

1 **Title Page**

2 **Title:** Using *gridCoal* to assess whether standard population genetic theory holds in the  
3 presence of spatio-temporal heterogeneity in population size

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## 30 **Abstract**

31 Spatially explicit population genetic models have long been developed, yet have rarely been  
32 used to test hypotheses about the spatial distribution of genetic diversity or the expected neutral  
33 levels of genetic divergence between populations. Here, we use spatially explicit coalescence  
34 simulations to explore the properties of the island model and the two-dimensional stepping  
35 stone model under a wide range of scenarios with spatio-temporal variation in deme size. We  
36 avoid the simulation of genetic data, using the fact that under the studied models, summary  
37 statistics of genetic diversity and divergence between demes can be approximated from  
38 coalescence times. We perform the simulations using *gridCoal*, a flexible spatial wrapper for the  
39 software *msprime* (Kelleher *et al.*, 2016) developed herein. In *gridCoal*, deme sizes can change  
40 arbitrarily across space and time, and migration rates between individual demes can be  
41 specified. We identify the different factors that can cause a deviation from the theoretical  
42 expectations, such as the simulation time in comparison to the effective deme size and the  
43 spatio-temporal autocorrelation across the grid. Our results highlight that  $F_{ST}$ , a measure of the  
44 strength of population structure, principally depends on recent demography, which makes it  
45 robust to temporal variation in deme size. We also warn that predicting genetic diversity from  
46 coalescence times requires a much longer run time than needed for the estimation of  $F_{ST}$ .  
47 Finally, we illustrate the use of *gridCoal* on a real-world example, the range expansion of silver  
48 fir (*Abies alba* Mill.) since the Last Glacial Maximum, using different degrees of spatio-temporal  
49 variation in deme size.

## 50 **1 Introduction**

51 The distribution and dynamics of genetic diversity within species are shaped by a myriad of  
52 evolutionary and ecological processes acting across different spatial and temporal scales

53 (Ellegren & Galtier, 2016). Although the role of space and, in particular, spatial autocorrelation  
54 in allele frequencies have been recognised since the dawn of population genetics (Wright, 1943;  
55 Malécot, 1948; Felsenstein, 1976; Sokal & Wartenberg, 1983), disproportionately more theoretical  
56 and methodological developments have been focused on understanding the effect of temporal  
57 changes in population size and gene flow among populations without spatial structure (*e.g.* Hey  
58 & Nielsen, 2007). Further, most statistical tools have been developed to detect past population  
59 size changes, either by testing different hypotheses such as exponential growth and bottlenecks  
60 (*e.g.* Excoffier *et al.*, 2013), or by using Bayesian methods to detect arbitrary population size  
61 changes from whole genome sequences (*e.g.* Drummond *et al.*, 2005). Researchers in landscape  
62 genetics have set the aim of overcoming the limitation imposed by population genetics methods  
63 that rely on the assumption of non-spatial and discrete populations (Manel *et al.*, 2003).  
64 However, the field has been mostly influenced by meta-population models (Hanski & Gilpin,  
65 1991) and by spatial statistics and geo-statistics (*e.g.* Guillot *et al.*, 2005; Smouse *et al.*, 2008;  
66 Forester *et al.*, 2016), rather than by population genetic theory.

67       There is increasing evidence that ignoring space can lead to biases and erroneous inferences  
68 (Bradburd & Ralph, 2019). Indeed, simulation studies have shown that ignoring isolation by  
69 distance can lead to false positives in efforts to detect hierarchical population structure and loci  
70 under selection (Meirmans, 2012). Similarly, ignoring space can severely bias common  
71 population genetics summary statistics, especially when the local effective population size (*i.e.*  
72 neighbourhood size) is small (Battey *et al.*, 2020). However, spatially explicit models are often  
73 mathematically intractable and theoretical predictions are valid only under limited conditions  
74 (Slatkin, 1985; Barton *et al.*, 2002; Kelleher *et al.*, 2014; Bradburd & Ralph, 2019). This is  
75 particularly true for spatially continuous models. For example, the coalescence process under  
76 the continuous space isolation-by-distance (IBD) model (Wright, 1943; Malécot, 1948) can be  
77 approximated using the Lambda-Fleming-Viot algorithm (Barton *et al.*, 2010a,b). However,  
78 results are inconsistent with large-scale patterns and often predict lower diversity than expected

79 from census numbers (Barton *et al.*, 2010a), although some of these issues have been solved by  
80 the subsequently introduced model of extinction and recolonisation (Kelleher *et al.*, 2014).  
81 Discrete spatial models are worse at capturing reality but are mathematically more tractable  
82 (Cox *et al.*, 2002), and several equivalences have been shown across island models,  
83 two-dimensional (2D) stepping stone models (Kimura, 1953) and IBD models assuming infinite  
84 or finite populations and the absence or presence of mutations (Malécot, 1975; Felsenstein, 1976;  
85 Slatkin, 1985). In particular, a 2D stepping stone model can approximate the decrease in genetic  
86 correlation with increasing distance of continuous space (Malécot, 1955; Kimura & Weiss, 1964),  
87 and when a sufficiently large lattice is used, it can produce summary statistics similar to those  
88 from a continuous space model (Battey *et al.*, 2020).

89 Efficient spatially explicit simulators have recently been developed, both those using a  
90 forward in time approach, such as *SLiM* (Haller & Messer, 2019), and those using a mixture of  
91 forward and coalescent approaches, such as *SPLATCHE 3* (Currat *et al.*, 2019). These  
92 developments have increasingly enabled the inclusion of space in population genetics  
93 applications (e.g. Battey *et al.*, 2020; González-Serna *et al.*, 2019; Ortego & Knowles, 2020;  
94 Quilodrán *et al.*, 2019). However, these spatial simulators can be challenging to parametrise.  
95 This is particularly true for forward simulations, as they require background knowledge on the  
96 demography, mating system and dispersal patterns. Backward, coalescent simulations have the  
97 advantage of allowing likelihood calculations while only tracing back the genealogy of sampled  
98 individuals (Felsenstein, 1992). Nevertheless, they still also require that past population size  
99 changes are known or follow a predictable pattern, such as constant size, expansion, decline or  
100 bottleneck. Ecological models, such as species distribution models coupled with recently  
101 developed paleo-climatic databases (e.g. Lima-Ribeiro *et al.*, 2015; Cook *et al.*, 2015; Karger *et al.*,  
102 2021), may be used to predict past species distributions in a spatially and temporally explicit  
103 manner (e.g. Tallavaara *et al.*, 2015; Wang *et al.*, 2017; Lima-Rezende *et al.*, 2019). Such time  
104 series of species distribution maps can provide potential input parameters for spatially explicit

105 coalescent simulations (He *et al.*, 2013).

106 The aim of this work is to explore the properties of island models and 2D stepping stone  
107 models under a wide range of scenarios with spatio-temporal variation in population size. To  
108 this end, we first develop a spatially explicit coalescent wrapper, *gridCoal*, for the most efficient  
109 coalescent simulator currently available, *msprime* (Kelleher *et al.*, 2016). In *gridCoal*, we  
110 implement the 2D stepping stone model with population sizes that may vary in space and time,  
111 and with migration rate that may differ between demes. *gridCoal* is different in several ways  
112 from *SPLATCHE 3*, the spatially explicit coalescent simulator that is currently used most  
113 frequently. Most importantly, in *gridCoal* (i) there is no forward simulation phase; (ii) demes do  
114 not follow a logistic growth model (as in *SPLATCHE 3*), instead instantly increasing or  
115 decreasing to user-defined deme sizes; and (iii) colonisation is possible from several seeds  
116 (without defining "layers" as in *SPLATCHE 3*). Further, unlike *SPLATCHE 3*, *gridCoal* does not  
117 simulate genetic marker data. Instead, we exploit the fact that under the 2D stepping stone  
118 model, summary statistics of genetic diversity and divergence between populations can be  
119 approximated from the coalescence times (Slatkin, 1991; Ralph *et al.*, 2020). After developing the  
120 coalescent wrapper, we use *gridCoal* to simulate various scenarios of spatial and temporal  
121 changes in population size and compare their outcome with theoretical expectations of the  
122 island models and 2D stepping stone models. In particular, we compare simulations with  
123 expectations for the mean coalescence time, which is proportional to the effective population  
124 size  $N_e$  and the amount of genetic diversity, for a measure of the strength of population  
125 structure  $F_{ST}$ , and for isolation-by-distance patterns. Our simulated scenarios include simplified  
126 and biologically realistic cases of population movement and expansion, where the spatial and  
127 temporal autocorrelation are decoupled. Finally, we illustrate the use of *gridCoal* on a real-world  
128 example, the range expansion of silver fir (*Abies alba* Mill.) since the Last Glacial Maximum,  
129 using different degrees of spatio-temporal variation in deme size.

## 2 Materials and Methods

### 2.1 The simulation tool: *gridCoal*

We developed a 2D stepping stone coalescent simulation tool, *gridCoal* (Appendix A), based on *msprime* (Kelleher *et al.*, 2016). Space is represented by a rectangular grid (size  $L \times L$  in most of our simulations). Each grid cell contains a single panmictic population, hereafter referred to as a deme, whose size ( $N$ ) can change in time at equally spaced time points comprising a given number of generations. A forward migration matrix defines the fraction of individuals that migrate from one deme to its four neighbouring demes. Forward migration rates ( $m$ ) are independent from deme sizes, and they can be symmetric or asymmetric between demes, and homo- or heterogeneous across space. The backward migration matrix, required for the coalescent process, contains elements that specify the fraction of individuals in a given deme that have a parent in another deme. Backward migration rates are calculated for each time point based on the deme sizes and the forward migration matrix.

The coalescent process consists of two phases: a scattering phase in the recent past with the fully defined demographic history of individual demes, and a collecting phase in the more distant past assuming panmictic population(s). While spatial structure is important in the scattering phase, its effect becomes smaller and even negligible in the collecting phase, which can be thus approximated by the standard coalescent process (Wakeley, 1998, 1999). This implies that it is unnecessary to run the spatially explicit simulations until all lineages coalesce; before that point, the lineages can instead be combined to a single or a few spatially non-explicit panmictic populations. It is, nevertheless, possible to specify multiple ancestral populations with low migration among them, and thus account for the spatial aspect of the collecting phase.

Time is managed in *gridCoal* using three parameters: (1) the number of time points  $T$  when the deme sizes are defined, (2) the time period that elapses between two time steps  $dt$  (in years, or

154 other suitable time units), and (3) the generation time  $gt$  (in years or other time units, compatible  
155 with  $dt$ ; see also Table 1). Thus,  $T \times dt$  determines the length of the spatially explicit phase. To  
156 achieve the highest efficiency, this time should be equal to the scattering phase. After this phase,  
157 all lineages are combined into one or more panmictic, spatially non-explicit, so-called ancestral  
158 population(s). This non-spatial phase ensures that all lineages coalesce even when the product of  
159 the effective population size and migration rate ( $Nm$ ) is small, and it facilitates the simulation of  
160 different refugial populations that may colonise different parts of the grid. Note that mutation is  
161 assumed to be negligible throughout this work.

## 162 **2.2 Simulated scenarios**

163 Here, we provide a brief summary of the simulated scenarios, while more details can be found in  
164 Appendix B. Across all scenarios, we used a forward migration rate that is constant in time and  
165 homogeneous across the grid. Simulations were run with an average deme size of  
166  $N \in (10, 50, 100, 250, 500)$ , with migration rate  $m \in (10^{-5}, 10^{-3}, 10^{-2}, 10^{-1}, 10^0)$  between  
167 neighbouring cells (see Table 1 for explanations of terms and symbols). To analyse the effect of  
168 spatial heterogeneity, we simulated various maps differing in the amount of spatial variation and  
169 autocorrelation in deme size (Fig. 1). Our simulated scenarios ranged from a homogeneous map,  
170 where all demes have the same size, to a map with large variance in deme size, with deme sizes  
171 drawn from a uniform distribution. To investigate the effect of temporal changes, we simulated  
172 scenarios with various demographic histories: static scenarios with fixed deme sizes in time;  
173 simple demographics, where all demes changed in the same manner on average, such as  
174 undergoing an expansion, decline or bottleneck; and more biologically realistic scenarios of  
175 colonisation from one side or from "seeds" (such as refugia), or range expansion and shift (Fig.  
176 1). For each scenario and combination of  $N$  and  $m$  in a factorial design, we ran 1000 replicates.

