p < 0.05$ ) greater than saline controls for all doses  
583 tested. All doses of *d*-METH significantly increased Freq compared to saline controls within 10  
584 min of administration, quickly reaching peak values by 10 – 15 min post-administration, which  
585 were maintained throughout the recording period. The dose-related effects of *d*-METH on basal  
586 TVb are shown in Figure 8B. Administration of *d*-METH significantly affected TVb [ $F(48, 444)$   
587  $= 8.704$ ;  $p < 0.0001$ ], characterized by transient depression at low doses (1.0 mg/kg) and gradual  
588 yet robust increases at the highest dose (10 mg/kg).

589 The effects of *d*-METH on Freq which were depressed by the administration of fentanyl  
590 (0.3 mg/kg) are shown in Figure 8C. In fentanyl-pretreated mice, there was a significant effect of  
591 *d*-METH over time [ $F(64, 560) = 25.52$ ;  $p < 0.0001$ ] on Freq. In contrast to the dose-dependent  
592 stimulation of Freq observed with *d*-METH under basal conditions, administration of *d*-METH  
593 to subjects that were pretreated with fentanyl (0.3 mg/kg) produced bidirectional, dose-  
594 dependent effects following a similar pattern as was observed with racemic METH and nearly  
595 identical to those observed on MVb with *d*-METH, demonstrating that METH and its  
596 enantiomers primarily modulate MVb via alterations in Freq. The effects of *d*-METH on TVb

597 that was depressed by the administration of fentanyl (0.3 mg/kg) are shown in Figure 8D. In  
598 fentanyl-pretreated mice, there was a significant effect of *d*-METH over time [F(64, 560) =  
599 7.637;  $p < 0.0001$ ] on TVb, albeit to a lesser degree than Freq. *d*-METH's effects on fentanyl-  
600 depressed TVb closely mirrored its effects on depressed Freq, albeit with a greater magnitude of  
601 depressant effects and milder stimulant effects at lower and higher doses, respectively. While the  
602 relationship between TVb and Freq was similar to what was observed under basal conditions, the  
603 balance of their contributions shifted toward TVb-driven depression. At low (1.0 mg/kg) and  
604 moderate (3.0 mg/kg) doses, *d*-METH significantly depressed TVb, in contrast to the significant  
605 compensatory elevation seen in fentanyl-treated controls who received saline at  $t = 20$ . The  
606 highest dose of 10 mg/kg *d*-METH had complex effects on TVb, inducing transient significant  
607 depression at  $t = 35$  followed by a gradual, nonsignificant increase compared to controls.



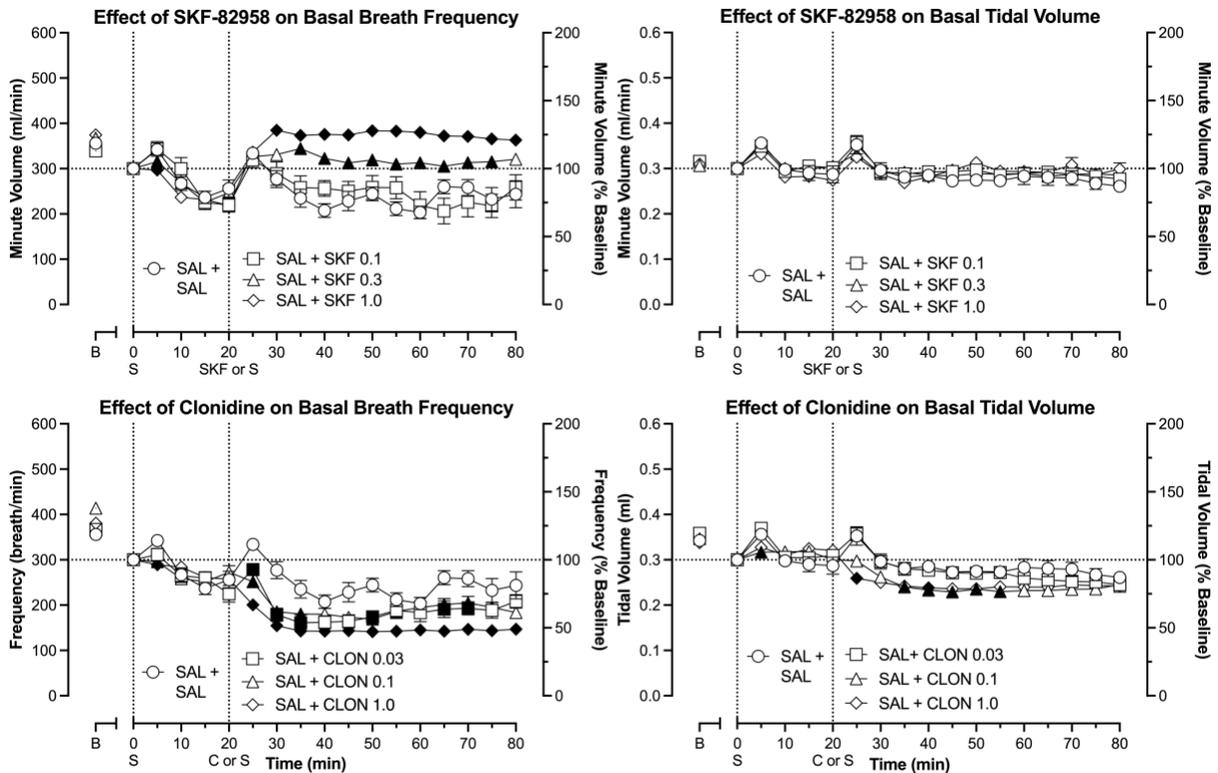
609 **Figure 8: Effects of *d*-METH on basal and fentanyl-depressed breath frequency and**  
 610 **volume.** Panel A) Dose- and time-effects of *d*-methamphetamine (*d*-METH) on breath frequency  
 611 (Freq); Panel B) dose- and time-effects of *d*-METH on tidal volume (TVb); C) dose- and time-  
 612 effects of *d*-METH on fentanyl- (FENT) depressed breath frequency (Freq); D) dose- and time-  
 613 effects of *d*-METH on FENT-depressed TVb. Left ordinate: mean raw Freq (breaths/min) or  
 614 TVb indexing values of symbols only during baseline (B) of Phase I. Right ordinate: normalized  
 615 (percent baseline) Freq or TVb indexing values of symbols during the 80-min test session  
 616 following Phase I baseline. Symbols indicate mean Freq or TVb expressed as a percentage of  
 617 baseline Freq or TVb for treatment groups consisting of 8 mice. Filled symbols indicate  
 618 significant differences compared to the respective Freq or TVb of saline-treated controls at

619 individual time points ( $p \leq 0.05$ ). Abscissa labels: B = mean baseline Freq or TVb, F, M or S =  
620 fentanyl, *d*-METH or saline (SAL) injection, respectively. Legend labels correspond to the dose  
621 in mg/kg. All other details are the same as in Figure 1.

