

## Dungeness crab populations fluctuate and correlate to megalopae abundance

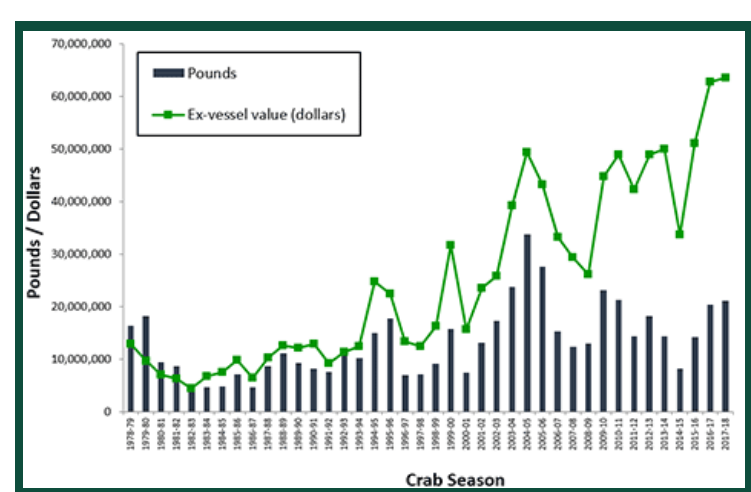


Figure 1. Oregon commercial Dungeness crab catch.<sup>1</sup>

The Dungeness crab fishery is one of the highest value fisheries in the US Pacific Northwest, but catch rates fluctuate interannually<sup>1</sup> (Fig. 1). Variable environmental conditions are hypothesized to be drivers, though precise mechanisms are not well understood. However, the abundance of the last larval stage, the megalopal stage, is correlated to the abundance of fishery catch in Oregon four years later<sup>2</sup> (Fig. 2).

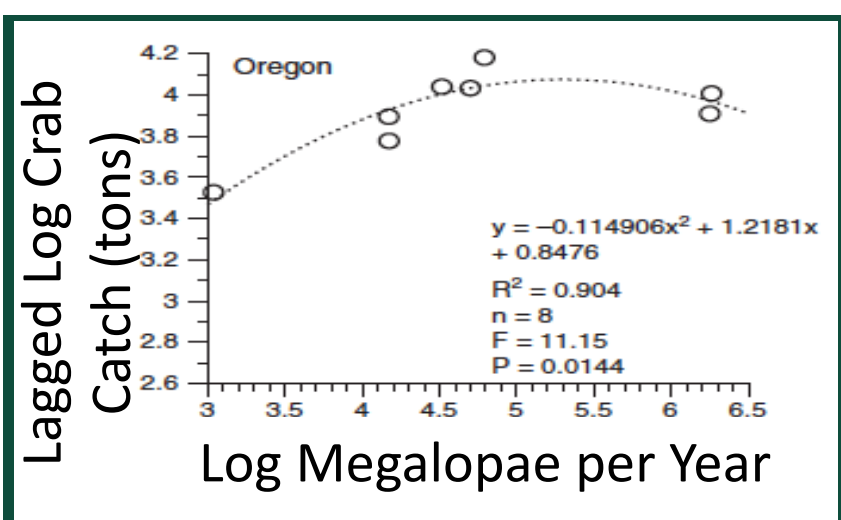


Figure 2. Correlation of megalopae abundance and adult catch.<sup>2</sup>

**Hypothesis:** Environmental exposure history of Dungeness megalopae is important for predicting their distribution. A statistical model for predicting megalopae occurrence that includes exposure history will out-perform a model that uses only ‘*in situ*’ ocean conditions coincident with megalopae sampling.

## Extracting megalopae habitat: ‘*in situ*’ conditions versus simulated exposure history

**J-SCOPE** (JISAO’s Seasonal Coastal Ocean Prediction of the Ecosystem<sup>3,4</sup>) produces historical ocean simulations (‘hindcasts’):