177 We sampled lineages across the grid in two different ways. In order to estimate the  
178 within-deme coalescence time, we sampled two lineages from each deme. In contrast, for

179 studying between-deme coalescence times, we sampled lineages along a horizontal line ( $L$   
180 demes) in the middle of the grid.

### 181 **2.3 Summary statistics**

182 Hudson *et al.* (1990) and Slatkin (1991) noted the close relationship between the probabilities of  
183 identity by descent and coalescence times, which makes it possible to bypass the simulation of  
184 genetic data, instead estimating diversity and divergence statistics directly from coalescence times  
185 (Ralph *et al.*, 2020). Additionally, for such calculations it is sufficient to simulate the genealogies  
186 of two lineages per deme.

187 **Coalescence times** For low and high  $Nm$ , the individual demes and the grid as a whole,  
188 respectively, are nearly panmictic, the distribution of coalescence times is close to exponential,  
189 and most lineages coalesce within the scattering phase. Under these conditions, the maximum  
190 likelihood estimate of the mean coalescence time is the sample mean. In contrast, for  
191 intermediate  $Nm$ , the probability that lineages migrate away from their present-time demes  
192 before coalescing is high, but the probability that they meet again and coalesce within the  
193 spatially explicit phase of the simulation is low. As a result, the distribution of coalescent times  
194 is no longer exponential and the sample mean is an incorrect estimate of the coalescent time. In  
195 order to still consider these intermediate  $Nm$  values, we use the sample median, which is  
196 expected to be less sensitive to the missing tail values (Fig. S1).

197 We calculated the expected within-deme coalescence time ( $T_W$ ) as the coalescence time of  
198 two lineages from the same deme. Assuming a mutation model, measures of within-population  
199 genetic diversity can be calculated from  $T_W$ . Here, we simply used  $T_W$  as a proxy for within-deme  
200 diversity and plotted it as a heatmap across the grid.  $T_B$  is the coalescence time between any two  
201 lineages from two different demes, and  $T_T$  is the coalescence time of any two lineages across the  
202 grid.

203 **Population structure ( $F_{ST}$ )** Under the infinite island model, the extent of population structure  
 204 can be described as in Whitlock & McCauley (1999):

$$F_{ST} = \frac{1}{4Nm + 1} \quad (1)$$

205 Under Kimura's 2D stepping stone model, given a homogeneous migration rate and equal  
 206 sized demes,  $F_{ST}$  can be defined as follows (Maruyama, 1977; Cox *et al.*, 2002):

$$F_{ST} = \frac{\frac{L^2 \log L}{2\pi\nu\sigma^2}}{\frac{L^2 \log L}{2\pi\nu\sigma^2} + 2NL^2} \quad (2)$$

207 where  $\sigma = 1/2$  is the average axial parent-offspring distance,  $\nu = 4\mu$ , and  $L$  is the grid size.  
 208 The value  $2\pi\nu\sigma^2$  is the neighbourhood size, which is the local panmictic unit that determines the  
 209 amount of variation between populations at the migration-drift equilibrium; thus, it is equivalent  
 210 to  $Nm$  in the island model. Note that when  $\frac{\log L}{2\pi\nu\sigma^2} \ll 2N$ , equation (2) simplifies to  $F_{ST} = \frac{\log L}{4N\pi\nu\sigma^2}$   
 211 (Cox *et al.*, 2002).

212 The link between  $F_{ST}$  and coalescent theory was introduced by Slatkin (1991):

$$F_{ST} = \frac{T_T - T_W}{T_T} \quad (3)$$

213 where  $T_T$  is the average total coalescence time and  $T_W$  is the within-deme coalescence time  
 214 averaged across demes. We refer to equation (3) as the *global* (population-wide)  $F_{ST}$ , which  
 215 measures the strength of the population structure and can be compared across different  
 216 simulated scenarios, and we use this definition throughout the manuscript. Note that  
 217 approximating summary statistics of genetic diversity and  $F_{ST}$  from coalescence times holds  
 218 only when the mutation rate is low, and migration is possible to neighbouring demes only  
 219 (Slatkin, 1985).

220 **Genetic distance ( $F^*$ )** In order to investigate the genetic differentiation between pairs of  
 221 demes and its relationship with physical distance, we used a measure of genetic distance based  
 222 on coalescence times. If only two demes are considered, Equation 3 transforms into:

$$F^* = \frac{T_B - T_W}{T_B + T_W} \quad (4)$$

223 where  $T_B$  is the mean coalescence time for two lineages sampled from different demes, and  $T_W$   
 224 is the mean within-deme coalescence time. Slatkin (1993) pointed out that this equation is not  
 225 appropriate to assess the strength of population structure in general, but it is a useful measure of  
 226 the genetic distances between demes. We used  $F^*$  between all pairs of sampled demes against the  
 227 physical (Euclidean) distance between demes to assess isolation-by-distance patterns across the  
 228 grid. Note that if  $T_W \approx T_B$  (which is the case for large  $Nm$ ),  $F^* = \frac{F_{ST}}{2 - F_{ST}}$  and  $F^*$  thus does not  
 229 provide any more information about the population structure than  $F_{ST}$ .

230 **Effective population size ( $N_e$ )** Under the island model,  $N_e$  is

$$N_e = N_s \left( 1 + \frac{(s-1)^2}{4N\nu s^2} \right), \quad (5)$$

231 where  $\nu = 4m$  (the total migration rate for each grid cell), and  $s$  is the number of demes in the  
 232 island model. While,  $N_e$  under two dimensional stepping stone model can be calculated as Cox  
 233 *et al.* (2002):

$$N_e = \frac{L^2 \log(L)}{4\pi\sigma^2\nu} \quad (6)$$

234 Effective population sizes predicted from simulations were obtained by halving the  
 235 coalescence time of lineages from the same deme.

## 3 Results

### 3.1 Coalescence times

The expected coalescence time of two samples drawn from the same deme is  $T_W = 2N$ , where  $N$  is the total number of diploid individuals in the deme. This result is independent of the migration matrix if all demes are connected by migration. Under the island model with  $d$  demes each containing  $N$  individuals, the expected coalescence time for two samples from the same deme is  $2Nd$  (Strobeck, 1987), which is higher than  $2N$  as a result of lineages escaping before coalescence occurs. Under the 2D stepping stone model the expected coalescence time is  $2NL^2$  (Cox *et al.*, 2002). However, in a 2D stepping stone model, where only the four neighbours are connected, strong local differentiation across demes occurs when  $Nm < 1$  (Kimura & Maruyama, 1971). In contrast, when  $Nm > 1$  local differentiation is less pronounced, and when  $Nm \geq 4$  the whole grid behaves like a single panmictic population (Kimura & Maruyama, 1971). Our simulations helped us to explore the effect of spatial heterogeneity in deme size on these theoretical predictions.

Spatial heterogeneity can be ignored when  $m = 0$  and thus each individual deme behaves like a panmictic population. In these cases, the expected coalescence time for two samples taken from the same deme is independent of the spatial heterogeneity of the grid and is thus expected to be  $2N$  (Fig. 2a and d). However, for the largest deme size considered ( $N = 500$ ),  $T_W$  was on average larger than  $2N$ . At the other extreme, when  $m$  was one, the whole grid behaved like a single panmictic population. Here,  $T_W$  was decoupled from the local deme size and was, on average, equal to  $2NL^2$  (Fig. 2c). Additionally, when there was large spatial variance in deme size across the grid, the uniform map, the coalescence time was systematically underestimated (Fig. 2f). This was because the spatial heterogeneity decreased the total effective population size across the grid.

For low migration rates, the expected coalescence time of two samples from the same deme

260 should be  $T_W = 2NL^2$ , independent of  $m$  (Maruyama, 1971). When  $m$  is very small, lineages  
 261 coalesce mostly within demes, on average, in time  $2N$ . However, very rarely, they escape and  
 262 coalesce only in time  $2NL^2/m$ , which results in a mean coalescence time of  $T_W = 2NL^2$ . The  
 263 spatially explicit phase of our simulations was not long enough for these samples to coalesce  
 264 after they escaped from the deme, and these were thus forced to coalesce sooner, leading to  
 265 underestimation of the average coalescence time (Fig. 2b and e). The problem of escaping  
 266 lineages matters the most in the transition phase from low to high  $Nm$ . Recall that in this part  
 267 of the parameter space we could not estimate the sample mean; therefore, we show the median  
 268  $T_W$  instead (Fig. 2). Note that these results should be treated with caution and cannot be  
 269 compared with any theoretical expectations. We found that for  $Nm \leq 0.05$ ,  $T_W$  was best  
 270 predicted by  $2\bar{N}$  (Fig. 2b;  $m = 0.001$  and  $N = 50$ ). Then, for  $Nm = 0.1$ , our simulations showed  
 271 that the median coalescence time was best predicted by twice the neighbourhood size, i.e. the  
 272 size of the deme plus its four neighbours (Fig. 2e;  $m = 0.001$  and  $N = 100$ ). However, we found  
 273 that already at  $Nm = 0.5$  the coalescence time was best predicted by  $2NL^2$  (Fig. 2; see also Fig.  
 274 S1b and e), suggesting that the transition phase is fast, which is in agreement with previous  
 275 observations by Kimura & Maruyama (1971).

### 276 3.2 Global $F_{ST}$

277 The island model (eq. 1) and 2D stepping stone model (eq. 2) provide expectations for the  
 278 strength of population structure ( $F_{ST}$ ) in subdivided populations. Here, we explored the  
 279 robustness of these predictions with respect to the spatio-temporal heterogeneity in deme size.  
 280 We found that all simulated scenarios deviated the most from theoretical predictions for  
 281 intermediate migration rates (or  $Nm$ ), where the predictions of the two models also differed the  
 282 most (Fig. 3a, b and c). Not surprisingly, the island model provided, on average, a better  
 283 approximation than the spatially explicit 2D stepping stone model when the deme sizes were  
 284 drawn from a uniform distribution across the grid, thus when there was no spatial

285 autocorrelation in deme size (Fig. 3a). In contrast, when deme sizes were homogeneous across  
286 space, and thus the spatial autocorrelation was maximised,  $F_{ST}$  was closer to the 2D stepping  
287 stone model predictions (Fig. 3a).  $F_{ST}$  of the clustered and low variance maps were in between  
288 the two model predictions.  $F_{ST}$  also varied considerably across replicates, with the largest  
289 variation occurring for the uniform map among all static scenarios considered (Fig. 3d).

290 Across the changing scenarios, we observed a consistent bias: scenarios where the mean  
291 deme size decreased over time gave a lower  $F_{ST}$ , while scenarios where the mean deme size  
292 increased (expanding population) gave a higher  $F_{ST}$  in comparison to the static equivalent  
293 scenarios (Fig. 3b and c). Similar to with the uniform static map,  $F_{ST}$  was relatively close to the  
294 predictions of the island model under the changing scenarios that ended in a uniform map (Fig.  
295 3b). More unexpectedly, under the realistic changing scenarios, where we decoupled the spatial  
296 and temporal autocorrelation, on average,  $F_{ST}$  did not deviate more from the island model  
297 prediction than the simple changing scenarios for the studied parameter combinations (Fig. 3c).  
298 These realistic changing scenarios also provided a relatively close match to their static  
299 equivalents (Fig. 3c). An exception is the range expansion and shift scenario. This is because  
300 here the theoretical expectation is shown for the average  $N$  across the grid, which is lower than  
301 the row of sampled demes situated in the middle of the grid (Fig. 1). Finally, the variation in  $F_{ST}$   
302 across replicates was important, and  $F_{ST}$  for different realisations of the same map overlapped  
303 between values of  $Nm$  that were one order of magnitude different, especially for low and  
304 intermediate values (Fig. 3d, e and f). The sampling variance in  $F_{ST}$  also increased with spatial  
305 variance in deme size across the grid, with the highest values corresponding to the two clustered  
306 maps (Fig. 3d and f).