622         The dose-dependent effects of SKF-82958 (1.0, 3.0, 10 mg/kg) on basal Freq in subjects  
623 who received saline prior to Phase II are shown in Figure 9A. Administration of SKF-82958  
624 significantly affected Freq over time [ $F(48, 444) = 7.861$ ;  $p < 0.0001$ ], producing dose-dependent  
625 elevations of Freq that were significantly greater than saline controls at 0.3 and 1.0 mg/kg. The  
626 two highest doses of SKF-82958 significantly increased Freq compared to saline controls within  
627 10 and 15 min of administration, respectively,, representing peak effects maintained throughout  
628 the recording period. The dose-related effects of SKF-82958 on basal TVb are shown in Figure  
629 9B. SKF-82958 had a significant effect on TVb over time [ $F(48, 444) = 1.999$ ;  $p = 0.0002$ ] but  
630 did not significantly alter TVb at any timepoint relative to saline-treated controls when data were  
631 analyzed via post-hoc comparisons.

632         The dose-dependent effects of clonidine (0.03, 0.1, 1.0 mg/kg) on basal Freq in subjects  
633 who received saline prior to Phase II are shown in Figure 9C. Administration of clonidine  
634 significantly affected Freq [ $F(48, 448) = 4.194$ ;  $p < 0.0001$ ], producing significant dose-  
635 dependent depression of Freq at all doses when compared with saline-treated controls. All doses  
636 of clonidine significantly decreased Freq from 5 – 15 min post-administration ( $t = 25 - 35$ ),  
637 which represented peak depressant effects that slowly dissipated for 0.03 and 0.1 mg/kg  
638 treatments, while the maximal depressant effects of 1.0 mg/kg were maintained throughout the  
639 recording period. The dose-related effects of clonidine on basal TVb are shown in Figure 9D.  
640 Clonidine had a significant effect on TVb over time [ $F(48, 448) = 4.173$ ;  $p < 0.0001$ ], with post-  
641 hoc comparisons showing significant depression by the highest dose (1.0 mg/kg) at two time

642 points ( $t = 25$  and  $40$ ) and significant depression by the intermediate dose ( $0.1$  mg/kg) from  $15 -$   
 643  $35$  min ( $t = 35 - 55$ ) compared with saline-treated controls.



644

645 **Figure 9: Effects of representative stimulant and depressant agonists on basal and**  
 646 **depressed breath frequency and tidal volume.** Panel A) Dose- and time-effects of SKF-82958  
 647 (SKF) on breath frequency (Freq); Panel B) dose- and time-effects of SKF on tidal volume  
 648 (TVb); Panel C) dose- and time-effects of clonidine (CLON) on breath frequency (Freq); Panel  
 649 D) dose- and time-effects of CLON on tidal volume (TVb). Left ordinate: mean raw Freq  
 650 (breaths/min) or TVb indexing values of symbols only during baseline (B) of Phase I. Right  
 651 ordinate: normalized (percent baseline) Freq or TVb indexing values of symbols during the 80-  
 652 min test session following Phase I baseline. Symbols indicate mean Freq or TVb expressed as a  
 653 percentage of baseline Freq or TVb for treatment groups consisting of 8 mice. Filled symbols

654 indicate significant differences compared to the respective Freq or TVb of saline-treated controls  
655 at individual time points ( $p \leq 0.05$ ). Abscissa labels: B = mean baseline, S, SKF, or C = saline  
656 (SAL), SKF-82958, or clonidine injection, respectively. Legend labels correspond to the dose in  
657 mg/kg. All other details are the same as in Figure 1.

#### 658 **4. Discussion and Summary**

659 Overall, previously published reports by other laboratories and the results of the present  
660 study show that monoamine receptors are: 1) present in brainstem regions relevant to respiration;  
661 2) involved in modulating the activity of respiratory networks; 3) able to be manipulated  
662 pharmacologically to alter respiration; and 4) capable of altering OIRD in laboratory animals  
663 (Ciarka et al., 2007; Imam et al., 2020; Lalley, 2008; Ramirez et al., 2012; van der Schier et al.,  
664 2014). In the first stage of the present study the effects of the two METH enantiomers, *d*- and *l*-  
665 METH, on basal and fentanyl-depressed respiration were evaluated to determine their individual  
666 contributions to the bidirectional respiratory modulation observed previously with racemic  
667 METH. There were two main findings from these experiments. First, *d*- and *l*-METH were  
668 shown to have opposing effects on basal respiration, as evidenced by the complete separation of  
669 respiratory stimulant and respiratory depressant effects between nominally equal doses of *d*-  
670 METH and *l*-METH, respectively (Figures 1A and 1B). Second, experiments that evaluated the  
671 two enantiomers under fentanyl-depressed conditions showed a recapitulation of the racemate's  
672 bidirectional respiratory effects with *d*-METH, while *l*-METH was shown to significantly  
673 exacerbate fentanyl-induced respiratory depression at all doses tested (Figures 1C and 1D).

674 While the enantiomers tended to modulate fentanyl-depressed respiration in the manner  
675 that was hypothesized, the unexpected recapitulation of bidirectional effects in fentanyl-  
676 depressed experiments with *d*-METH provides insight into the pharmacological determinants of

677 METH's respiratory activity and how it may be altered in the presence of fentanyl. While these  
678 results support the hypothesis that efficacy for releasing monoamines, more specifically relative  
679 efficacy for DA/5HT release, is correlated with respiratory stimulation, it remains to be  
680 determined whether such stimulation is the result of a direct effect of METH on monoamine  
681 transmission in brainstem respiratory networks or relies on monoamine-induced increases in  
682 downstream glutamate transmission to these areas (Fischer et al., 2021). Since amphetamines  
683 exert their effects via multiple mechanisms that include TAAR1 activation, their ability to  
684 influence synaptic monoamine levels may vary substantially based on the neurophysiology of the  
685 CNS regions in question (Abekawa et al., 1994; Stephans and Yamamoto, 1995; Underhill et al.,  
686 2019, 2014). Furthermore, since glutamate is known to play a primary role in controlling  
687 respiratory network activity, both intrinsically and extrinsically, to carry signals from distal  
688 inputs like chemosensors, it is possible that METH modulates glutamatergic inputs to respiratory  
689 networks through its effects on monoamines in regions that interact with those projections (Ang  
690 et al., 1992; Martelli et al., 2013; Pilowsky et al., 2009). Regardless of how monoamine release  
691 by METH specifically leads to increased respiratory output, these results provide compelling  
692 evidence that potency and selectivity for monoamine release is a key determinant of the nature of  
693 respiratory stimulation, as evidenced by *l*-METH's opposing effects on MVb.

