Advancement in treatment, prevention and control strategies of avian pathogenic Escherichia coli : A literature review

Priyanka Devkota¹, Bishal Koirala², and Bhuminanda Devkota³

¹Duke University Human Vaccine Institute ²Dangisharan Basic Hospital ³Agriculture and Forestry University

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Abstract

Avian pathogenic *Escherichia coli* (APEC) poses significant health risks in poultry, leading to infections that impact productivity. APEC strains exhibit antibiotic resistance, complicating treatment options. Current vaccination strategies are limited in their protective scope, necessitating the exploration of innovative alternatives such as probiotics, bacteriophages, immune stimulants, and antimicrobial peptides (AMPs). AMPs show promise due to their rapid action against resistant bacteria and minimal resistance development. Additionally, small molecules have demonstrated effectiveness against various APEC serotypes, supporting the development of new antimicrobial therapies. Overall, a multifaceted approach addressing APEC's virulence factors and incorporating novel therapies is crucial for controlling colibacillosis in poultry and mitigating risks to human health.

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Priyanka Devkota¹, Bishal Koirala², Bhuminanda Devkota³

¹Lab Research Analyst, Duke Human Vaccine Institute, NC, USA

²Medical Officer, Dangisharan Basic Hospital, Dang, Nepal

³Department of Theriogenology, Faculty of Animal Science, Veterinary Science and Fisheries, Agriculture and Forestry University, Chitwan, Nepal

ABSTRACT

Key Clinical Message

Avian pathogenic *Escherichia coli* (APEC) poses significant health risks in poultry, leading to infections that impact productivity. APEC strains exhibit antibiotic resistance, complicating treatment options. Current vaccination strategies are limited in their protective scope, necessitating the exploration of innovative alternatives such as probiotics, bacteriophages, immune stimulants, and antimicrobial peptides (AMPs). AMPs show promise due to their rapid action against resistant bacteria and minimal resistance development. Additionally, small molecules have demonstrated effectiveness against various APEC serotypes, supporting the development of new antimicrobial therapies. Overall, a multifaceted approach addressing APEC's virulence factors and incorporating novel therapies is crucial for controlling colibacillosis in poultry and mitigating risks to human health.

2. DISCUSSION

2.1 Control and biosecurity measures

E. coli can spread between flocks mainly through dirty hatching eggs, so preventing contamination is crucial. Key steps include collecting eggs often, discarding damaged or soiled ones, and disinfecting eggs within two hours of laying. Cleaning the egg surface and using electrostatic sprayers can help reduce bacteria. UV light can kill E. coli without harming the eggs. Handling eggs carefully is important, as broken eggs can contaminate others. Good airflow in incubators and avoiding cross-contamination are also important. Chicks that may carry E. coli should be kept warm and fed by hand.^{4,5} To control APEC infections in poultry, several strategies are essential. Antibiotics and vaccines are commonly used, but newer options like probiotics and bacteriophages are being explored. Other innovative treatments such as immune stimulants, virulence inhibitors, and antimicrobial peptides target the bacteria directly or strengthen the chicken's natural defenses. Minimizing stress in poultry is crucial—keeping ammonia and dust levels low, providing good airflow, and maintaining comfortable temperature, humidity, and space helps birds stay healthier. Vaccinating against certain diseases and ensuring balanced nutrition also boosts immunity, further protecting chickens from infection.^{1,6} To prevent vertical APEC transmission in poultry breeding, breeders focus on developing resistant breeds, ensuring clean hatching eggs, and avoiding the use of eggs laid on the floor. Horizontal transmission is managed through controlled production cycles, removing weak chicks early, and maintaining rigorous sanitation. Strong biosecurity is vital—this includes chlorinating feed and water, disinfecting poultry houses and equipment, and restricting entry points for potential carriers like houseflies, wild birds, and rodents to keep APEC out of facilities.^{1,6,7}