### 307 **3.3 Genetic distance ( $F^*$ )**

308 Varying  $N$  and  $m$  across a homogeneous map showed that increasing the deme size and/or the  
309 migration rate led, as expected from equation (1), to weaker differentiation between demes (Fig.

310 S2). The degree of spatial variance in deme size affected both the average genetic distance between  
311 demes and the shape of the isolation by distance curves (Fig. 3g). Maps with homogeneous deme  
312 size had the lowest and flattest isolation by distance curves. Note that these can be treated as a  
313 baseline expectation under the 2D stepping stone model (Slatkin, 1993). The uniform map gave  
314 higher  $F^*$  values across all the distance classes, i.e. the isolation by distance curve was shifted  
315 upwards, because the compared pairs of demes had, on average, a different size. The clustered  
316 map resulted in a lower mean  $F^*$  for small distance classes and a higher  $F^*$  for larger distance  
317 classes, meaning that the isolation by distance curve was steeper. This was because pairs of demes  
318 located close to each other tended to have similar sizes, and those for large distance classes often  
319 had different sizes. Varying deme classes also caused a large variance in the genetic distance  
320 across replicates (Fig. 3g, h and i).

321 Demographic, i.e. temporal, changes introduced a bias in the same direction as in  $F_{ST}$ :  
322 scenarios where the mean deme size decreased over time had a lower  $F^*$ , while scenarios with  
323 increasing average deme size had a larger  $F^*$  value in comparison to a static uniform map.  
324 However, the shape of the isolation by distance curve did not change (Fig. 3h). The realistic  
325 changing scenarios all had increasing deme sizes, so we observed the same upward bias as  
326 before (Fig. 3i). The fact that the population sizes were changing had the strongest influence on  
327  $F^*$  when the spatial and temporal autocorrelation was the most decoupled, i.e. for the side  
328 colonisation scenario, and for large distance classes (Fig. 3i).

### 329 **3.4 Effect of spatial and temporal resolution**

330 Our simulations were carried out on a finite square grid of  $L \times L$  (not a torus), which implies a  
331 finite number of demes and that demes on the edge of the grid had only two or three neighbours.  
332 Not surprisingly, we found that  $F_{ST}$  estimated from a larger grid provided a better fit to the  
333 predictions of the island model and the 2D stepping stone model (Fig. 4a). Further, we found that  
334 there was an edge effect, which led to the overestimation of  $F^*$  for demes that were  $L$  or nearly  $L$

335 steps away from each other (Fig. 4b). Analysing  $F^{*}$  against distance from samples in the middle  
336 the grid allowed us to disentangle the effect of grid size from the edge effect. We found that close  
337 to the edges the genetic distance is overestimated between the demes, mainly due to edge effects,  
338 while grid size principally influences the precision of the estimates, i.e. larger grids provide more  
339 precise estimates of  $F^{*}$  for a given distance class (Fig. 4c).

340 We investigated the effect of temporal resolution in the case of a simple changing scenario  
341 where all demes increased linearly in size (Fig. 1). The coarser time resolution ( $T = 5$ ) did not  
342 have a noticeable effect on the estimation of the mean coalescence time within demes, but the  
343 between-deme coalescence times were systematically overestimated (not shown). As a result, the  
344 genetic distances between demes were also overestimated (Fig. 4e). This is because when  $T = 5$ ,  
345 the population size at any time is larger than in the finer time-resolution scenario ( $T = 25$ ). Time  
346 resolution is also important in more complex setting such as range expansion and shift (Fig. 1).

347 The time necessary for lineages to coalesce during the spatially explicit phase of the  
348 simulations may become a limitation in practical applications. When the spatially explicit phase  
349 is too short compared with the deme sizes, the coalescence time between lineages is determined  
350 by the non-spatial coalescence process of the panmictic ancestral population. Extremely long  
351 simulations may be required to reliably estimate the coalescence time when the deme sizes are  
352 large. Fig. 4e shows estimates of  $N_e$  calculated as half of the mean total coalescence time. In  
353 contrast, Fig. 4f demonstrates that it is possible to obtain relatively precise estimates of  $F_{ST}$  with  
354 much shorter simulation times. This is because  $F_{ST}$  is defined as a ratio of coalescence times  
355 and the biases cancel out. Indeed, both the estimation of within-deme and total coalescence  
356 times are biased because of the same process, i.e. the limited length of the spatially explicit  
357 phase, which means that their distributions are missing the same amount from the tails on the  
358 right side. This result also highlights that  $F_{ST}$  is dependent only on recent demographic events  
359 and is independent of the deeper ancestry, which makes it a useful measure.

### 3.5 Application example: *Abies alba* post-glacial colonisation history

Silver fir (*Abies alba* Mill.) is a coniferous tree species that has progressively colonised the mountainous regions of Europe from different refugia since the Last Glacial Maximum (LGM, 21 kyrs BP). While the exact location of the refugia are debated, it is generally agreed that the Central and/or Northern Apennines hosted the largest populations in pre-LGM times, with other important populations occurring on the Balkan Peninsula (Tinner *et al.*, 2013). Mitochondrial DNA variation clearly suggests the presence of two haplotypes corresponding to the Italian and Balkan Peninsulas (Ziegenhagen *et al.*, 2005; Liepelt *et al.*, 2009) (Fig. S3).

The demographic history of silver fir over the past 22 kyrs BP was obtained from the LPX-Bern dynamic global vegetation model with a resolution of 1° by 1° Lat/Lon (Sitch *et al.*, 2003; Ruosch *et al.*, 2016). The model was forced with climate anomalies and included competition between common tree species and plant functional types. The output of LPX-Bern is the Foliar Projective Cover (FPC), which is the fraction of a grid cell that is covered by silver fir. We estimated the number of trees ( $N$ ) in each deme from FPC, assuming that a mature tree occupies 40 m<sup>2</sup>, and that  $N/N_e = 0.001$  (an arbitrary but realistic value (Waples *et al.*, 2011)). The full input data consisted 221 time points spaced at 100 year (i.e. four generation) intervals on a 53 × 24 grid. In the following we shall refer to one grid cell of LPX-Bern as one deme. While the population size of the whole species (i.e. all demes) showed an overall increasing trend with time (post-LGM colonisation), the size fluctuations of individual demes were highly variable (see Fig. 5a). We used the expected coalescence time for two samples taken from the same deme as an approximation for the genetic diversity in a deme, thus assumed that mutations can be neglected. Finally, we note that LPX-Bern has several shortcomings and does not predict the current distribution of silver fir accurately. However, the objective of this example was not to make predictions for the expected levels of genetic diversity in silver fir, but to study the effect of spatio-temporal heterogeneity in population size in a biologically realistic scenario.

385 We performed four simulated scenarios. First, we used a homogeneous deme size (i.e.  $N_e$ )  
386 in space and time, which represented our null model. We fixed the deme size to its average size  
387 based on the last step of the LPX-Bern data. Second, we included the spatial variation in deme  
388 size, represented by the last step of the LPX-Bern data, but kept deme sizes constant in time.  
389 Third, we used the full LPX-Bern input data, thus considering realistic deme sizes changing both  
390 in space and in time. Fourth, we explored the effect of having two ancestral populations, i.e. using  
391 pre-LGM historical information. For this, we used simulations identical to the third scenario,  
392 but at the oldest time point (i.e. 22 kyrs BP), and we combined the demes into one of the two  
393 most plausible ancestral populations based on the spatial distribution of mtDNA haplotypes in  
394 contemporary samples (Fig. ??a). We achieved this by simply assigning each deme with mtDNA  
395 data to the dominant haplotype (i.e. more than 50% Balkan or Italian type) or to the origin of the  
396 nearest deme of known origin, in case of missing data (Fig. ??b).

397 We found that both space and time had an effect on the coalescence times, and thus on the  
398 distribution of genetic diversity in space (Fig. 5b). As expected, when the deme size was constant  
399 in space and time, the distribution of genetic diversity only reflected stochastic effects of the  
400 coalescence process (Fig. 5b, Scenario 1). Spatial variation in deme size introduced variation  
401 in the expected levels of genetic diversity, which was also proportional to the deme size (Fig.  
402 5a and b, Scenario 2). When deme size varied both in space and time, the spatial variation in  
403 the mean coalescence time became even stronger. In particular, the recently colonised areas of  
404 Northern Europe had a lower expected level of genetic diversity (Fig. 5b, Scenario 3). Finally, when  
405 we assumed two ancestral populations, their contact zone had much higher levels of expected  
406 genetic diversity (Fig. 5b, Scenario 4). This is because there was a much longer waiting time for  
407 the two ancestral populations to coalesce, which is determined by the size of these populations  
408 and also by the migration rate between them. For a real data application, calibration of these  
409 two parameters would be necessary to match the observed genetic diversity data. Alternatively,  
410 the match between simulated and observed data could be used to estimate the divergence time

<sup>411</sup> between the two mtDNA haplotypes (e.g. Hickerson *et al.*, 2007).

## 4 Discussion

### The role of spatial and temporal autocorrelation

Using a wide range of simplified and biologically realistic simulations, we have identified several factors that may cause a deviation from theoretical expectations of the island model and the 2D stepping stone model. We found that non-spatial null models, such as the island model, are inappropriate in the presence of spatial autocorrelation in deme size (Fig. 3). Most real-life situations involve some degree of spatial autocorrelation. Previous studies have already demonstrated the limitations of non-spatial null models, for example in the presence of isolation by distance (Wang & Whitlock, 2003; Meirmans, 2012), due to population structure or biased sampling schemes (Chikhi *et al.*, 2010), or to local variation in deme size or barriers to gene flow across the landscape (Duforet-Frebourg & Blum, 2014). Here, we show that the 2D stepping stone model can account for spatial autocorrelation, at least when it is homogeneous across the landscape, and to some extent when there is local variation in deme size (clustered scenario) (Fig. 3). Thus far, the 2D stepping stone model has rarely been used as a null model (but see Duforet-Frebourg & Blum 2014 and Battey *et al.* 2020), partly due to the lack of a simulation tool. *gridCoal* could facilitate more widespread use of the 2D stepping stone model to generate the null distributions of neutral statistics, such as genetic diversity (assuming a non-zero mutation rate) or  $F_{ST}$ , in the presence of spatial autocorrelation in population size.

Demography, or temporal change in population size, is well known to contribute to deviations from theoretical expectations of the island model, and can limit the validity of statistical procedures that are based on this model. This is particularly true for  $F_{ST}$ -outlier tests used to detect loci under selection (*e.g.* Chikhi *et al.*, 2010; Bierne *et al.*, 2013; De Mita *et al.*, 2013; Lotterhos & Whitlock, 2014) because  $F_{ST}$  is dependent on recent ancestry (Slatkin, 1991). Here, we simulated realistic scenarios with the presence of both spatial and temporal heterogeneity in

436 deme size, and observed that deviations from the theoretical expectations are strongest when  
437 the spatial and temporal autocorrelation in deme size are decoupled (Fig. 3c, range expansion  
438 and shift). Our simulations demonstrate that neutral  $F_{ST}$  is well below the theoretical  
439 expectations for such a range expansion and shift (Fig. 3c). This result is in agreement with the  
440 findings of Lotterhos & Whitlock (2014), who showed that spatial autocorrelation in deme size  
441 or recent range expansion resulted in the largest number of false positives for most methods in  
442 efforts to detect spatially divergent selection. Spatio-temporal trends in population size are  
443 expected to be common in nature, especially in the Northern hemisphere, where the  
444 demographic history is often dominated by expansion from glacial refugia and a shift towards  
445 the north (e.g. Excoffier *et al.*, 2009). Our example application also illustrates such a case (Fig. 5).