694         In the second stage of this study, six selective monoamine receptor agonists  
695 (phenylephrine, clonidine, SKF-82958, quinpirole, 8-OH-DPAT, and DOI) were initially  
696 characterized for their effects on basal respiration to identify receptor-agonist pairings with  
697 respiratory stimulant effects. Subsequent experiments evaluated the ability of agonists that did  
698 not depress basal respiration to reverse respiratory depression induced by an ED<sub>50</sub> dose of  
699 fentanyl. The results obtained from these experiments provided two primary findings. First,

700 experimental results showed that agonists of excitatory catecholamine receptors, phenylephrine  
701 ( $\alpha_1$ ) and SKF-82958 ( $D_1$ ), stimulated MVb in a dose-dependent manner under both basal and  
702 fentanyl-depressed conditions. Second, agonists of inhibitory catecholamine receptors often  
703 associated with presynaptic neurons, clonidine ( $\alpha_2$ ) and quinpirole ( $D_2$ ), were shown to depress  
704 basal respiration following administration dose-dependently. These results are in line with  
705 previously published research demonstrating respiratory stimulant effects of  $D_1$  receptor agonists  
706 and the rhythm-enhancing effects of NE inputs to brainstem nuclei (Errchidi et al., 1991, 1990;  
707 Lalley, 2008, 2005, 2004). Additionally, these results show a dichotomy between respiratory  
708 stimulant effects of post-synaptic receptors and depressant effects of pre-synaptic autoreceptors,  
709 further supporting the hypothesis that mild increases in synaptic monoamines induced by low  
710 doses of amphetamines may preferentially activate autoreceptors to induce respiratory  
711 depression.

712         Although neither of the two serotonin receptor agonists strongly modulated basal or  
713 depressed MVb, a similar pattern emerged whereby 8-OH-DPAT ( $5HT_{1a}$ ), an agonist of an  
714 inhibitory presynaptic receptor subtype, tended to be pro-depressant, while DOI ( $5HT_{2a}$ ), an  
715 agonist of excitatory post-synaptic receptor subtypes, tended to be mildly stimulating. Although  
716 ample evidence in the literature supports the respiratory activity of 8-OH-DPAT, many of the  
717 studies published on the respiratory effects of 5HT agonists were conducted in neonates and in  
718 the context of various existing respiratory pathologies (Bodineau et al., 2004; Guenther et al.,  
719 2009; Günther et al., 2006; Mathew, 2011; Stettner et al., 2008; Veasey, 2003). Our results  
720 contradict some earlier findings from experiments with morphine, fentanyl, and remifentanyl-  
721 treated animals that showed  $5HT_{1a}$ -mediated selective reversal of respiratory depression, but are  
722 in line with negative results that have been reported from other preclinical and clinical trials of

723 5HT<sub>1a</sub> agonists, including buspirone, for the treatment of central apneas (Guenther et al., 2012,  
724 2010; Oertel et al., 2007; Ren et al., 2015). Although *in vivo* models have demonstrated  
725 respiratory stimulant effects of DOI previously in different contexts (Andrzejewski et al., 2017;  
726 Budzinska, 2009), these data represent the first time the stimulant effects of DOI on OIRD have  
727 been reported. The limited efficacy of DOI under fentanyl-depressed conditions may be a  
728 product of its psychedelic pharmacology, which is known to induce broad enhancements in  
729 neuronal network activity and glutamate transmission within the CNS (Inserra et al., 2021;  
730 Mason et al., 2020; Nichols, 2016, 2004; Vollenweider and Komater, 2010). Taken together,  
731 these findings indicate that monoaminergic inputs influence respiration, which can be  
732 manipulated in either direction using agonists selective for excitatory post-synaptic receptors or  
733 inhibitory presynaptic receptors.

734         The primary findings from experiments conducted in stages one and two of this study are  
735 complementary and provide insight into the monoaminergic mechanisms that mediate METH's  
736 bidirectional effects on respiration. Findings in stage one demonstrated that the enantiomer with  
737 the greatest absolute and relative potency for releasing DA, *d*-METH, acted almost entirely as a  
738 respiratory stimulant under basal and depressed conditions at the doses tested. Conversely, *l*-  
739 METH, a substantially less potent monoamine releaser biased toward NE release, primarily acted  
740 as a depressant when tested at the same doses. However, when assayed at a dose beyond those  
741 used in experiments with other enantiomeric compositions, 30 mg/kg *l*-METH displayed  
742 moderate yet significant stimulant effects similar to those of 3.0 mg/kg *d*-METH or 10 mg/kg  
743 racemic METH, thus confirming that *l*-METH also displays dose-related bidirectionality rather  
744 than solely dose-dependent depression. These findings demonstrate that the two enantiomers  
745 have differential potency as respiratory stimulants, as opposed to exerting opposing influences

746 on respiration as was originally hypothesized. The respiratory stimulant activity of high-dose *l*-  
747 METH may be explained as a consequence of the administration of an adequate dose to produce  
748 sufficient release of NE, and possibly DA, to cause a shift in the balance of indirect agonism  
749 toward an overall excitatory effect on neurotransmission and respiratory output. This hypothesis  
750 could also be extended in the opposite direction for *d*-METH, whereby the administration of  
751 lower doses ( $\leq 0.3$  mg/kg) may engender depressant effects as a function of lesser NE release  
752 leading to a shift in the balance toward overall inhibition. When considered alongside the results  
753 obtained from experiments of catecholamine receptor agonists, the evidence suggests that  
754 increased signaling at excitatory post-synaptic catecholamine receptors may underlie the  
755 respiratory stimulant properties of METH and represents a mechanism that could potentially be  
756 exploited in the future development of respiratory stimulant therapeutics. Similarly, inhibitory  
757 presynaptic catecholamine receptors may be responsible for the respiratory depressant effects of  
758 METH and represent a target for medications development efforts to rescue METH-  
759 compromised respiration.