2.2 Antibiotics

APEC strains are resistant to most antibiotics, with only a few carbapenems still effective—although resistance to imipenem has started appearing. These strains often withstand drugs like ampicillin, tetracycline, trimethoprim, sulfamethoxazole, and streptomycin. The high resistance to critical antibiotics, especially β -lactams and colistin, raises serious concerns about spreading these resistant bacteria and genes to humans through the food chain.⁸ The US and EU restrict non-therapeutic antibiotic use for growth and limit critical antibiotics in animal farming to help reduce antibiotic resistance risks.⁹ A study showed chickens treated with enrofloxacin showed better feed efficiency, lower death rates, healthier organs, and less bacterial presence than those given oxytetracycline. Oral enrofloxacin was especially effective, providing strong protection within 2 hours and lasting all day.¹⁰ Colistin, a last-resort antibiotic for Gram-negative infections, becomes more effective and less prone to resistance in *E. coli* when combined with small molecule adjuvants that target the pmrAB system.¹¹

2.3 "No Antibiotics Ever" Broiler Production Strategy

Pathogenic *E. coli* is present in both no antibiotics ever (NAE) broilers and their environment, which can increase the likelihood of colibacillosis outbreaks, particularly when birds are under stress. NAE programs can lower feed efficiency, slow growth, and negatively impact gut health, ultimately reducing overall poultry production. Research shows that raising broilers without antibiotics can lead to more pathogens, higher stress levels, and poorer growth. Additionally, stress makes birds more vulnerable to infections like colibacillosis, highlighting the challenges of NAE approaches.^{12,13}For decades, antimicrobial growth promoters were the primary approach to managing APEC, but the move to NAE production has resulted in more cases of colibacillosis. The prevalence of APEC-like virulent strains tends to be high overall, but it varies with the seasons, reaching peak in spring and dropping during the hotter months. Key environmental factors such as temperature, humidity, and housing conditions significantly influence APEC levels.^{12,13}

2.5 Probiotics and Prebiotics

Beneficial live microbes, known as probiotics, play a key role in preventing infections, while prebiotics serve as non-digestible ingredients that encourage the growth of healthy gut bacteria. Research indicates that including Lactobacillus plantarum B1 in the diets of broilers enhances levels of ileal mucosal secretory IgA and decreases pro-inflammatory cytokines. This not only improves growth performance but also reduces cecal $E. \ coli$ counts. Specifically, L. plantarum B1 has been effective in lowering $E. \ coli$ levels and bolstering immune responses.^{20,21} Other strains, such as Lactobacillus plantarum 15-1 with fructooligosaccharides and

Enterococcus faecalis-1, have also shown benefits, resulting in reduced mortality and improved immunity against APEC infections. A combination of multiple probiotic strains was able to eliminate deaths from APEC O78 and decrease bacterial levels in the liver and spleen. Furthermore, a commercial probiotic tested alongside a recombinant Salmonella vaccine demonstrated potential in enhancing both growth and immunity in poultry.²

2.6 Bacteriophages

The effectiveness of phage mixtures against APEC infections in chickens was tested using various methods of administration. Phages SPR02 and DAF6 significantly lowered mortality rates when given before or 24-48 hours after the APEC challenge.²² An oral or spray phage cocktail (phi F78E, phi F258E, phi F61E) reduced mortality in both experimental and natural infections. Another intramuscular cocktail (TM1, TM2, TM3, TM4) also decreased mortality, lowered APEC levels in the lungs, and improved body weight. Additionally, phage-loaded chitosan nanoparticles (C- Φ KAZ14 NPs) given orally reduced mortality and intestinal APEC levels while improving health and weight. Overall, these studies show that bacteriophages could be a promising way to control APEC infections in chickens, especially with the C- Φ KAZ14 NP treatment.^{2,22}

2.8 Virulence inhibitors

Virulence inhibitors function by disabling or weakening pathogens instead of just stopping their growth like antibiotics. They target virulence mechanisms, such as the quorum sensing (QS) system. These inhibitors can overcome issues associated with traditional antibiotics, like antibiotic resistance and harm to helpful bacteria, while making pathogens more vulnerable to the host's immune system. This makes them a promising alternative to standard antibiotics.^{2,23}

2.9 Growth Inhibitors

Growth inhibitors targeting bacterial membranes are promising new antibacterial agents with a lower risk of resistance. Baicalin, from *Scutellaria baicalensis*, reduced mortality and lung damage in chickens with APEC lung injury. Rutin lowered AI-2 secretion, reduced biofilm formation, and protected chicken lung cells. Additionally, andrographolide from *Andrographis paniculata* lowered inflammation in chicken lung cells. ^{11,24} Small molecules (SMs) are small compounds (200–500 Da) that can stop bacterial growth and enzymes. They work against different APEC types, including antibiotic-resistant ones, without harming eukaryotic cells. These findings support new treatments for APEC infections in poultry, which could also lower human ExPEC infections and reduce antibiotic resistance. Screening a small molecule library found several APEC growth inhibitors that effectively killed bacteria at low doses, improved survival in wax moth larvae, and reduced APEC levels.^{2,11,24}