446  $F_{ST}$  and  $F^*$  are based on the same information, but  $F_{ST}$  is a more integrative and therefore  
447 more robust measure, while  $F^*$  is more sensitive to local differentiation patterns (Fig. 3g–i). Note  
448 that our  $F^*$  is closely related to  $\hat{M}$  of Slatkin (1993), which has the advantage of being independent  
449 of the mutation rates when they are small across loci. Based on a wide range of scenarios, we found  
450 that spatial and temporal variation in deme size can influence the steepness of the isolation-by-  
451 distance curve. In agreement with Duforet-Frebourg & Blum (2014), we found that local variation  
452 in population size, as in our clustered map, caused large variance in local  $F^*$  (Fig. 3g). The most  
453 complex range expansion and shift scenario led to a relatively flat isolation-by-distance curve  
454 (Fig. 3i). Indeed, Slatkin (1993) already proposed that the lack of an isolation-by-distance pattern  
455 in a natural population can indicate non-equilibrium populations or recent colonisation, a pattern  
456 that has been confirmed through empirical studies (e.g. Leblois *et al.*, 2000; De Kort *et al.*, 2014).

## 457 ***gridCoal*: Guidelines for future users**

458 *gridCoal* is a wrapper for the most efficient algorithm to simulate genealogies: the optimised  
459 continuous time approximation of the coalescence process implemented in *msprime* (Kelleher  
460 *et al.*, 2016). It complements the existing arsenal of spatially explicit simulators (Guillaume &

461 Rougemont, 2006; Landguth & Cushman, 2010; Haller & Messer, 2019; Currat *et al.*, 2019; Dellicour  
462 *et al.*, 2014; Becheler *et al.*, 2019). The choice of parameters and model calibration are essential for  
463 running spatially coalescent simulations. Here we provide some guidelines in the case of *gridCoal*.

464 The spatially explicit phase (given by the number of steps  $T$  and the time step  $dt$ ) should be  
465 long enough so that lineages coalesce during this phase, but also short enough to avoid wasting  
466 computational time. The choice of  $dt$  should be driven by the particular biological question. For  
467 example, throughout this paper we used a combination of parameter values (number of steps  $T$ ,  
468 time step  $dt$  and generation time  $gt$ ) such that most lineages coalesced in the spatially explicit  
469 phase across all combinations of  $N$  and  $m$  (Fig. 4d–f). Note that the largest  $dt$  was necessary for  
470 intermediate values of  $Nm$ , where lineages can escape and take a long time to coalesce. We  
471 suggest that users perform test simulations with the required values of  $N$  and  $m$  to choose an  
472 appropriate  $dt$ . This is particularly important if it is necessary that all lineages coalesce during  
473 the spatially explicit phase, e.g. for estimating genetic diversity maps such as those shown in the  
474 example of post-glacial colonisation of silver fir (Fig. 5). In contrast, if the question concerns a  
475 particular organism with a given generation time and across a particular time period, the  
476 parameters can be chosen accordingly. For example, setting  $dt = 100$  and using 210 time points  
477 takes the ancestral population back to the Last Glacial Maximum (LGM, 21 kya), which could be  
478 a suitable parameter combination for several species that expanded after the LGM.

479 *gridCoal* avoids the simulation of genetic data and instead simulates summary statistics that  
480 can be derived from coalescence times, i.e. gene diversity, the strength of population structure  
481 ( $F_{ST}$ ), and the genetic distance between pairs of demes ( $F^*$ ). We emphasise that approximating  
482 summary statistics of genetic diversity and  $F_{ST}$  from coalescence times holds only when the  
483 mutation rate is low and when migration is possible to neighbouring demes only (Slatkin, 1985).  
484 Further, for comparing *gridCoal* simulations to real data, a calibration of  $N_e$  and  $\mu$  is necessary  
485 because these parameters are non-identifiable. Such a calibration can be achieved by using  
486 additional information about the mutation rate of particular genetic markers used and by

487 estimating  $N/N_e$  (Waples *et al.*, 2011). Finally, simulations from *gridCoal* are closer to that of a  
488 continuous space model, and thus to biological reality, for large grid sizes. Nevertheless, at least  
489 for small neighbourhood sizes, a grid of  $50 \times 50$  already appears to be sufficient to accurately  
490 approximate a continuous space process for many commonly used summary statistics (see  
491 details in Battey *et al.* 2020).

### 492 ***gridCoal* for eco-evolutionary data fusion**

493 *gridCoal* might be useful for empirical applications of eco-evolutionary data-fusion approaches,  
494 such as integrative Distributional Demographic Coalescent (iDDC) approach (He *et al.*, 2013;  
495 Brown & Knowles, 2012). In this context, one key feature of *gridCoal* is that it is not only  
496 spatially but also temporally explicit. Temporal explicitness means that the exact population size  
497 of each deme has to be set by the user at regularly placed time intervals. In this way, *gridCoal* is  
498 fully deterministic in terms of the forward-time demography, and stochastic in terms of the  
499 backward coalescence events. Although this feature may appear as a limitation in some  
500 situations, it is necessary for applications that make use of species distribution data issued from  
501 ecological models and paleological data (Svenning *et al.*, 2011; Gavin *et al.*, 2014). This feature  
502 also represents an important contrast to *SPLATCHE 3* (Currat *et al.*, 2019), where each deme  
503 follows a logistic growth model. As a result, in *SPLATCHE 3*, user-provided population sizes are  
504 only approximately achieved, no population declines, and only local extinctions are possible.  
505 Indeed, to set up explicit temporal changes in population size, Ortego & Knowles (2020) updated  
506 the population sizes only three times from 21 ky BP to the present, which is a rough  
507 approximation of actual population size changes and may bias the isolation-by-distance patterns  
508 (Fig. 4d).

509 There is a wide range of possible input data sets that can be used for eco-evolutionary data-  
510 fusion approaches. First, paleo-climatic databases have opened possibilities for running species  
511 distribution models (SDMs; Elith & Leathwick 2009; Sexton *et al.* 2009) and for producing a time

512 series of species distribution maps. Second, process-based dynamic vegetation models (DVMs;  
513 Pereira *et al.* 2010) offer important advantages over SDMs, which are static, correlative approaches,  
514 and thus DVMs hold a great potential for use in data-fusion approaches. Even though DVMs suffer  
515 from limitations related to their complex parametrisation, they are continually improving as the  
516 quality and richness of climatic, remote sensing, and other biological data increases (*e.g.* Hartig  
517 *et al.*, 2012). Third, fossil data is increasingly being organised in databases (Davis *et al.*, 2013; Peters  
518 *et al.*, 2019). The most abundant type of fossil data is pollen, particularly from forest trees, which  
519 has been used to reconstruct past population size fluctuations (*e.g.* Ruosch *et al.*, 2016; Kaufman  
520 *et al.*, 2020). Indeed, our example of post-glacial colonisation history in silver fir (see Fig. 5) could  
521 be made more realistic by using the spatio-temporal interpolation of pollen records (Ruosch *et al.*,  
522 2016).

## 523 **Data accessibility**

524 Detailed information about the software, a user manual, and example simulations are available  
525 for download at <https://github.com/Trubenova/gridCoal>.

## 526 **Author contributions**

527 KC designed the research. ES and BT developed *gridCoal*, ran the simulations and analysed the  
528 data. ES, BT and KC wrote the manuscript.

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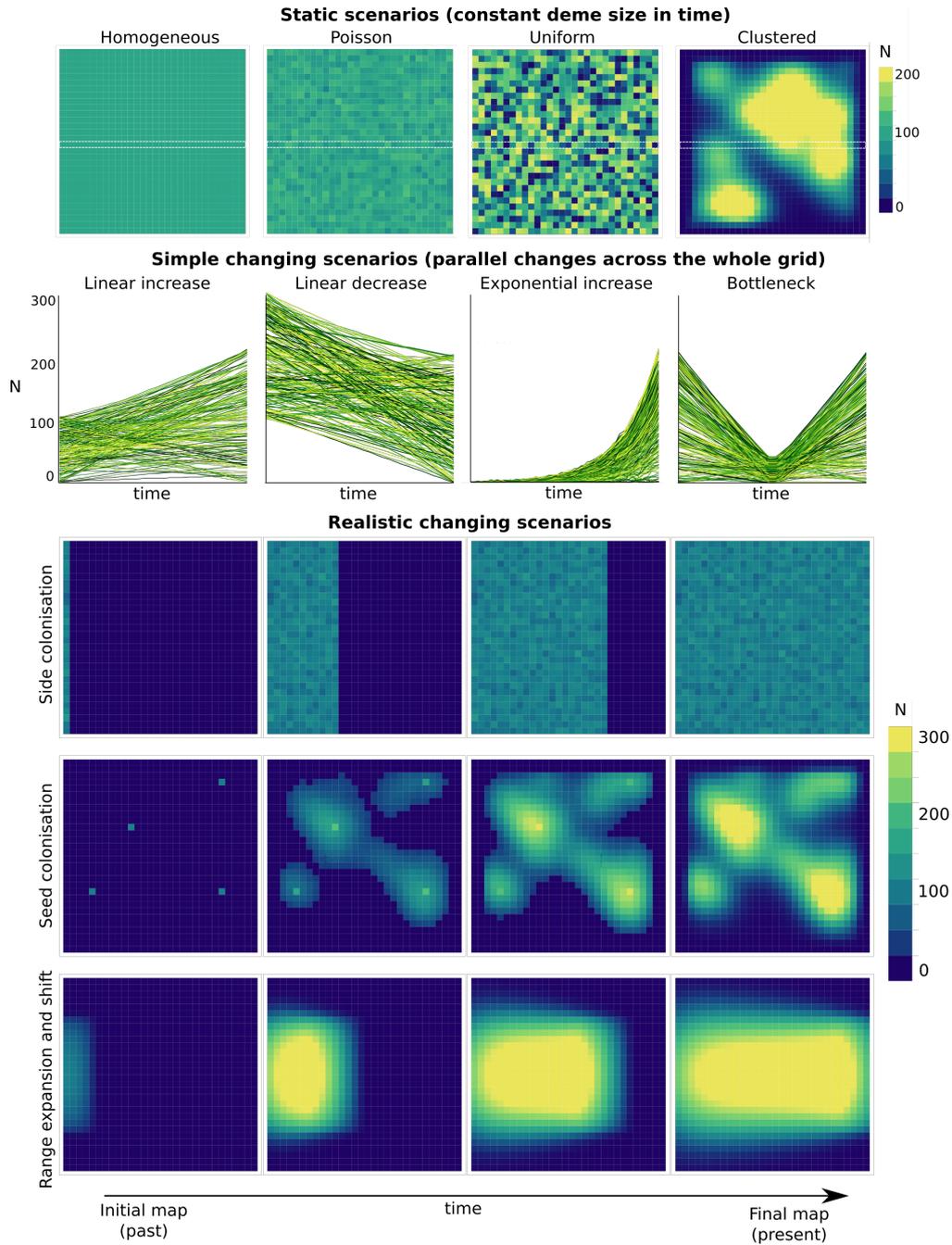
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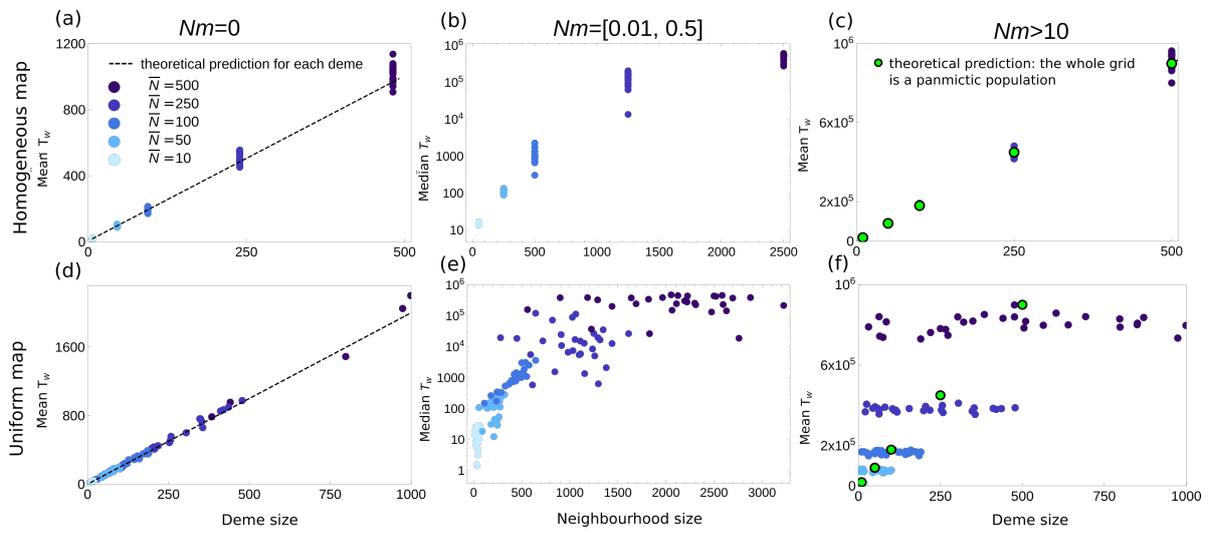
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**Table 1:** Symbols and terms and their definitions.