760           Interestingly, enhancement of glutamate may be a shared mechanism among the agonists  
761 that produced elevations of fentanyl-depressed respiration as well as the amphetamines (Chen et  
762 al., 2006; Kalivas and Duffy, 1995; Kanbayashi et al., 2000; Nishino et al., 1998). A growing  
763 body of recent evidence points to the involvement of AMPA receptor activation in the generation  
764 and stimulation of respiratory network activity, which could potentially be the downstream  
765 effector mediating the effects of excitatory monoamine receptors (Dahan et al., 2018; Imam et  
766 al., 2020; Oertel et al., 2010; Ren et al., 2009; van der Schier et al., 2014). In fact, preliminary  
767 data collected in early experiments with the  $\alpha_1$  antagonist prazosin, which is thought to decrease  
768 glutamatergic transmission from regions such as the hypothalamus, spinal cord and brainstem,

769 showed dose-dependent depression of MVb following administration (data not shown) (Chen et  
770 al., 2006).

771 Overall, these data demonstrate that activation of monoaminergic receptors can  
772 differentially modulate respiration based on the receptor subtype's effect on cellular and network  
773 activity. Furthermore, while data from these experiments cannot confirm whether the respiratory  
774 effects of METH are mediated by monoamine receptors in the brainstem, similarities between  
775 the effects of METH and catecholamine receptor agonists on respiratory frequency suggest they  
776 are likely involved. To this point, the findings reported here have identified potential targets for  
777 future analeptic development ( $D_1$  &  $\alpha_1$ ) based on their ability to mitigate OIRD from fentanyl, as  
778 well as the receptors that are likely mediators of enhanced OIRD toxicity ( $D_2$  &  $\alpha_2$ ). . Future  
779 studies should be conducted specifically in human subjects to test the cross-species consistency  
780 of the bi-directional effects of racemic methamphetamine, and to evaluate the use of  
781 monoaminergic analeptics by themselves and in combination with opioid antagonists such as  
782 naloxone for their ability to provide rapid and sustained reversal of OIRD, especially in the  
783 context of fentanyl.

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790 **REFERENCES**

- 791 Abekawa, T., Ohmori, T., Koyama, T., 1994. Effects of repeated administration of a high dose of  
 792 methamphetamine on dopamine and glutamate release in rat striatum and nucleus  
 793 accumbens. *Brain Res* 643, 276–281. [https://doi.org/10.1016/0006-8993\(94\)90033-7](https://doi.org/10.1016/0006-8993(94)90033-7)  
 794 Andersen, P.H., Grønvald, F.C., Jansen, J.A., 1985. A comparison between dopamine-stimulated  
 795 adenylyl cyclase and 3H-SCH 23390 binding in rat striatum. *Life Sciences* 37, 1971–  
 796 1983. [https://doi.org/10.1016/0024-3205\(85\)90028-1](https://doi.org/10.1016/0024-3205(85)90028-1)  
 797 Andrzejewski, K., Kaczyńska, K., Zaremba, M., 2017. Serotonergic system in hypoxic  
 798 ventilatory response in unilateral rat model of Parkinson’s disease. *J Biomed Sci* 24.  
 799 <https://doi.org/10.1186/s12929-017-0331-2>  
 800 Ang, R.C., Hoop, B., Kazemi, H., 1992. Role of glutamate as the central neurotransmitter in the  
 801 hypoxic ventilatory response. *Journal of Applied Physiology* 72, 1480–1487.  
 802 <https://doi.org/10.1152/jappl.1992.72.4.1480>  
 803 Ashok, A.H., Mizuno, Y., Volkow, N.D., Howes, O.D., 2017. Association of Stimulants With  
 804 Dopaminergic Alterations in Users of Cocaine, Amphetamine, and Methamphetamine: A  
 805 Systematic Review and Meta-analysis. *JAMA Psychiatry* 74, 511–519.  
 806 <https://doi.org/10.1001/jamapsychiatry.2017.0135>  
 807 Bodineau, L., Cayetanot, F., Marlot, D., Collin, T., Gros, F., Frugière, A., 2004. Endogenous 5-  
 808 HT1/2 systems and the newborn rat respiratory control. *Respiratory Physiology &*  
 809 *Neurobiology* 141, 47–57. <https://doi.org/10.1016/j.resp.2004.03.007>  
 810 Boess, F.G., Martin, I.L., 1994. Molecular biology of 5-HT receptors. *Neuropharmacology* 33,  
 811 275–317. [https://doi.org/10.1016/0028-3908\(94\)90059-0](https://doi.org/10.1016/0028-3908(94)90059-0)  
 812 Bolme, P., Corrodi, H., Fuxe, K., Hökfelt, T., Lidbrink, P., Goldstein, M., 1974. Possible  
 813 involvement of central adrenaline neurons in vasomotor and respiratory control. *Studies*  
 814 *with clonidine and its interactions with piperoxane and yohimbine. European Journal of*  
 815 *Pharmacology* 28, 89–94. [https://doi.org/10.1016/0014-2999\(74\)90116-2](https://doi.org/10.1016/0014-2999(74)90116-2)  
 816 Borsini, F., Giraldo, E., Monferini, E., Antonini, G., Parenti, M., Bietti, G., Donetti, A., 1995.  
 817 BIMT 17, a 5-HT<sub>2A</sub> receptor antagonist and 5-HT<sub>1A</sub> receptor full agonist in rat cerebral  
 818 cortex. *Naunyn Schmiedebergs Arch Pharmacol* 352, 276–282.  
 819 <https://doi.org/10.1007/BF00168557>  
 820 Boundy, V.A., Luedtke, R.R., Gallitano, A.L., Smith, J.E., Filtz, T.M., Kallen, R.G., Molinoff,  
 821 P.B., 1993. Expression and characterization of the rat D<sub>3</sub> dopamine receptor:  
 822 pharmacologic properties and development of antibodies. *J Pharmacol Exp Ther* 264,  
 823 1002–1011.  
 824 Boyajian, C.L., Leslie, F.M., 1987. Pharmacological evidence for alpha-2 adrenoceptor  
 825 heterogeneity: differential binding properties of [3H]rauwolscine and [3H]idazoxan in rat  
 826 brain. *J Pharmacol Exp Ther* 241, 1092–1098.  
 827 Budzinska, K., 2009. Serotonergic modulation of cortical and respiratory responses to episodic  
 828 hypoxia. *Eur J Med Res* 14 Suppl 4, 32–37. <https://doi.org/10.1186/2047-783x-14-s4-32>  
 829 Campiani, G., Nacci, V., Bechelli, S., Ciani, S.M., Garofalo, A., Fiorini, I., Wikström, H., de  
 830 Boer, P., Liao, Y., Tepper, P.G., Cagnotto, A., Mennini, T., 1998. New antipsychotic  
 831 agents with serotonin and dopamine antagonist properties based on a pyrrolo[2,1-  
 832 b][1,3]benzothiazepine structure. *J Med Chem* 41, 3763–3772.  
 833 <https://doi.org/10.1021/jm9706832>