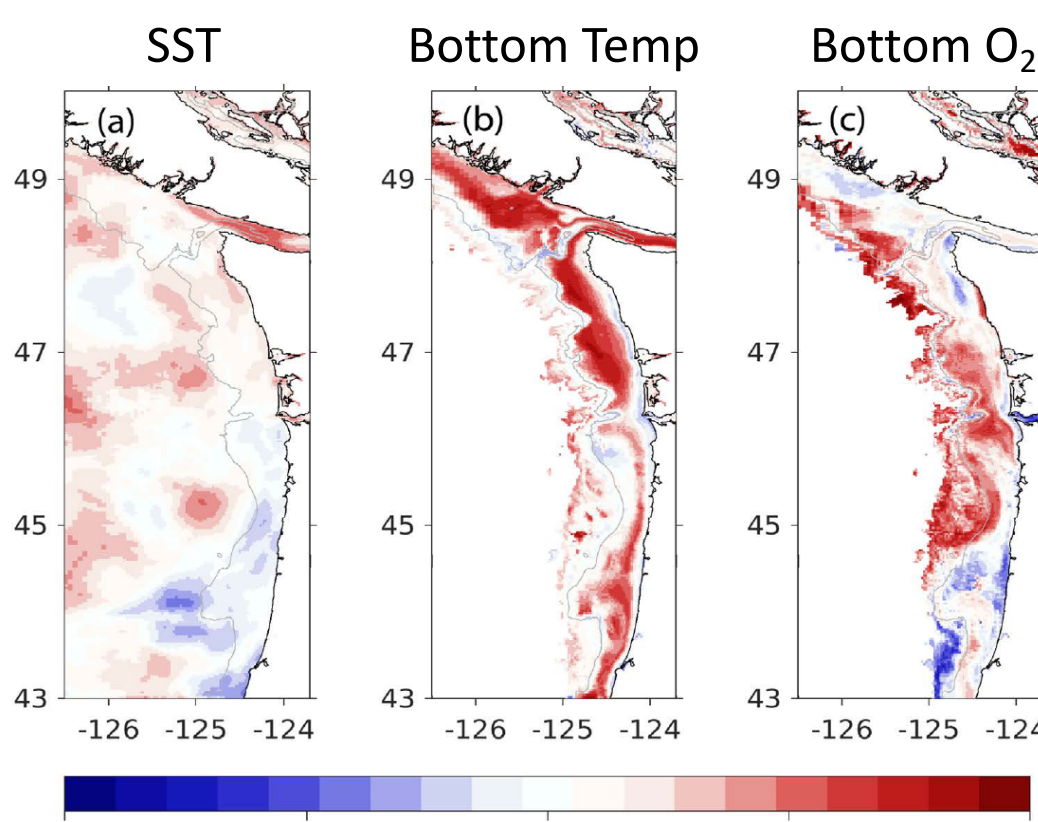
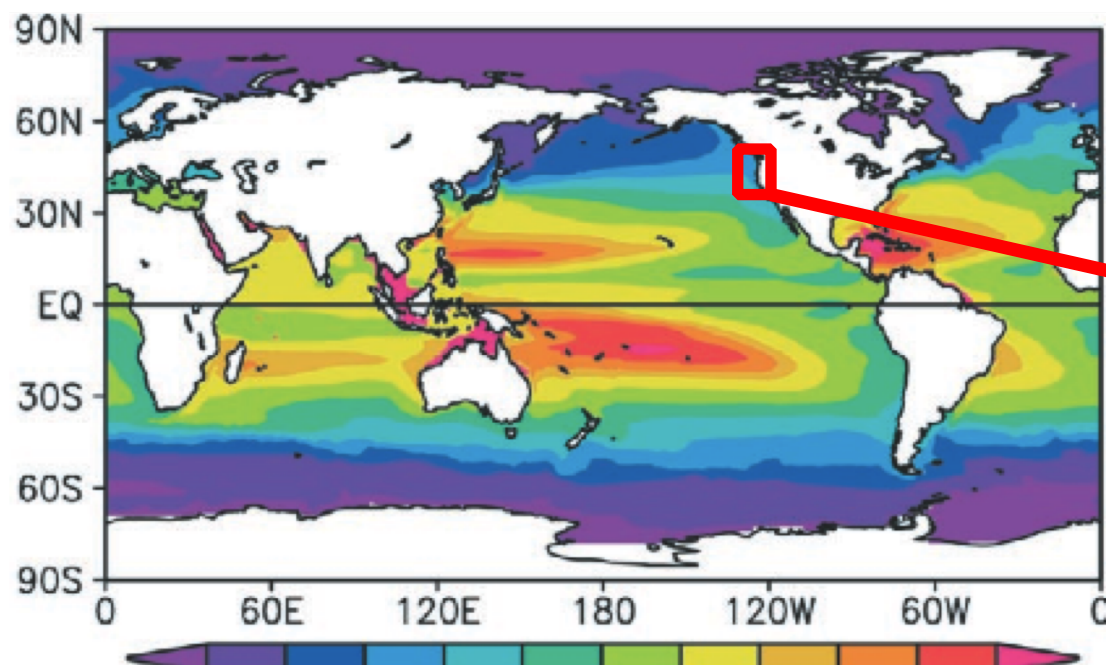