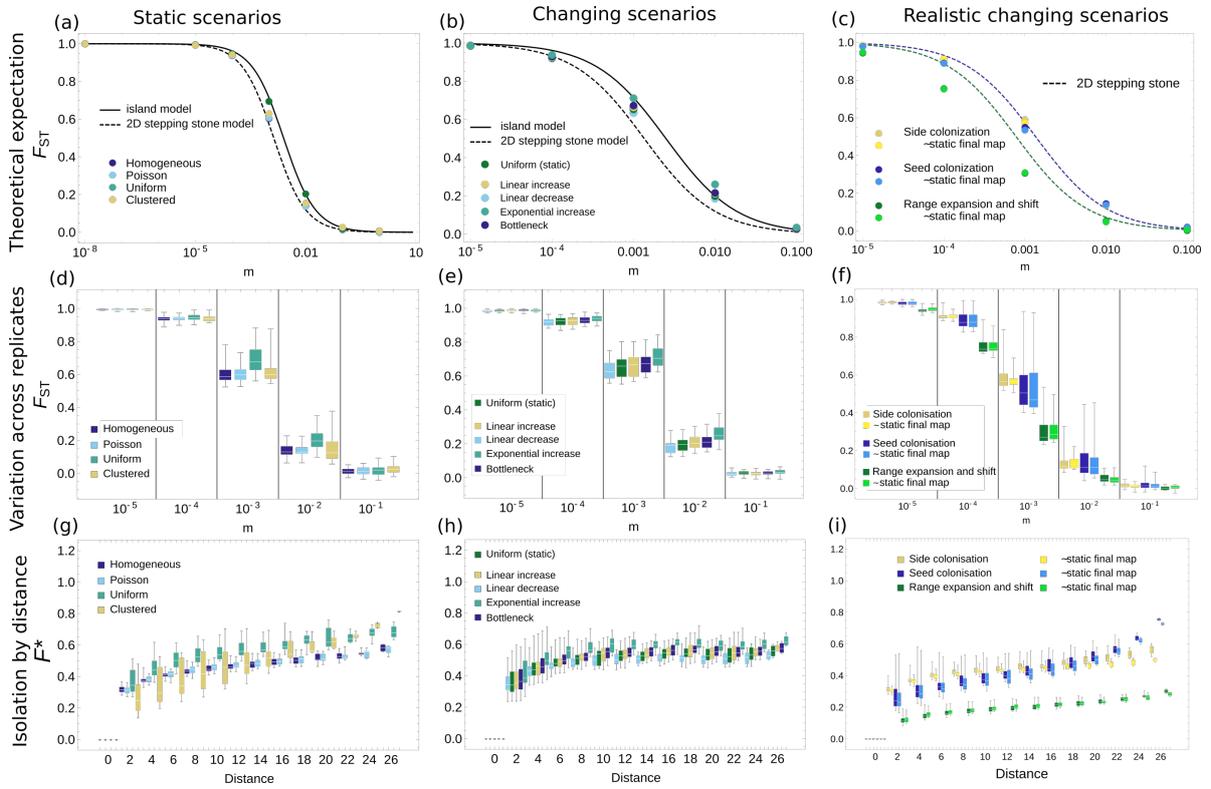
Symbol	Term	Definition
$d$	deme	panmictic population in a single grid cell
	map	grid with a defined distribution of deme sizes
$L$	grid size	number of rows (columns) in a square grid
$N$	deme size	number of individuals in a deme
$N_b$	neighbourhood size	size of a focal deme and its four neighbours
$T$	number of time points	number of time points with the defined demographic history
$gt$	generation time	the interval between the birth of an individual and the birth of its offspring
$dt$	time step	time between two defined time points, in years
$m$	migration rate	fraction of population moving from the ancestral cell to a neighbouring cell
$T_W$	within-deme coalescence time	coalescence time between two lineages drawn from the same deme
$T_B$	between-deme coalescence time	coalescence time between two lineages drawn from different demes
$T_T$	average coalescence time	average coalescence time of any two lineages drawn from the grid



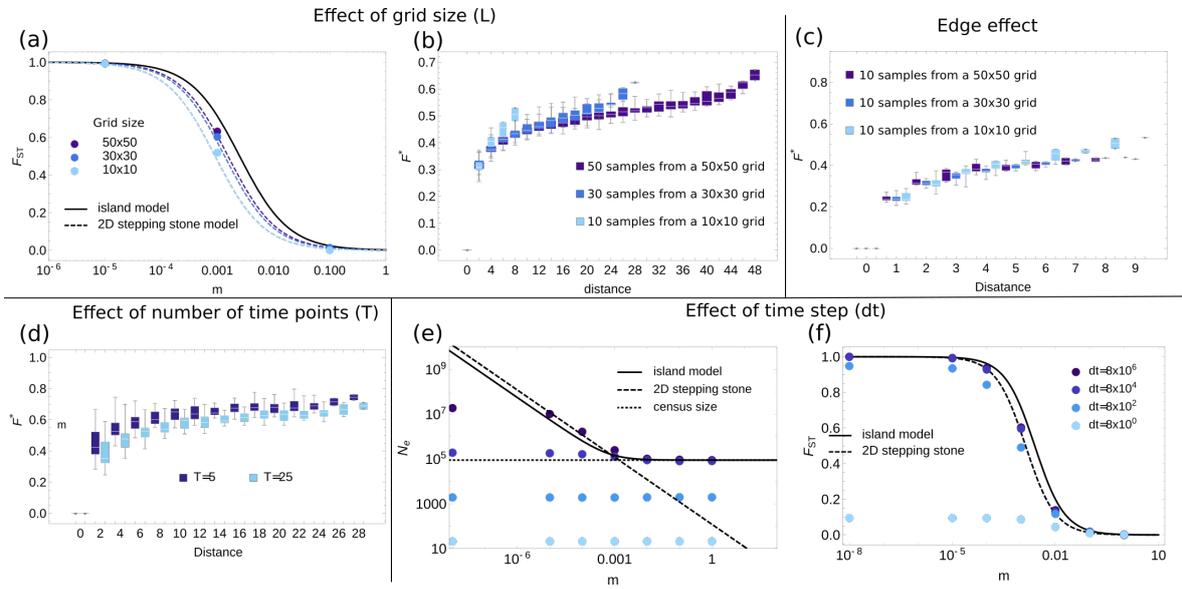
**Figure 1:** The three different groups of scenarios simulated. **Static scenarios:** demes had a constant size across the spatially explicit phase of the simulations. **Simple changing scenarios:** the size of all demes changed in a correlated manner. In the present time step, all scenarios were identical to the deme sizes drawn from a uniform distribution. **Realistic changing scenarios:** deme sizes changed in space and time to model a colonisation event. The grid size was 30 across all scenarios. To estimate  $T_W$  two lineages were sampled in each deme, and to estimate  $T_B$  two lineages were sampled from 30 demes in a row in the middle of the grid.



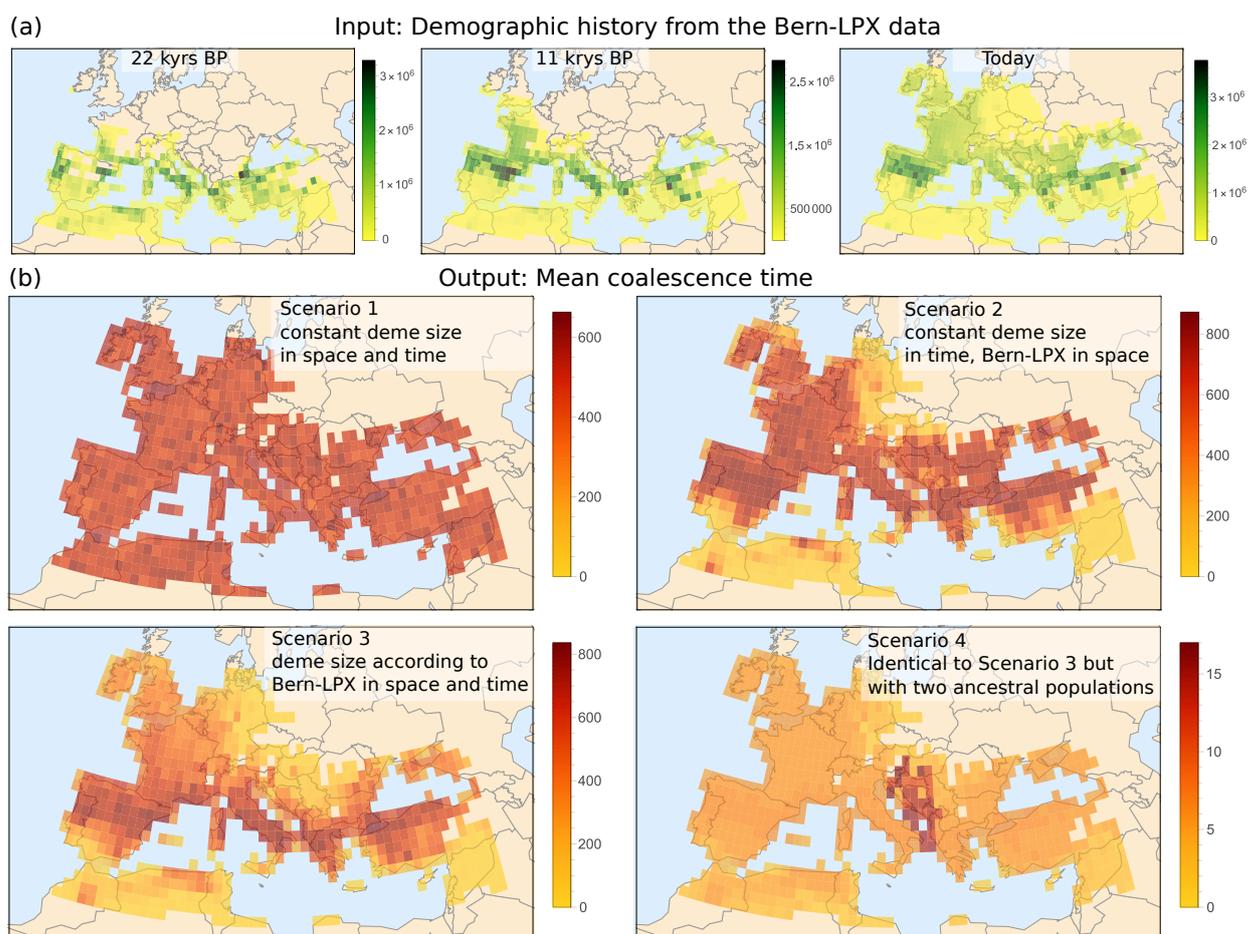
**Figure 2:** Within-deme coalescence times ( $T_W$ ) for different ranges of  $Nm$  from zero (a, d) to  $m = 1$  (panmictic grid) (c, f) across two maps: homogeneous (a, b, c) and uniform (d, e, f). The theoretical predictions are for a Wright-Fisher model with  $N_i$ , where  $i$  is an index for demes (a, d) and for  $2L^2\bar{N}$ , where  $L$  is the grid size and  $\bar{N}$  is the average deme size across the grid. There is no theoretical prediction for intermediate  $Nm$  (b, e) because for those parameter ranges the median  $T_W$  is shown. Each parameter combination ( $N$  and  $m$ ) is represented by 30 dots showing  $T_W$  for individual demes.