834 Cano, M., Huang, Y., 2021. Overdose deaths involving psychostimulants with abuse potential,  
835 excluding cocaine: State-level differences and the role of opioids. *Drug and Alcohol*  
836 *Dependence* 218, 108384. <https://doi.org/10.1016/j.drugalcdep.2020.108384>

837 Chen, Q., Li, D.-P., Pan, H.-L., 2006. Presynaptic  $\alpha 1$  Adrenergic Receptors Differentially  
838 Regulate Synaptic Glutamate and GABA Release to Hypothalamic Presympathetic  
839 Neurons. *J Pharmacol Exp Ther* 316, 733–742. <https://doi.org/10.1124/jpet.105.094797>

840 Ciarka, A., Vincent, J.-L., van de Borne, P., 2007. The effects of dopamine on the respiratory  
841 system: Friend or foe? *Pulmonary Pharmacology & Therapeutics* 20, 607–615.  
842 <https://doi.org/10.1016/j.pupt.2006.10.011>

843 Corcoran, A.E., Commons, K.G., Wu, Y., Smith, J.C., Harris, M.B., Richerson, G.B., 2014. Dual  
844 Effects of 5-HT<sub>1a</sub> Receptor Activation on Breathing in Neonatal Mice. *J Neurosci* 34,  
845 51–59. <https://doi.org/10.1523/JNEUROSCI.0864-13.2014>

846 Cruickshank, C.C., Dyer, K.R., 2009. A review of the clinical pharmacology of  
847 methamphetamine. *Addiction* 104, 1085–1099. [https://doi.org/10.1111/j.1360-](https://doi.org/10.1111/j.1360-0443.2009.02564.x)  
848 [0443.2009.02564.x](https://doi.org/10.1111/j.1360-0443.2009.02564.x)

849 Cunningham, J.K., Maxwell, J.C., Campollo, O., Liu, L.-M., Lattyak, W.J., Callaghan, R.C.,  
850 2013. Mexico's precursor chemical controls: Emergence of less potent types of  
851 methamphetamine in the United States. *Drug and Alcohol Dependence* 129, 125–136.  
852 <https://doi.org/10.1016/j.drugalcdep.2012.10.001>

853 Dahan, A., van der Schrier, R., Smith, T., Aarts, L., van Velzen, M., Niesters, M., 2018.  
854 Averting Opioid-induced Respiratory Depression without Affecting Analgesia.  
855 *Anesthesiology* 128, 1027–1037. <https://doi.org/10.1097/ALN.0000000000002184>

856 Desai, R.I., Terry, P., Katz, J.L., 2005. A comparison of the locomotor stimulant effects of D1-  
857 like receptor agonists in mice. *Pharmacology Biochemistry and Behavior* 81, 843–848.  
858 <https://doi.org/10.1016/j.pbb.2005.06.006>

859 Egan, C.T., Herrick-Davis, K., Miller, K., Glennon, R.A., Teitler, M., 1998. Agonist activity of  
860 LSD and lisuride at cloned 5HT<sub>2A</sub> and 5HT<sub>2C</sub> receptors. *Psychopharmacology* 136,  
861 409–414. <https://doi.org/10.1007/s002130050585>

862 Eilam, D., Szechtman, H., 1989. Biphasic effect of D-2 agonist quinpirole on locomotion and  
863 movements. *Eur J Pharmacol* 161, 151–157. [https://doi.org/10.1016/0014-](https://doi.org/10.1016/0014-2999(89)90837-6)  
864 [2999\(89\)90837-6](https://doi.org/10.1016/0014-2999(89)90837-6)

865 Elder, H.J., Varshneya, N.B., Walentiny, D.M., Beardsley, P.M., 2023a. Amphetamines  
866 modulate fentanyl-depressed respiration in a bidirectional manner. *Drug Alcohol Depend*  
867 243, 109740. <https://doi.org/10.1016/j.drugalcdep.2022.109740>

868 Elder, H.J., Walentiny, D.M., Beardsley, P.M., 2023b. Theophylline reverses oxycodone's but  
869 not fentanyl's respiratory depression in mice while caffeine is ineffective against both  
870 opioids. *Pharmacology Biochemistry and Behavior* 229, 173601.  
871 <https://doi.org/10.1016/j.pbb.2023.173601>

872 Errchidi, S., Hilaire, G., Monteau, R., 1990. Permanent release of noradrenaline modulates  
873 respiratory frequency in the newborn rat: an in vitro study. *J Physiol* 429, 497–510.

874 Errchidi, S., Monteau, R., Hilaire, G., 1991. Noradrenergic modulation of the medullary  
875 respiratory rhythm generator in the newborn rat: an in vitro study. *J Physiol* 443, 477–  
876 498.

877 Fischer, K.D., Knackstedt, L.A., Rosenberg, P.A., 2021. Glutamate homeostasis and dopamine  
878 signaling: Implications for psychostimulant addiction behavior. *Neurochem Int* 144,  
879 104896. <https://doi.org/10.1016/j.neuint.2020.104896>