2.10 Antimicrobial peptides

AMPs, also known as host defense peptides, are short, positively charged proteins found in many organisms, including humans. They have the ability to directly eliminate harmful microbes or enhance the body's immune response. With the growing issue of antibiotic resistance, AMPs are being recognized as potential therapeutic solutions. They provide quick and targeted action against bacteria that are resistant to traditional antibiotics and have a low risk of developing resistance, positioning them as excellent options for antibacterial treatments.²⁵ Bacteria often produce AMPs to fight off rival bacteria within their environment.²⁶Gram-positive and Gram-negative bacteria have membranes with negatively charged lipids that attract the positively charged AMPs. In Gram-negative bacteria, AMPs first need to breach the outer membrane, which is strengthened by cations like calcium and magnesium attached to lipopolysaccharides. AMPs likely use a method where they displace these cations, creating temporary openings in the outer membrane for entry.^{25,27} Antimicrobial peptides (AMPs) fight bacteria mainly by interacting with their membranes in different ways. Some peptides insert into the membrane to create pores, while others coat the membrane surface and disrupt it like a detergent when they reach a high concentration. Additionally, some peptides cause the membrane to bend around a central opening. AMPs can also block the creation of cell walls and inhibit the production of proteins or genetic material. Among the most studied AMPs are

insect-derived peptides, particularly cecropins, which effectively target bacteria like *E. coli* by damaging their membranes.²⁸ AMPs causes ion leakage and depolarization that lead to membrane dysfunction and rapid cell lysis.²⁹ Although bacteria are less likely to resist AMPs due to their multiple targets, recent studies show that resistance can develop under selective pressure, and AMPs also face challenges due to low stability and high production costs.^{25,30} In ovo treatment with D-CATH-2 reduced APEC mortality and bacterial load in chickens. Combining surfactin with amoxicillin enhanced its effectiveness, lowering mortality and APEC levels while boosting cytokine genes. Peptides like A3 and cecropin A-D-Asn also decreased *E. coli* in the chicken gut, suggesting antimicrobial peptides as alternatives or supplements to antibiotics in poultry.^{2,31}

3. CONCLUSION AND FUTURE PROSPECTIVES

APEC's diverse virulence factors interact to cause systemic infections in poultry, necessitating a comprehensive strategy that targets iron acquisition, quorum sensing, bacterial metabolism, and secretion systems for effective therapeutic development. APEC strains, especially ST95, ST131, and serogroups O1, O2, and O18, pose a risk of extra-intestinal infections in humans. With rising antibiotic resistance and the potential for transmission of resistant bacteria and genes to humans, developing antibacterials for animals that avoid cross-resistance with existing antibiotics is essential. An effective APEC vaccine offering cross-protection against multiple serotypes is also needed, and insights into APEC's virulence should guide the identification of new vaccine candidates. Additionally, alternative therapies, such as small molecule inhibitors and antimicrobial peptides targeting novel pathways, should be explored to control colibacillosis in poultry.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

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Priyanka Devkota : Conceptualization; data curation; supervision; validation; visualization; writing – original draft; writing – review and editing.

Bishal Koirala : Conceptualization; data curation; supervision; validation; visualization; writing – original draft; writing – review and editing.