**Figure 3:** Comparison of  $F_{ST}$  from different scenarios with theoretical predictions of the island model and the 2D stepping stone model (a, b, c). Variation in  $F_{ST}$  across 1 000 replicate simulations of the same map (d, e, f). Isolation-by-distance patterns characterised as  $F^*$  against distance for different scenarios (g, h, i). Simulation parameters:  $L = 30$ ,  $T = 30$ ,  $dt = 50000$ ,  $gt = 25$ .

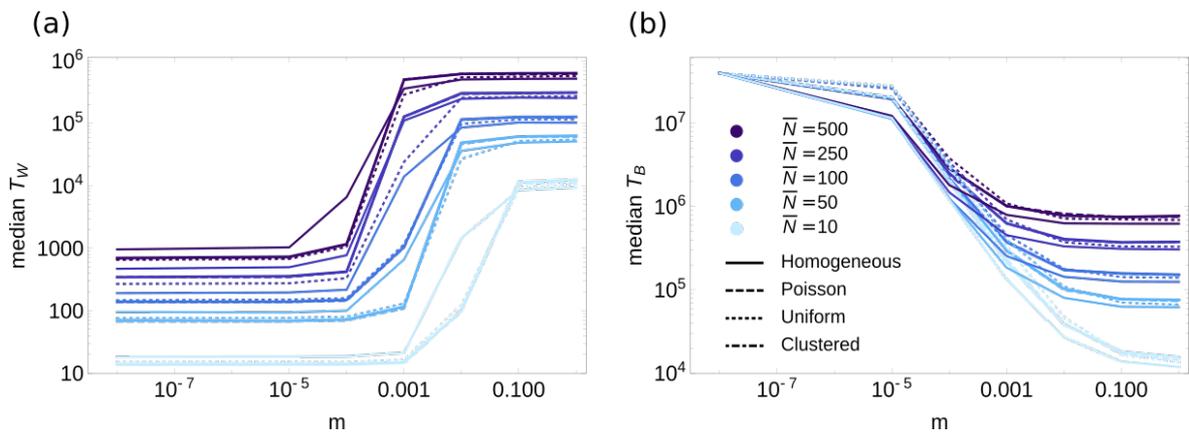


**Figure 4:** Sensitivity of  $F_{ST}$  and  $F^*$  to  $L$  (a, b),  $T$  (d), and  $N_e$  (estimated as  $T_T/2$ , where  $T_T$  is the average coalescence time) and  $F_{ST}$  to  $dt$  (e, f) using a homogeneous map,  $\bar{N} = 100$ , and  $m = 10^3$ . Unless otherwise specified,  $dt = 2 \times 10^8$ ,  $gt = 25$ , and  $T = 5$ . The edge effect (c) was explored using inner demes sampled along a line in the middle of the grid.

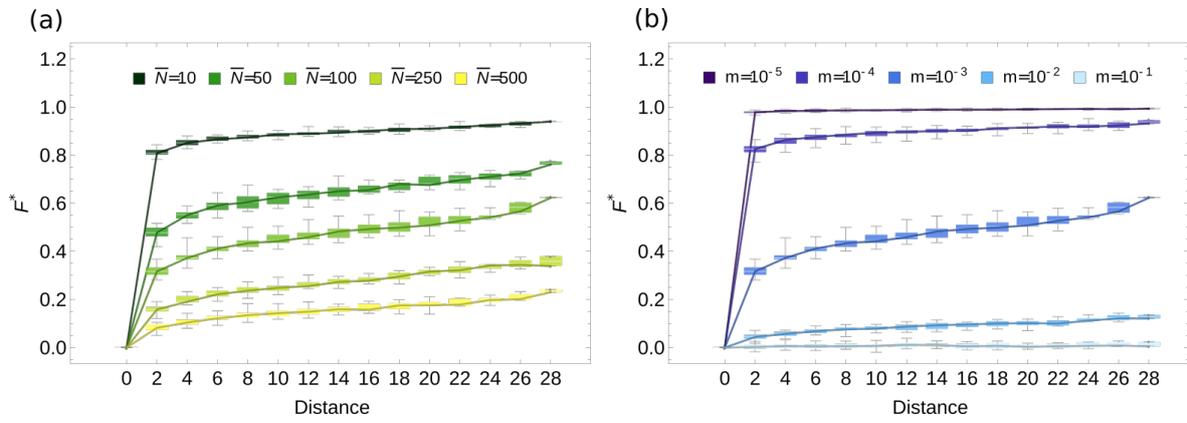


**Figure 5:** Real-world example: range expansion of silver fir (*Abies alba* Mill.) since the Last Glacial Maximum (LGM, 22 kyrs BP). **(a)** Raw input data for *gridCoal*: the demographic history from the global dynamic vegetation model LPX-Bern. Three time points are shown out of the 220: the LGM, the beginning of the Holocene, and today. **(b)** Mean coalescence time from the simulated scenarios with increasing complexity in terms of spatio-temporal variation in deme size from the top left to the bottom right panel. Note that the colour scale differs between maps.

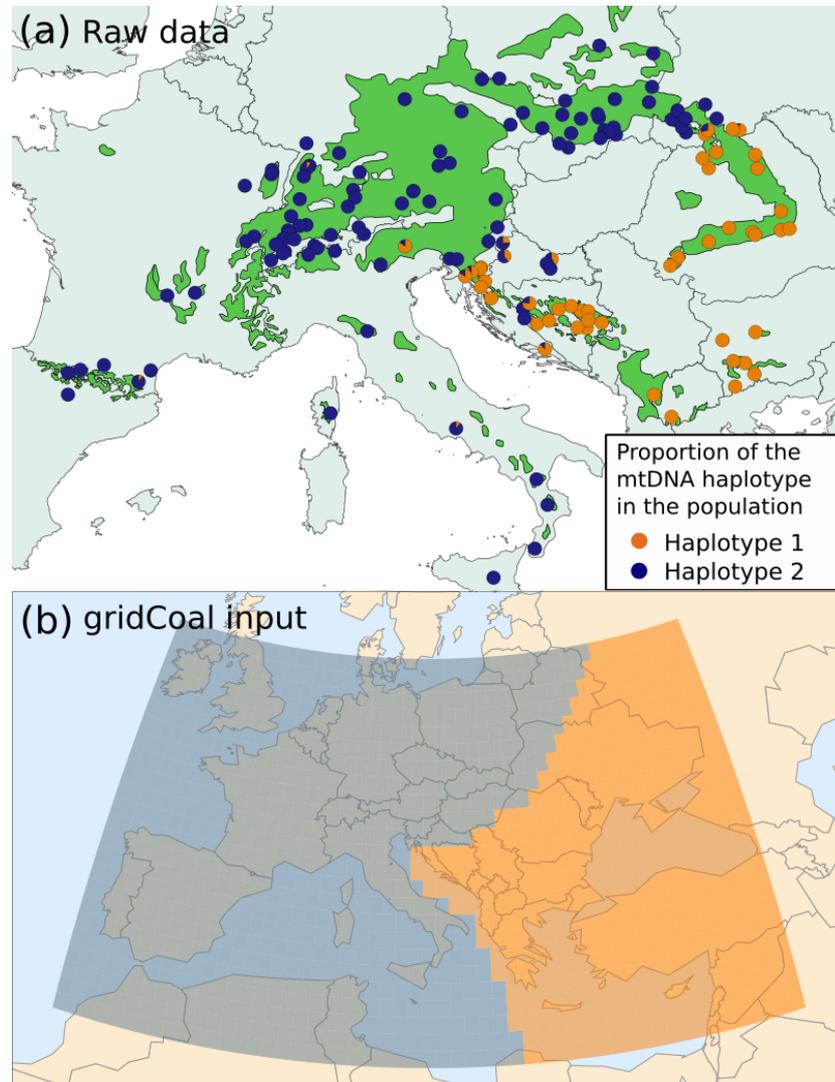
711 **Supplementary figures**



**Figure S1:** Median coalescence time of two lineages from the same deme  $T_W$  (a) and from different demes  $T_B$  (b). Different line types represent different maps (see Fig. 1 for details).



**Figure S2:** Isolation-by-distance patterns for different average deme sizes,  $\bar{N}$  (a), and migration rates,  $m$  (b).



**Figure S3:** (a) The distribution of silver fir (*Abies alba* Mill.) mtDNA haplotypes in present day samples following (Liepelt *et al.*, 2009) (pie charts) and the distribution of silver fir (in green) from EUFORGEN (<http://www.euforgen.org>). (b) mtDNA data used to assign each grid cell to one of the two ancestral populations in Scenario 4.

## 712 **A *gridCoal* description and user manual**

### 713 **A.1 Input files and parameters**

714 In order to run the simulations, it is necessary to define the following input files and parameters.

715 **Demographic history input files [required]:** The demographic history of the collection of  
716 demes distributed on a grid is represented by a matrix of size  $T \times n$ , where  $T$  is the number of  
717 time points at which one wishes to define the population sizes and  $n$  is the number of grid cells.  
718 The matrix contains the population sizes of the grid cells at given time points. Each row is the  
719 flattened two dimensional grid, indexed from 0 to  $n - 1$ , defining the sizes of the subpopulations.  
720 The first line is the oldest time point. In *msprime*, a population is not allowed to have size 0. In  
721 our case, however, we do not want to exclude the possibility that populations become extinct and  
722 the demes are subsequently recolonised, even repeatedly. We therefore set populations with size  
723 0 as  $10^{-10}$ . This is done automatically – before the simulations start, the program replaces any 0  
724 in the input data with  $10^{-10}$ .

725 *Example:* demographic history with three data points

726

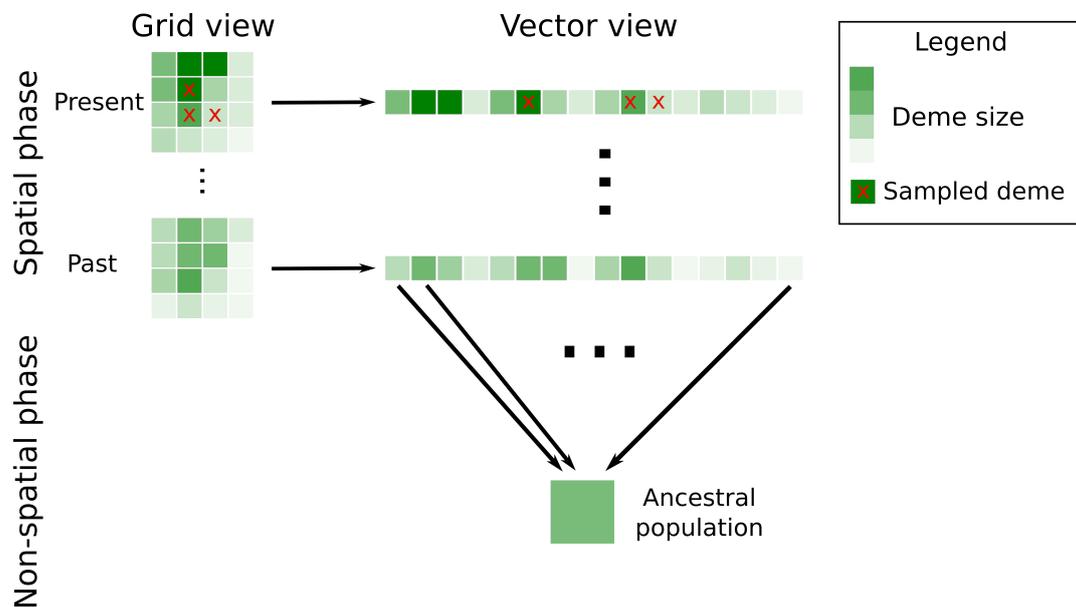
1	2	0	4	2	3	4	5	3	4	5	5	4	5	6	7
1	1	1	1	2	2	2	2	3	0	0	3	4	4	4	4
1	2	3	4	2	3	4	5	3	4	5	0	4	5	6	7

728

729 *Enter as:* `-pop InputData.txt` or `--pop_sizes InputData.txt`

730 **Row number [required]:** This number, together with the demographic history file, defines  
731 the shape of the spatial map. It must be an integer. Additionally, the size of the grid (number of  
732 all cells – row length of the demographic history input file) must be divisible by the row number.