- 880 Friedman, J., Shover, C.L., 2023. Charting the fourth wave: Geographic, temporal, race/ethnicity  
881 and demographic trends in polysubstance fentanyl overdose deaths in the United States,  
882 2010–2021. *Addiction* n/a. <https://doi.org/10.1111/add.16318>
- 883 Guenther, U., Manzke, T., Wrigge, H., Dutschmann, M., Zinserling, J., Putensen, C., Hoeft, A.,  
884 2009. The Counteraction of Opioid-Induced Ventilatory Depression by the Serotonin 1A-  
885 Agonist 8-OH-DPAT Does Not Antagonize Antinociception in Rats In Situ and In Vivo.  
886 *Anesthesia & Analgesia* 108, 1169–1176. <https://doi.org/10.1213/ane.0b013e318198f828>
- 887 Guenther, U., Theuerkauf, N.U., Huse, D., Boettcher, M.F., Wensing, G., Putensen, C., Hoeft,  
888 A., 2012. Selective 5-HT1A-R-agonist Repinotan Prevents Remifentanil-induced  
889 Ventilatory Depression and Prolongs Antinociception. *Anesthesiology* 116, 56–64.  
890 <https://doi.org/10.1097/ALN.0b013e31823d08fa>
- 891 Guenther, U., Wrigge, H., Theuerkauf, N., Boettcher, M.F., Wensing, G., Zinserling, J.,  
892 Putensen, C., Hoeft, A., 2010. Repinotan, a selective 5-HT1A-R-agonist, antagonizes  
893 morphine-induced ventilatory depression in anesthetized rats. *Anesth Analg* 111, 901–  
894 907. <https://doi.org/10.1213/ANE.0b013e3181eac011>
- 895 Günther, S., Maroteaux, L., Schwarzacher, S.W., 2006. Endogenous 5-HT2B receptor activation  
896 regulates neonatal respiratory activity in vitro. *J Neurobiol* 66, 949–961.  
897 <https://doi.org/10.1002/neu.20253>
- 898 Hassan, S.F., Wearne, T.A., Cornish, J.L., Goodchild, A.K., 2016. Effects of acute and chronic  
899 systemic methamphetamine on respiratory, cardiovascular and metabolic function, and  
900 cardiorespiratory reflexes. *J Physiol* 594, 763–780. <https://doi.org/10.1113/JP271257>
- 901 Imam, M.Z., Kuo, A., Smith, M.T., 2020. Countering opioid-induced respiratory depression by  
902 non-opioids that are respiratory stimulants. *F1000Res* 9, 91.  
903 <https://doi.org/10.12688/f1000research.21738.1>
- 904 Insera, A., Gregorio, D.D., Gobbi, G., 2021. Psychedelics in Psychiatry: Neuroplastic,  
905 Immunomodulatory, and Neurotransmitter Mechanisms. *Pharmacol Rev* 73, 202–277.  
906 <https://doi.org/10.1124/pharmrev.120.000056>
- 907 Jaster, A.M., Elder, H., Marsh, S.A., de la Fuente Revenga, M., Negus, S.S., González-Maeso, J.,  
908 2022. Effects of the 5-HT2A receptor antagonist volinanserin on head-twitch response  
909 and intracranial self-stimulation depression induced by different structural classes of  
910 psychedelics in rodents. *Psychopharmacology*. <https://doi.org/10.1007/s00213-022-06092-x>
- 911
- 912 Kalivas, P.W., Duffy, P., 1995. D1 receptors modulate glutamate transmission in the ventral  
913 tegmental area. *J. Neurosci.* 15, 5379–5388. <https://doi.org/10.1523/JNEUROSCI.15-07-05379.1995>
- 914
- 915 Kanbayashi, T., Honda, K., Kodama, T., Mignot, E., Nishino, S., 2000. Implication of  
916 dopaminergic mechanisms in the wake-promoting effects of amphetamine: a study of d-  
917 and l-derivatives in canine narcolepsy. *Neuroscience* 99, 651–659.  
918 [https://doi.org/10.1016/S0306-4522\(00\)00239-6](https://doi.org/10.1016/S0306-4522(00)00239-6)
- 919 Kuczenski, R., Segal, D., Cho, A., Melega, W., 1995. Hippocampus norepinephrine, caudate  
920 dopamine and serotonin, and behavioral responses to the stereoisomers of amphetamine  
921 and methamphetamine. *J Neurosci* 15, 1308–1317.  
922 <https://doi.org/10.1523/JNEUROSCI.15-02-01308.1995>
- 923 Lalley, P.M., 2008. OPIOIDERGIC AND DOPAMINERGIC MODULATION OF  
924 RESPIRATION. *Respir Physiol Neurobiol* 164, 160–167.  
925 <https://doi.org/10.1016/j.resp.2008.02.004>

- 926 Lalley, P.M., 2005. D1-dopamine receptor agonists prevent and reverse opiate depression of  
927 breathing but not antinociception in the cat. *Am J Physiol Regul Integr Comp Physiol*  
928 289, R45-51. <https://doi.org/10.1152/ajpregu.00868.2004>
- 929 Lalley, P.M., 2004. Dopamine1 receptor agonists reverse opioid respiratory network depression,  
930 increase CO2 reactivity. *Respir Physiol Neurobiol* 139, 247–262.  
931 <https://doi.org/10.1016/j.resp.2003.10.007>
- 932 Lalley, P.M., Bischoff, A.M., Schwarzacher, S.W., Richter, D.W., 1995. 5-HT2 receptor-  
933 controlled modulation of medullary respiratory neurones in the cat. *J Physiol* 487, 653–  
934 661.
- 935 Lawler, C.P., Prioleau, C., Lewis, M.M., Mak, C., Jiang, D., Schetz, J.A., Gonzalez, A.M.,  
936 Sibley, D.R., Mailman, R.B., 1999. Interactions of the Novel Antipsychotic Aripiprazole  
937 (OPC-14597) with Dopamine and Serotonin Receptor Subtypes. *Neuropsychopharmacol*  
938 20, 612–627. [https://doi.org/10.1016/S0893-133X\(98\)00099-2](https://doi.org/10.1016/S0893-133X(98)00099-2)
- 939 Losacker, M., Zörntlein, S., Schwarze, B., Staudt, S., Röhrich, J., Hess, C., 2021. Determination  
940 of the enantiomeric composition of amphetamine, methamphetamine and 3,4-  
941 methylenedioxy-N-methylamphetamine (MDMA) in seized street drug samples from  
942 southern Germany. *Drug Testing and Analysis* n/a. <https://doi.org/10.1002/dta.3118>
- 943 Lovenberg, T.W., Baron, B.M., de Lecea, L., Miller, J.D., Prosser, R.A., Rea, M.A., Foye, P.E.,  
944 Racke, M., Slone, A.L., Siegel, B.W., 1993. A novel adenylyl cyclase-activating  
945 serotonin receptor (5-HT7) implicated in the regulation of mammalian circadian rhythms.  
946 *Neuron* 11, 449–458. [https://doi.org/10.1016/0896-6273\(93\)90149-1](https://doi.org/10.1016/0896-6273(93)90149-1)
- 947 Mark, K.A., Soghomonian, J.-J., Yamamoto, B.K., 2004. High-Dose Methamphetamine Acutely  
948 Activates the Striatonigral Pathway to Increase Striatal Glutamate and Mediate Long-  
949 Term Dopamine Toxicity. *J Neurosci* 24, 11449–11456.  
950 <https://doi.org/10.1523/JNEUROSCI.3597-04.2004>
- 951 Martelli, D., Stanić, D., Dutschmann, M., 2013. The emerging role of the parabrachial complex  
952 in the generation of wakefulness drive and its implication for respiratory control.  
953 *Respiratory Physiology & Neurobiology* 188, 318–323.  
954 <https://doi.org/10.1016/j.resp.2013.06.019>
- 955 Mason, N.L., Kuypers, K.P.C., Müller, F., Reckweg, J., Tse, D.H.Y., Toennes, S.W., Hutten,  
956 N.R.P.W., Jansen, J.F.A., Stiers, P., Feilding, A., Ramaekers, J.G., 2020. Me, myself,  
957 bye: regional alterations in glutamate and the experience of ego dissolution with  
958 psilocybin. *Neuropsychopharmacol.* <https://doi.org/10.1038/s41386-020-0718-8>
- 959 Mathew, O.P., 2011. Apnea of prematurity: pathogenesis and management strategies. *J Perinatol*  
960 31, 302–310. <https://doi.org/10.1038/jp.2010.126>
- 961 Mattson, C.L., Tanz, L.J., Quinn, K., Kariisa, M., Patel, P., Davis, N.L., 2021. Trends and  
962 Geographic Patterns in Drug and Synthetic Opioid Overdose Deaths — United States,  
963 2013–2019. *MMWR Morb. Mortal. Wkly. Rep.* 70, 202–207.  
964 <https://doi.org/10.15585/mmwr.mm7006a4>
- 965 Mendelson, J., Uemura, N., Harris, D., Nath, R.P., Fernandez, E., Jacob, P., Everhart, E.T.,  
966 Jones, R.T., 2006. Human pharmacology of the methamphetamine stereoisomers. *Clin*  
967 *Pharmacol Ther* 80, 403–20. <https://doi.org/10.1016/j.clpt.2006.06.013>
- 968 Nelson, D.L., Lucaites, V.L., Wainscott, D.B., Glennon, R.A., 1999. Comparisons of  
969 hallucinogenic phenylisopropylamine binding affinities at cloned human 5-HT2A, 5-  
970 HT2B and 5-HT2C receptors: Naunyn-Schmiedeberg's *Arch Pharmacol* 359, 1–6.  
971 <https://doi.org/10.1007/PL00005315>