Figure 4. Anomaly Correlation Coefficient for seasonal forecast vs hindcast.<sup>3,4</sup>

- NOAA’s CFS (coupled air/sea/land model; Fig. 3) provides boundary & atm forcing of ROMS-based regional model with biogeochemistry
- Modeled fields: T, S, O, NO<sub>3</sub>, Chl a; derived variables: pH, Ω
- Model skill evaluated<sup>3,4</sup> (Fig. 4)
- Fields applied to habitat modeling: sardine<sup>5</sup>, crab, pteropods<sup>6</sup>, and hake<sup>7</sup>

Figure 3. Climate Forecast System (CFS)



Check out our website: [www.nanoos.org/products/j-scope](http://www.nanoos.org/products/j-scope)

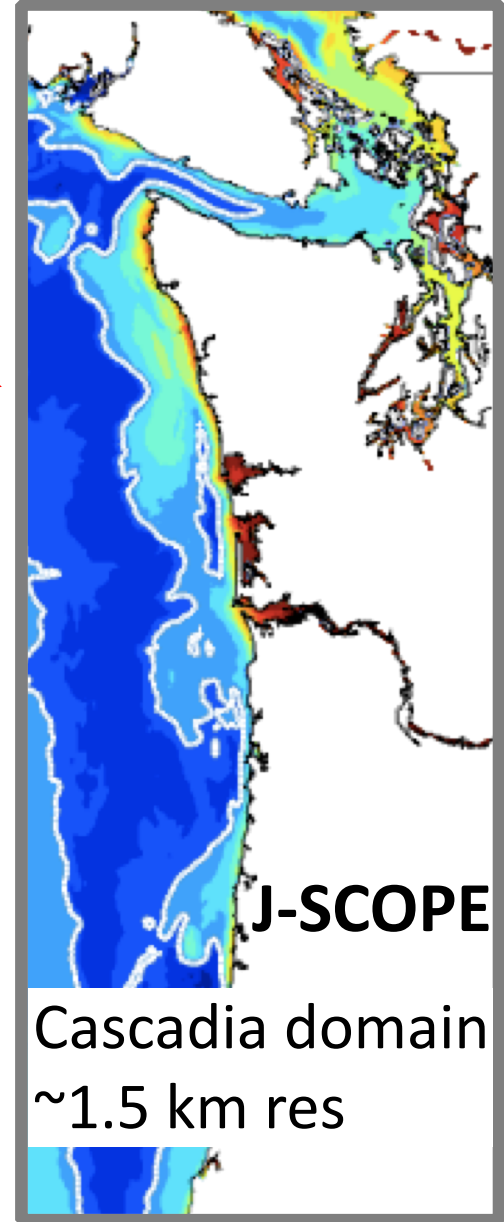
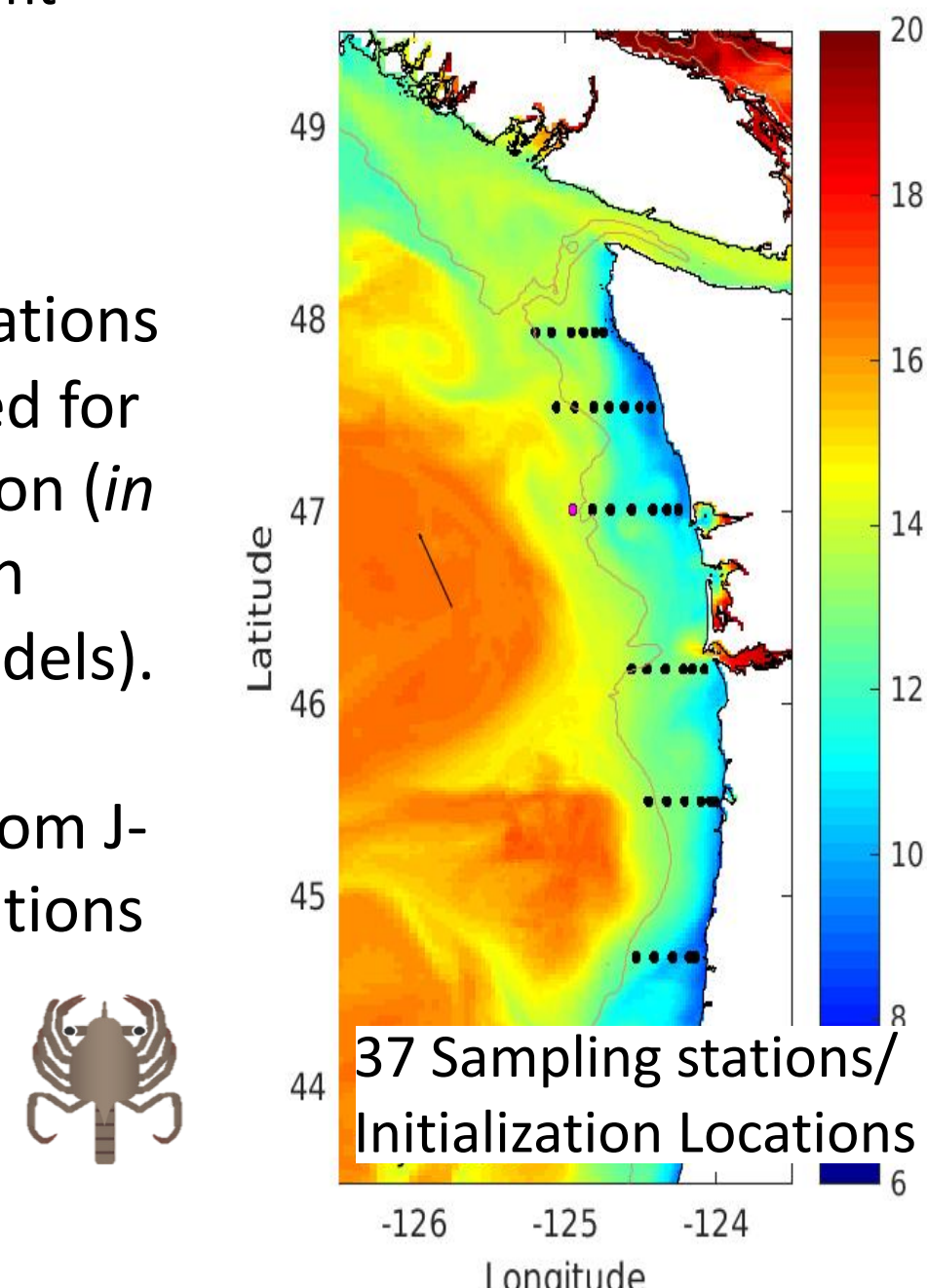


Figure 5. Megalopae sampling locations (37 stations, years 2009-2017) used for environmental conditions extraction (*in situ* model) and particle simulation initialization (exposure history models).

A. ‘*in situ*’ conditions extracted from J-SCOPE hindcasts at times and locations where megalopae were sampled, averaged between 0-30m depth.