733 *Enter as:* `-row 5` or `--row_number 5`



**Figure A1:** Input data preparation.

734 **List or a fraction of sampled cells [optional]:** A list of cells from which the samples are  
 735 taken is can be supplied. <sup>1</sup>. These cells must not be empty at the final time point (present), but  
 736 could be empty in the past. For efficiency, two samples are taken from each sampled deme. <sup>2</sup> If  
 737 only a number (float, < 1) is supplied, random, non-empty cells will be sampled, with the number  
 738 representing the sampled fraction. If no file is supplied, all samples that are not empty at present  
 739 are sampled.

740 *Example: SampleList.txt*

741 0 1 2 3 4 7 8 33 34 35

742 *Enter as: -sam SampleList.txt or --sample\_coords SampleList.txt*

<sup>1</sup>Mind the indexing of the sampled cells, which also must start from 0.

<sup>2</sup>It is more efficient to run more replicates with fewer samples than fewer replicates with more samples. In the coalescence process, the waiting time until the next coalescence event happens is exponentially distributed, with its mean proportional to the number of lineages. Thus, in the beginning, several coalescence events happen in quick succession, yet the mean coalescence time of the deme is largely dominated by the amount of time that the last remaining few samples took to coalesce.

743 **Migration matrix** A two-dimensional migration matrix capturing backward migration is  
 744 needed to run the simulation itself; however, as an input file we only need the forward  
 745 migration matrix,  $M$ <sup>3</sup>. We assume that  $M$  remains constant in time because it depends on the  
 746 dispersal ability of the species. The element  $(M_{i,j})$  defines the fraction of the lineages in  
 747 population  $i$  that migrates to population  $j$ . In order to run coalescence simulations, *gridCoal*  
 748 calculates a backward migration matrix,  $BM$ .  $BM$  changes through time, as it depends on the  
 749 actual population sizes of the neighbouring cells.  $BM_{i,j}(t)$  defines the fraction of lineages in  
 750 population  $i$  at time  $t$  that have parents in population  $j$  and is thus calculated as:

$$BM_{i,j} = \frac{M_{j,i}N_j(t-1)}{N_i(t)} \quad (\text{A1})$$

751 The fact that the population size changes and migration matrices are not updated at every  
 752 generation, but at arbitrary time steps, represents a significant gain in computing time in  
 753 comparison to other tools such as *SPLATCHE 3*.

754 In the newer version of *gridCoal*, only a migration list is needed, capturing the migration rate  
 755 from source to target cell.

756 A migration list indicating the migration rate between two connected cells is used to build a  
 757 migration matrix, and to calculate backward migration during the simulation. Even if two cells are  
 758 connected by an edge, if the migration is not specified in the list, it is considered 0. The migration  
 759 list must be formatted as lines of three values: source cell  $i$  (integer), target cell  $j$  (integer), and  
 760 migration rate  $m_{ij}$  (positive float between 0 and 1). Note that if migration involves the exchange  
 761 of migrants, both directions need to be specified.

762 If a file is not specified but rather a single number  $m$  (float) is supplied, a migration matrix  
 763 is generated in which migration is assumed to occur between adjacent cells symmetrically with  
 764 rate  $m$ .

765 *Example: MigrationList.txt:*

---

<sup>3</sup>The diagonal elements of  $M$  must be zero, a requirement for *msprime*.

766 0 1 1.00e-06  
767 0 5 1.00e-06  
768 1 0 1.00e-06  
769 1 2 1.00e-06  
770 1 6 1.00e-06  
771 2 1 1.00e-06  
772 2 3 1.00e-06  
773 2 7 1.00e-06  
774 3 2 1.00e-06

775

776 Enter as: `-mig MigrationList.txt` or `-mig 0.000001`

777 or `--migration_matrix MigrationList.txt`

778 or `--migration_matrix 0.000001`

779 **Time step and generation time [optional]:** The amount of time between two time points,  
780 denoted  $dt$ , is given in arbitrary time units (years, months, days, minutes).

781 Time is measured in generations in *msprime*, and we therefore need to specify the generation  
782 time of the population at hand. We define the generation time,  $dt$ , as the time it takes for a species  
783 to reach a reproductive age, expressed in the same units as other supplied times. The timing  
784 of demographic events (expressed in the same units) is re-calculated by dividing the time point  
785 of events specified in demography input files by the generation time of the simulated organism  
786 expressed in the same units. Therefore, it is possible to run the simulations for any organism with  
787 an arbitrary generation time, from bacterial populations to trees.

788 By default, generation time is set to 1, and the time between two supplied data points is set at  
789 10.

790 Enter as: `-dt 100 -gen 25` or `--delta_t 100 --generation_time 25`

791 **Ancestral populations** At the point in time beyond which the demography is unknown, all  
792 lineages are merged into spatially non-explicit ancestral populations where they follow the  
793 standard coalescence process. We assume either a single or multiple panmictic ancestral  
794 populations with specified sizes and a very low rate of migration between them ( $10^{-8}$ ).  
795 Furthermore, it is necessary to specify which of the cells originate in each ancestral population.

796 A list determining the origin of each cell can be supplied as a txt file with n (number of all  
797 cells) lines. If no file is supplied, all cells are expected to originate in a single spatially non-explicit  
798 population. The size of the ancestral populations can be set, with the default as 1.

799 **Other inputs** It is further possible to specify an **output directory**, into which outputs (log file  
800 with input parameters, demography debugger, random seed numbers and coalescence times) are  
801 saved. The default value is OUTPUT.

802 *Enter as: --output\_dir 7 or -odir 7*

803 **Print demography file** This option can be used to print a detailed demography debugger  
804 file, supplied by *msprime*. The replicate number must be set to 1. This makes it possible to  
805 simultaneously run many simulations with only one debugger file (identical for all).

806 *Enter as: -pdeb BOOL or --print\_debugger BOOL*

807 Finally, it is possible to set a **random seed number**, which makes it possible to reproduce a  
808 given simulation.

809 *Enter as: --set\_seed INT or -seed INT*

## 810 **A.2 Demographic events**

811 All demographic changes, including population size and migration rate changes, need to be  
812 defined as a demographic event at a given time point, going backwards in time, and collected to  
813 a list, which is used by *msprime* for the coalescence simulation. This is done automatically by  
814 *gridCoal* based on the file containing the demographic history.

815 At each generation, we calculate the backward migration as described above.

816 Similarly, we update the population sizes, with one additional constraint. If a deme has  
817 individuals at a given time point  $t$  but was empty in the preceding time point  $t - 1$ , we need to  
818 define the source of those individuals. Specifically, we define a mass migration event from this  
819 cell to its 4 neighbouring cells, at rates proportional to their population sizes in the preceding  
820 generation. Thinking forward in time, this corresponds to the idea of an empty cell being  
821 colonised by its neighbours, at rates proportional to their population sizes.

822 After a list of demographic events is created, these are supplied to *msprime*, together with the  
823 initial conditions. Single repeats are run in each simulation. The coalescence tree is extracted,  
824 and the time of coalescence of each pair of samples is saved in matrix format, with sampled cells  
825 in rows and columns. The matrix is symmetrical, with the diagonal representing the within-deme  
826 coalescence time.

### 827 **A.3 Output files**

828 **Summary of inputs** For `REPLICATE = 1`, All the input files are collected and saved into a  
829 created `OUTPUT_DIR_NAME` directory as  
830 `OUTPUT_DIR_NAME/Output.txt`. The random seed number is saved in the same file.

831 **Print demography debugger** If `--print_debugger` is set to `True`, the detailed  
832 demographic history – with all changes in population size and migration rate – is produced by  
833 *msprime* and printed into the `OUTPUT_DIR_NAME` directory as  
834 `OUTPUT_DIR_NAME + 'DemographyDebugger.txt'`. Note that this output file can  
835 be very large.

836 **Simulation results** The result of the simulation itself is a square matrix of coalescence times  
837 of samples from all sampled demes, saved into `OUTPUT_DIR_NAME` directory as

```
838 OUTPUT_DIR_NAME + 'CoalTimes' + REPLICATE + '.txt'
```

## 839 B Simulated scenarios

### 840 B.1 Static scenarios

841 In these simulations, all demes had constant sizes across  $T = 5$  time points and  $dt = 2 \times 10^8$   
 842 years ( $8 \times 10^6$  generations, 25 years per generation), on a square grid of size  $L = 30$ . Different  
 843 scenarios (maps) were simulated, with increasing spatial variance (see Fig. 1):

- 844 • *homogeneous (no spatial variance)*: equal-sized demes of size  $N$
- 845 • *Poisson (low spatial variance)*: deme sizes drawn from a Poisson distribution with mean  $N$
- 846 • *uniform (high spatial variance)*: deme sizes drawn from a uniform distribution with a range  
 847 from 0 to  $2N$
- 848 • *clustered (spatially autocorrelated)*: randomly placed seeds grown into clusters, where  
 849 neighbouring deme sizes were correlated and the average deme size across the whole map  
 850 was  $N$

**Table B1:** Summary of parameter values used in the simulations of static populations.

Variable	Symbol	Values
map type		homogeneous, Poisson, uniform, clustered
average deme size size	$N$	10, 50, 100, 250, 500
migration rate	$m$	0, $10^{-8}$ , $10^{-5}$ , $10^{-3}$ , $10^{-2}$ , $10^{-1}$ , $10^0$

851 Additionally, to address the effects of grid size and edges due to the finite grid size, we  
 852 simulated the homogeneous maps on grids with  $L \in (10, 30, 50)$  sampling 10, 30 or 50 cells in a  
 853 line, for only three migration rates,  $m \in (10^{-5}, 10^{-3}, 10^{-1})$ . To analyse the effect of time  
 854 resolution and the overall simulation time, we ran simulations on a homogeneous map with  
 855 deme size  $N = 100$  on a square grid of size  $L = 30$  with a time step of  
 856  $dt \in (2 \times 10^2, 2 \times 10^4, 2 \times 10^6, 2 \times 10^8)$  years (generation time was assumed to be 25 years),  
 857 with migration rates  $m \in (0, 10^{-8}, 10^{-5}, 10^{-3}, 10^{-2}, 10^{-1}, 10^0)$ .

## 858 B.2 Simple changing scenarios

859 In these simulations, all demes had a changing but similar demographic history (Fig. 1). Maps  
860 were updated at each time point and all scenarios had the same uniform map at the last (most  
861 recent) time point  $U_{final} = \mathcal{U}(0, 2N)$ , where  $N = 100$ , which ensured a valid comparison across  
862 scenarios. Since the deme sizes were changing, we used a finer resolution of  $T = 30$  time points  
863 and a time step of  $dt = 50\,000$  years (2000 generations, 25 years per generation). The following  
864 scenarios were simulated:

- 865 • *Stable*: there was no change in the average deme size across the map, but we started with a  
866 uniform map  $U_{startStable} = \mathcal{U}(0, 2N)$  that differed from  $U_{final}$ . Therefore, individual deme  
867 sizes changed linearly between the initial and final time points.
- 868 • *Linear increase*: individual deme sizes changed linearly from a uniform map, with deme  
869 sizes drawn from  $U_{startLinE} = \mathcal{U}(0, N)$  to  $U_{final}$ . Note that while the average deme size  
870 increased, some individual demes shrank.
- 871 • *Decline*: individual demes changed linearly from a uniform map, with deme sizes drawn  
872 from  $U_{startDec} = \mathcal{U}(2N, 3N)$  to  $U_{final}$ . Note that while the overall population was declining,  
873 some demes became larger.
- 874 • *Exponential increase*: individual deme sizes increased exponentially from their original sizes,  
875 drawn from a uniform distribution  $U_{startExpE} = \mathcal{U}(0, 2N)$ , to their final sizes  $U_{final}$ . Note  
876 that while the whole population was expanding, a few individual demes became smaller.
- 877 • *Bottleneck*: deme sizes first linearly declined from  $U_{startBot} = \mathcal{U}(0, 2N)$  to  
878  $U_{midBot} = \mathcal{U}(0, 0.4N)$ , then expanded to  $U_{final}$ . Note that while the mean deme size  
879 changed from  $N$  to  $0.2N$  to  $N$  again, some individual demes experienced different  
880 demographic histories.