972 Neumeyer, J.L., Kula, N.S., Bergman, J., Baldessarini, R.J., 2003. Receptor affinities of  
973 dopamine D1 receptor-selective novel phenylbenzazepines. *European Journal of*  
974 *Pharmacology* 474, 137–140. [https://doi.org/10.1016/S0014-2999\(03\)02008-9](https://doi.org/10.1016/S0014-2999(03)02008-9)  
975 Nichols, D.E., 2016. Psychedelics. *Pharmacol Rev* 68, 264–355.  
976 <https://doi.org/10.1124/pr.115.011478>  
977 Nichols, D.E., 2004. Hallucinogens. *Pharmacology & Therapeutics* 101, 131–181.  
978 <https://doi.org/10.1016/j.pharmthera.2003.11.002>  
979 Nishimura, T., Takahata, K., Kosugi, Y., Tanabe, T., Muraoka, S., 2017. Psychomotor effect  
980 differences between l-methamphetamine and d-methamphetamine are independent of  
981 murine plasma and brain pharmacokinetics profiles. *J Neural Transm* 124, 519–523.  
982 <https://doi.org/10.1007/s00702-017-1694-y>  
983 Nishino, S., Mao, J., Sampathkumaran, R., Shelton, J., 1998. Increased dopaminergic  
984 transmission mediates the wake-promoting effects of CNS stimulants. *Sleep Res Online*  
985 1, 49–61.  
986 Oertel, B., Schneider, A., Rohrbacher, M., Schmidt, H., Tegeder, I., Geisslinger, G., Lötsch, J.,  
987 2007. The Partial 5-Hydroxytryptamine1A Receptor Agonist Buspirone does not  
988 Antagonize Morphine-induced Respiratory Depression in Humans. *Clinical*  
989 *Pharmacology & Therapeutics* 81, 59–68. <https://doi.org/10.1038/sj.clpt.6100018>  
990 Oertel, B.G., Felden, L., Tran, P.V., Bradshaw, M.H., Angst, M.S., Schmidt, H., Johnson, S.,  
991 Greer, J.J., Geisslinger, G., Varney, M.A., Lötsch, J., 2010. Selective antagonism of  
992 opioid-induced ventilatory depression by an ampakine molecule in humans without loss  
993 of opioid analgesia. *Clin Pharmacol Ther* 87, 204–211.  
994 <https://doi.org/10.1038/clpt.2009.194>  
995 Onimaru, H., Shamoto, A., Homma, I., 1998. Modulation of respiratory rhythm by 5-HT in the  
996 brainstem-spinal cord preparation from newborn rat. *Pflugers Arch* 435, 485–494.  
997 <https://doi.org/10.1007/s004240050543>  
998 Ortman, H.A., Newby, M.L., Acevedo, J., Siegel, J.A., 2021. The acute effects of multiple doses  
999 of methamphetamine on locomotor activity and anxiety-like behavior in adolescent and  
1000 adult mice. *Behavioural Brain Research* 405, 113186.  
1001 <https://doi.org/10.1016/j.bbr.2021.113186>  
1002 Pilowsky, P.M., Lung, M.S.Y., Spirovski, D., McMullan, S., 2009. Differential regulation of the  
1003 central neural cardiorespiratory system by metabotropic neurotransmitters. *Philos Trans*  
1004 *R Soc Lond B Biol Sci* 364, 2537–2552. <https://doi.org/10.1098/rstb.2009.0092>  
1005 Ramirez, J.M., Doi, A., Garcia, A.J., Elsen, F.P., Koch, H., Wei, A.D., 2012. The Cellular  
1006 Building Blocks of Breathing. *Compr Physiol* 2, 2683–2731.  
1007 <https://doi.org/10.1002/cphy.c110033>  
1008 Rauhut, A.S., Bialecki, V., 2011. Development and Persistence of Methamphetamine  
1009 Conditioned Hyperactivity in Swiss-Webster Mice. *Behav Pharmacol* 22, 228–238.  
1010 <https://doi.org/10.1097/FBP.0b013e328345f741>  
1011 Ren, J., Ding, X., Funk, G.D., Greer, J.J., 2009. Ampakine CX717 protects against fentanyl-  
1012 induced respiratory depression and lethal apnea in rats. *Anesthesiology* 110, 1364–1370.  
1013 <https://doi.org/10.1097/ALN.0b013e31819faa2a>  
1014 Ren, J., Ding, X., Greer, J.J., 2015. 5-HT1A Receptor Agonist Befiradol Reduces Fentanyl-  
1015 induced Respiratory Depression, Analgesia, and Sedation in Rats. *Anesthesiology* 122,  
1016 424–434. <https://doi.org/10.1097/ALN.0000000000000490>