Bhuminanda Devkota: Supervision; validation; visualization; writing- review and editing

REFERENCES

- 1. Dho-Moulin, M. and Fairbrother, J.M., 1999. Avian pathogenic Escherichia coli (APEC). Veterinary research , 30 (2-3), pp.299-316
- Kathayat D, Lokesh D, Ranjit S, Rajashekara G. Avian Pathogenic*Escherichia coli* (APEC): An Overview of Virulence and Pathogenesis Factors, Zoonotic Potential, and Control Strategies. *Pathogens* . 2021;10(4):467. Published 2021 Apr 12. doi:10.3390/pathogens10040467.
- Allan B, Wheler C, Köster W, et al. In Ovo Administration of Innate Immune Stimulants and Protection from Early Chick Mortalities due to Yolk Sac Infection. Avian Dis . 2018;62(3):316-321. doi:10.1637/11840-041218-Reg.1

- Kunert Filho, H. C., K. C. T. Brito, L. S. Cavalli, and B. G. Brito. "Avian Pathogenic Escherichia coli (APEC)-an update on the control." The battle against microbial pathogens: basic science, technological advances and educational programs, A Méndez-Vilas Ed 1 (2015): 598-618.
- Hu J, Afayibo DJA, Zhang B, et al. Characteristics, pathogenic mechanism, zoonotic potential, drug resistance, and prevention of avian pathogenic *Escherichia coli* (APEC). *Front Microbiol*. 2022;13:1049391. Published 2022 Dec 13. doi:10.3389/fmicb.2022.1049391
- Dziva F, Stevens MP. Colibacillosis in poultry: unravelling the molecular basis of virulence of avian pathogenic Escherichia coli in their natural hosts. Avian Pathol . 2008;37(4):355-366. doi:10.1080/03079450802216652.
- Christensen H, Bachmeier J, Bisgaard M. New strategies to prevent and control avian pathogenic Escherichia coli (APEC). Avian Pathol . 2021;50(5):370-381. doi:10.1080/03079457.2020.1845300.
- Osman KM, Kappell AD, Elhadidy M, et al. Poultry hatcheries as potential reservoirs for antimicrobialresistant Escherichia coli: A risk to public health and food safety. *Sci Rep*. 2018;8(1):5859. Published 2018 Apr 11. doi:10.1038/s41598-018-23962-7
- Xia H, Tang Q, Song J, Ye J, Wu H, Zhang H. A yigP mutant strain is a small colony variant of E. coli and shows pleiotropic antibiotic resistance. Can J Microbiol . 2017;63(12):961-969. doi:10.1139/cjm-2017-0347
- Chansiripornchai, Niwat. "Comparative efficacy of enrofloxacin and oxytetracycline by different administration methods in broilers after experimental infection with avian pathogenic Escherichia coli." The Thai Journal of Veterinary Medicine 39, no. 3 (2009): 231-236.
- Kathayat D, Antony L, Deblais L, Helmy YA, Scaria J, Rajashekara G. Small Molecule Adjuvants Potentiate Colistin Activity and Attenuate Resistance Development in *Escherichia coli* by Affectingpmr AB System. *Infect Drug Resist*. 2020;13:2205-2222. Published 2020 Jul 10. doi:10.2147/IDR.S260766
- Fancher, Courtney A., Li Zhang, Aaron S. Kiess, Pratima A. Adhikari, Thu TN Dinh, and Anuraj T. Sukumaran. "Avian pathogenic Escherichia coli and Clostridium perfringens: Challenges in no antibiotics ever broiler production and potential solutions." *Microorganisms* 8, no. 10 (2020): 1533.
- Fancher CA, Thames HT, Colvin MG, et al. Prevalence and Molecular Characteristics of Avian Pathogenic Escherichia coli in "No Antibiotics Ever" Broiler Farms. *Microbiol Spectr* . 2021;9(3):e0083421. doi:10.1128/Spectrum.00834-21
- Ghunaim H, Abu-Madi MA, Kariyawasam S. Advances in vaccination against avian pathogenic Escherichia coli respiratory disease: potentials and limitations. *Vet Microbiol*. 2014;172(1-2):13-22. doi:10.1016/j.vetmic.2014.04.019
- Watts A, Wigley P. Avian Pathogenic Escherichia coli : An Overview of Infection Biology, Antimicrobial Resistance and Vaccination. Antibiotics (Basel) . 2024;13(9):809. Published 2024 Aug 26. doi:10.3390/antibiotics13090809
- 16. Subedi M, Luitel H, Devkota B, et al. Antibiotic resistance pattern and virulence genes content in avian pathogenic Escherichia coli (APEC) from broiler chickens in Chitwan, Nepal [published correction appears in BMC Vet Res. 2018 May 22;14(1):166. doi: 10.1186/s12917-018-1453-9]. BMC Vet Res . 2018;14(1):113. Published 2018 Mar 27. doi:10.1186/s12917-018-1442-z
- Awad AM, El-Shall NA, Khalil DS, et al. Incidence, Pathotyping, and Antibiotic Susceptibility of Avian Pathogenic *Escherichia coli* among Diseased Broiler Chicks. *Pathogens*. 2020;9(2):114. Published 2020 Feb 12. doi:10.3390/pathogens9020114
- Grakh K, Mittal D, Prakash A, Jindal N. Characterization and antimicrobial susceptibility of biofilmproducing Avian Pathogenic Escherichia coli from broiler chickens and their environment in India. Vet Res Commun. 2022;46(2):537-548. doi:10.1007/s11259-021-09881-5
- McPeake SJ, Smyth JA, Ball HJ. Characterisation of avian pathogenic Escherichia coli (APEC) associated with colisepticaemia compared to faecal isolates from healthy birds. *Vet Microbiol*. 2005;110(3-4):245-253. doi:10.1016/j.vetmic.2005.08.001
- 20. Wang S, Peng Q, Jia HM, et al. Prevention of Escherichia coli infection in broiler chickens with Lactobacillus plantarum B1. *Poult Sci*. 2017;96(8):2576-2586. doi:10.3382/ps/pex061
- 21. Joseph J, Zhang L, Adhikari P, Evans JD, Ramachandran R. Avian Pathogenic Escherichia coli