881 Furthermore, we ran a set of simulations where, instead of the uniform map, we used a  
882 spatially autocorrelated map (clustered). We ran similar demographic histories, with no change

883 in time, bottleneck, decline or expansion. We used the same parameters of  $m$ ,  $T$  and step time as  
 884 used for the homogeneous map.

**Table B2:** Summary of parameter values used in the simulations of simple demographic histories.

Variable	Symbol	Values
map type		high-variance (uniform)
average deme size	$N$	100
migration rate	$m$	$10^{-5}, 10^{-4}, 10^{-3}, 10^{-2}, 10^0$
duration of known demographic history	$T$	5, 20

### 885 **B.3 Realistic changing scenarios**

886 These scenarios were inspired by a species range expansion (Fig. 1). Populations colonised the  
 887 grid from different places so that demes had different demographic histories. As a result, the  
 888 temporal and spatial autocorrelation were decoupled to some extent. We used the finer resolution  
 889 of  $T = 30$  time points and  $dt = 50\,000$  years (2000 generations, 25 years per generation). The  
 890 average deme size at the last step was  $N = 100$ . The following scenarios were simulated:

- 891 • *Side colonisation:* demes were colonised from one side of the grid by one column in each of  
 892 the 30 time points. Once a deme was colonised, its size remained constant.
- 893 • *Seed colonisation:* clusters of populations grew from a small number of initially occupied  
 894 demes or "seeds".
- 895 • *Range expansion and shift:* a single expanding population travelled across the grid, while  
 896 colonising its surroundings. At the first time point, only a kernel of  $20 \times 10$  demes were  
 897 occupied. During the following time steps, the kernel demes colonised their neighbours and  
 898 their size increased by 20% at the next time points.

## 899 C Computational run-time of *gridCoal* simulations

900 We performed 17 500 simulations of *gridCoal* for various parameter sets, to illustrate the  
901 dependence of the computational run-time on individual parameters. All simulations were  
performed on a personal computer with the following specifications:

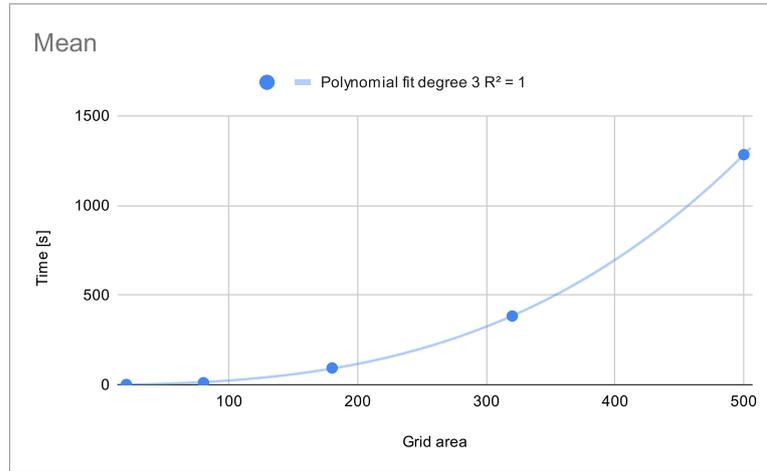
<b>Operating system:</b>	Mac OS Catalina version 10.15.7
<b>Processor:</b>	2.6 GHz 6-Core Intel Core i7
<b>Memory:</b>	16 GB 2667 MHz DDR4

902

903 For each parameter set, 5 batches of 100 simulations (run in series) were run and timed, to  
904 account for random effects (e.g. CPU usage by other programs). In the tables below, individual  
905 batch measurements are shown and run-time trends are displayed in graphs as a mean and  
906 standard deviation of five batches against a changing parameter (in bold). Lines highlighted in  
907 green indicate the simulation batch for the same set of parameters. Time was measured using  
908 the "time" shell command, with "user" time (measured in seconds) shown in the tables.

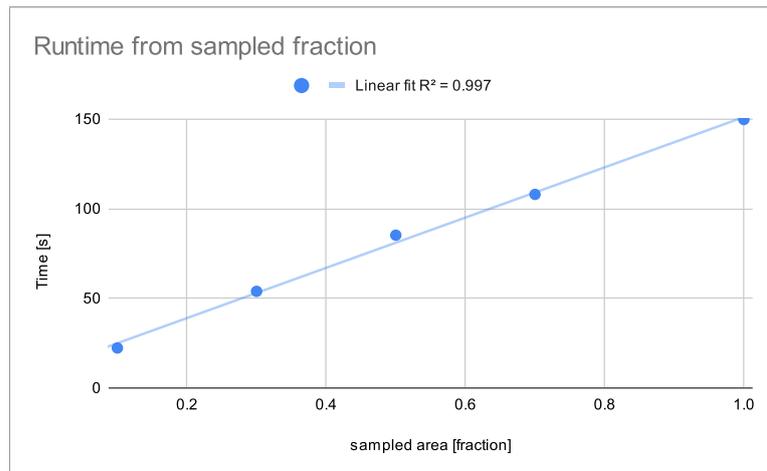
### Effect of grid size

909	Output Directory	Seed	Dimensions	Mean population size	Time points	Time step	Generation time	Migration rate	Sampling fraction	Run 1	Run 2	Run 3	Run 4	Run 5	Mean	Standar deviation	Area
	GridSize1	1	5x4	100	10	200	5	0.1	0.5	1.233	1.207	1.197	1.177	1.208	<b>1.2044</b>	0.0203	20
	GridSize2	1	10x8	100	10	200	5	0.1	0.5	11.108	11.023	10.98	11.099	11.078	<b>11.0576</b>	0.0545	80
	Baseline	1	15x12	100	10	200	5	0.1	0.5	92.962	93.677	93.532	93.095	93.2	<b>93.2932</b>	0.3008	180
	GridSize4	1	20x16	100	10	200	5	0.1	0.5	385.675	381.328	382.483	384.137	381.438	<b>383.0122</b>	1.8675	320
	GridSize5	1	25x20	100	10	200	5	0.1	0.5	1289.598	1280.77	1282.215	1279.916	1281.191	<b>1282.738</b>	3.9231	500



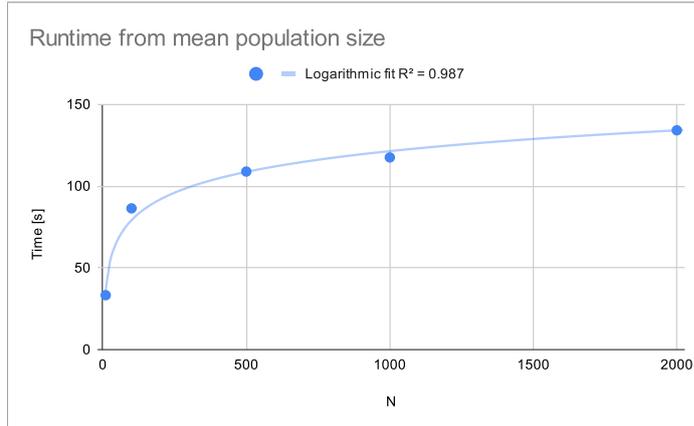
### Effect of sampled fraction

	Output Directory	Seed	Dimensions	Mean population size	Time points	Time step	Generation time	Migration rate	Sampling fraction	Run 1	Run 2	Run 3	Run 4	Run 5	Mean	Standar deviation
	SampledFraction01	1	15x12	100	10	200	5	0.1	0.1	22.264	22.241	22.368	22.352	22.225	<b>22.29</b>	0.0656
	SampledFraction03	1	15x12	100	10	200	5	0.1	0.3	53.872	53.88	53.743	53.947	53.933	<b>53.875</b>	0.0806
	SampledFraction05	1	15x12	100	10	200	5	0.1	0.5	86.26	84.539	84.725	84.597	85.83	<b>85.1902</b>	0.7978
	SampledFraction07	1	15x12	100	10	200	5	0.1	0.7	109.865	107.255	107.284	107.682	107.587	<b>107.9346</b>	1.0950
	SampledFraction1	1	15x12	100	10	200	5	0.1	1	151.201	148.181	148.255	150.387	150.401	<b>149.685</b>	1.3794



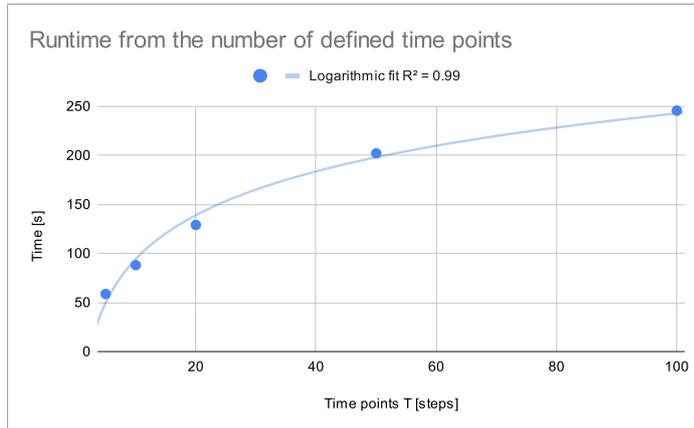
### Effect of mean population size

910	Output Directory	Seed	Dimensions	Mean population size	Time points	Time step	Generation time	Migration rate	Sampling fraction	Run 1	Run 2	Run 3	Run 4	Run 5	Mean	Standard deviation
	MeanPopSize10	1	15x12	10	10	200	5	0.1	0.5	33.411	33.438	32.921	33.389	33.163	<b>33.2644</b>	0.2210
	MeanPopSize100	1	15x12	100	10	200	5	0.1	0.5	86.761	86.665	86.316	85.975	86.143	<b>86.372</b>	0.3355
	MeanPopSize500	1	15x12	500	10	200	5	0.1	0.5	109.53	108.45	108.626	109.001	108.803	<b>108.882</b>	0.4161
	MeanPopSize1000	1	15x12	1000	10	200	5	0.1	0.5	117.689	117.189	117.565	117.577	117.559	<b>117.5158</b>	0.1903
	MeanPopSize2000	1	15x12	2000	10	200	5	0.1	0.5	127.408	129.057	137.914	138.803	137.408	<b>134.118</b>	5.4273



### Effect of the number of defined time points

	Output Directory	Seed	Dimensions	Mean population size	Time points	Time step	Generation time	Migration rate	Sampling fraction	Run 1	Run 2	Run 3	Run 4	Run 5	Mean	Standard deviation
	TimePoints5	1	15x12	100	5	200	5	0.1	0.5	59.327	60.09	57.196	58.492	58.273	<b>58.6756</b>	1.0966
	TimePoints10	1	15x12	100	10	200	5	0.1	0.5	87.903	84.501	91.794	88.03	88.135	<b>88.0726</b>	2.5807
	TimePoints20	1	15x12	100	20	200	5	0.1	0.5	128.064	123.701	130.296	129.128	133.872	<b>129.0122</b>	3.6874
	TimePoints50	1	15x12	100	50	200	5	0.1	0.5	201.55	199.504	199.865	209.857	199.463	<b>202.0478</b>	4.4483
	TimePoints100	1	15x12	100	100	200	5	0.1	0.5	242.78	246.714	246.313	246.027	246.14	<b>245.5948</b>	1.5950



**Effect of the generation time**

911	Output Directory	Seed	Dimensions	Mean population size	Time points	Time step	Generation time	Migration rate	Sampling fraction	Run 1	Run 2	Run 3	Run 4	Run 5	Mean	Standar deviation
	GenTime1	1	15x12	100	10	200	1	0.1	0.5	185.025	178.495	174.344	176.235	176.189	<b>178.0576</b>	4.1635
	GenTime5	1	15x12	100	10	200	5	0.1	0.5	91.379	89.823	88.743	96.7	89.727	<b>91.2744</b>	3.1764
	GenTime10	1	15x12	100	10	200	10	0.1	0.5	55.029	54.663	53.67	54.292	57.213	<b>54.9734</b>	1.3489
	GenTime50	1	15x12	100	10	200	50	0.1	0.5	22.041	23.049	23.138	22.052	21.9	<b>22.436</b>	0.6040
	GenTime100	1	15x12	100	10	200	100	0.1	0.5	16.679	16.876	16.592	16.422	16.675	<b>16.6488</b>	0.1642

## Runtime from generation time