1017 Richards, C.F., Clark, R.F., Holbrook, T., Hoyt, D.B., 1995. The effect of cocaine and  
1018 amphetamines on vital signs in trauma patients. *The Journal of Emergency Medicine* 13,  
1019 59–63. [https://doi.org/10.1016/0736-4679\(94\)00123-5](https://doi.org/10.1016/0736-4679(94)00123-5)

1020 Rothman, R.B., Baumann, M.H., 2003. Monoamine transporters and psychostimulant drugs.  
1021 *European Journal of Pharmacology* 479, 23–40.  
1022 <https://doi.org/10.1016/j.ejphar.2003.08.054>

1023 Rothman, R.B., Baumann, M.H., Dersch, C.M., Romero, D.V., Rice, K.C., Carroll, F.I., Partilla,  
1024 J.S., 2001. Amphetamine-type central nervous system stimulants release norepinephrine  
1025 more potently than they release dopamine and serotonin. *Synapse* 39, 32–41.  
1026 [https://doi.org/10.1002/1098-2396\(20010101\)39:1<32::AID-SYN5>3.0.CO;2-3](https://doi.org/10.1002/1098-2396(20010101)39:1<32::AID-SYN5>3.0.CO;2-3)

1027 Sokoloff, P., Giros, B., Martres, M.-P., Bouthenet, M.-L., Schwartz, J.-C., 1990. Molecular  
1028 cloning and characterization of a novel dopamine receptor (D3) as a target for  
1029 neuroleptics. *Nature* 347, 146–151. <https://doi.org/10.1038/347146a0>

1030 Sprouse, J., Reynolds, L., Li, X., Braselton, J., Schmidt, A., 2004. 8-OH-DPAT as a 5-HT7  
1031 agonist: phase shifts of the circadian biological clock through increases in cAMP  
1032 production. *Neuropharmacology* 46, 52–62.  
1033 <https://doi.org/10.1016/j.neuropharm.2003.08.007>

1034 Stephans, S.E., Yamamoto, B.K., 1995. Effect of repeated methamphetamine administrations on  
1035 dopamine and glutamate efflux in rat prefrontal cortex. *Brain Research* 700, 99–106.  
1036 [https://doi.org/10.1016/0006-8993\(95\)00938-M](https://doi.org/10.1016/0006-8993(95)00938-M)

1037 Stettner, G.M., Zanella, S., Hilaire, G., Dutschmann, M., 2008. 8-OH-DPAT suppresses  
1038 spontaneous central apneas in the C57BL/6J mouse strain. *Respiratory Physiology &*  
1039 *Neurobiology* 161, 10–15. <https://doi.org/10.1016/j.resp.2007.11.001>

1040 Stone, L.S., German, J.P., Kitto, K.F., Fairbanks, C.A., Wilcox, G.L., 2014. Morphine and  
1041 Clonidine Combination Therapy Improves Therapeutic Window in Mice: Synergy in  
1042 Antinociceptive but Not in Sedative or Cardiovascular Effects. *PLoS One* 9.  
1043 <https://doi.org/10.1371/journal.pone.0109903>

1044 Underhill, S.M., Hullihen, P.D., Chen, J., Fenollar-Ferrer, C., Rizzo, M.A., Ingram, S.L., Amara,  
1045 S.G., 2019. Amphetamines signal through intracellular TAAR1 receptors coupled to  
1046 Gα13 and GαS in discrete subcellular domains. *Mol Psychiatry*.  
1047 <https://doi.org/10.1038/s41380-019-0469-2>

1048 Underhill, S.M., Wheeler, D.S., Li, M., Watts, S.D., Ingram, S.L., Amara, S.G., 2014.  
1049 Amphetamine modulates excitatory neurotransmission through endocytosis of the  
1050 glutamate transporter EAAT3 in dopamine neurons. *Neuron* 83, 404–416.  
1051 <https://doi.org/10.1016/j.neuron.2014.05.043>

1052 van der Schier, R., Roozkrans, M., van Velzen, M., Dahan, A., Niesters, M., 2014. Opioid-  
1053 induced respiratory depression: reversal by non-opioid drugs. *F1000Prime Rep* 6.  
1054 <https://doi.org/10.12703/P6-79>

1055 Van Tol, H.H.M., Bunzow, J.R., Guan, H.-C., Sunahara, R.K., Seeman, P., Niznik, H.B., Civelli,  
1056 O., 1991. Cloning of the gene for a human dopamine D4 receptor with high affinity for  
1057 the antipsychotic clozapine. *Nature* 350, 610–614. <https://doi.org/10.1038/350610a0>

1058 Veasey, S.C., 2003. Serotonin agonists and antagonists in obstructive sleep apnea: therapeutic  
1059 potential. *Am J Respir Med* 2, 21–29. <https://doi.org/10.1007/BF03256636>

1060 Volkow, N.D., Chang, L., Wang, G.-J., Fowler, J.S., Franceschi, D., Sedler, M., Gatley, S.J.,  
1061 Miller, E., Hitzemann, R., Ding, Y.-S., Logan, J., 2001. Loss of Dopamine Transporters

1062 in Methamphetamine Abusers Recovers with Protracted Abstinence. *J Neurosci* 21,  
1063 9414–9418. <https://doi.org/10.1523/JNEUROSCI.21-23-09414.2001>  
1064 Vollenweider, F.X., Kometer, M., 2010. The neurobiology of psychedelic drugs: implications for  
1065 the treatment of mood disorders. *Nat. Rev. Neurosci.* 11, 642–651.  
1066 <https://doi.org/10.1038/nrn2884>  
1067 Wang, T., Yu, Z., Shi, Y., Xiang, P., 2015. Enantiomer Profiling of Methamphetamine in White  
1068 Crystal and Tablet Forms (Ma Old) Using LC–MS-MS. *J Anal Toxicol* 39, 551–556.  
1069 <https://doi.org/10.1093/jat/bkv060>  
1070 Zarrindast, M.-R., Ramezani-Tehrani, B., Ghadimi, M., 2002. Effects of adrenoceptor agonists  
1071 and antagonists on morphine-induced Straub tail in mice. *Pharmacology Biochemistry*  
1072 *and Behavior* 72, 203–207. [https://doi.org/10.1016/S0091-3057\(01\)00749-3](https://doi.org/10.1016/S0091-3057(01)00749-3)  
1073