(APEC) in Broiler Breeders: An Overview. *Pathogens* . 2023;12(11):1280. Published 2023 Oct 26. doi:10.3390/pathogens12111280

- Kaikabo AA, AbdulKarim SM, Abas F. Evaluation of the efficacy of chitosan nanoparticles loaded ΦKAZ14 bacteriophage in the biological control of colibacillosis in chickens. *Poult Sci*. 2017;96(2):295-302. doi:10.3382/ps/pew255
- 23. Dou X, Gong J, Han X, et al. Characterization of avian pathogenic Escherichia coli isolated in eastern China. *Gene*. 2016;576(1 Pt 2):244-248. doi:10.1016/j.gene.2015.10.012
- Kathayat D, Helmy YA, Deblais L, Rajashekara G. Novel small molecules affecting cell membrane as potential therapeutics for avian pathogenic Escherichia coli. *Sci Rep*. 2018;8(1):15329. Published 2018 Oct 17. doi:10.1038/s41598-018-33587-5
- Mahlapuu M, Håkansson J, Ringstad L, Björn C. Antimicrobial Peptides: An Emerging Category of Therapeutic Agents. Front Cell Infect Microbiol. 2016;6:194. Published 2016 Dec 27. doi:10.3389/fcimb.2016.00194
- Hassan M, Kjos M, Nes IF, Diep DB, Lotfipour F. Natural antimicrobial peptides from bacteria: characteristics and potential applications to fight against antibiotic resistance. J Appl Microbiol . 2012;113(4):723-736. doi:10.1111/j.1365-2672.2012.05338.x
- 27. Clifton LA, Skoda MW, Le Brun AP, et al. Effect of divalent cation removal on the structure of gramnegative bacterial outer membrane models. *Langmuir*. 2015;31(1):404-412. doi:10.1021/la504407v
- Wang S, Zeng X, Yang Q, Qiao S. Antimicrobial Peptides as Potential Alternatives to Antibiotics in Food Animal Industry. Int J Mol Sci . 2016;17(5):603. Published 2016 May 3. doi:10.3390/ijms17050603
- Li Y. Recombinant production of antimicrobial peptides in Escherichia coli: a review [published correction appears in Protein Expr Purif. 2012 Mar;82(1):252]. Protein Expr Purif. 2011;80(2):260-267. doi:10.1016/j.pep.2011.08.001
- Lofton H, Pränting M, Thulin E, Andersson DI. Mechanisms and fitness costs of resistance to antimicrobial peptides LL-37, CNY100HL and wheat germ histones. *PLoS One* . 2013;8(7):e68875. Published 2013 Jul 23. doi:10.1371/journal.pone.0068875
- 31. Antão EM, Glodde S, Li G, et al. The chicken as a natural model for extraintestinal infections caused by avian pathogenic Escherichia coli (APEC). *Microb Pathog* . 2008;45(5-6):361-369. doi:10.1016/j.micpath.2008.08.005