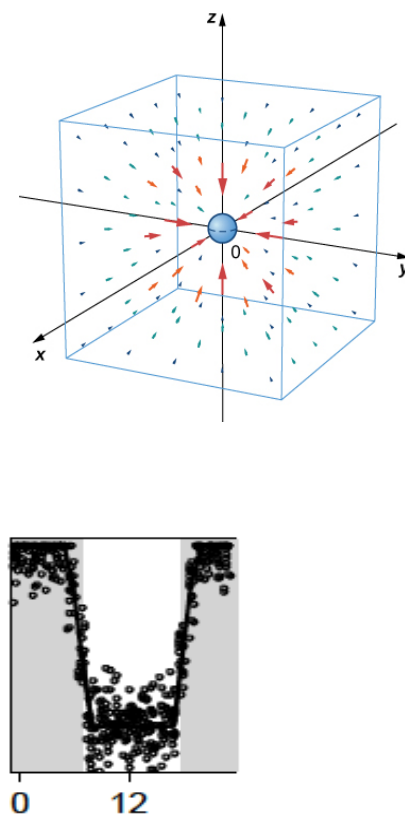


B. **Exposure history simulations:** particles initialized and tracked backward for 30 days (LTRANSv2<sup>8</sup>)

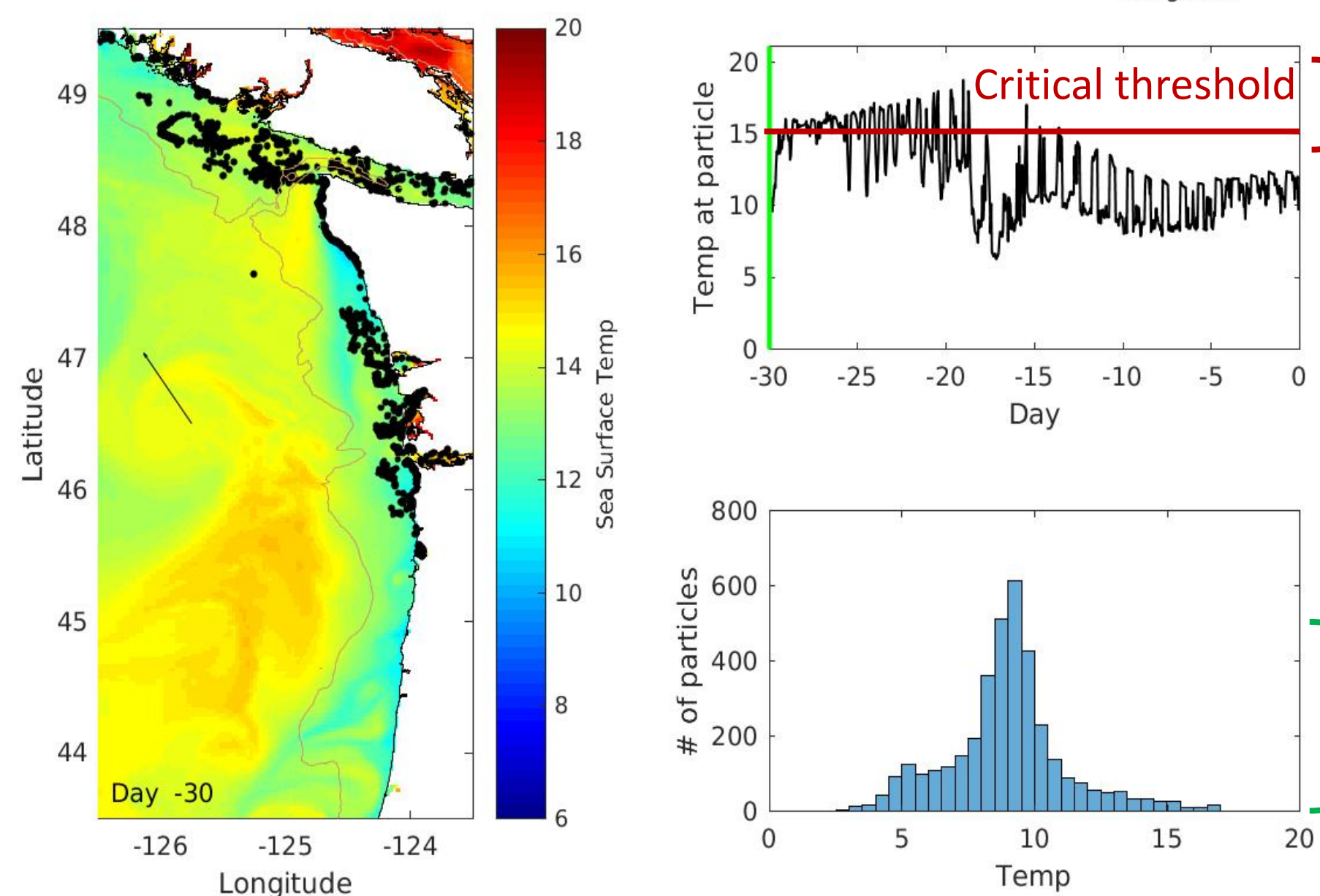
- **Advection, random displacement, and environmental conditions** from JSCOPE hindcasts<sup>3,4</sup>

• **Larval Behavior**

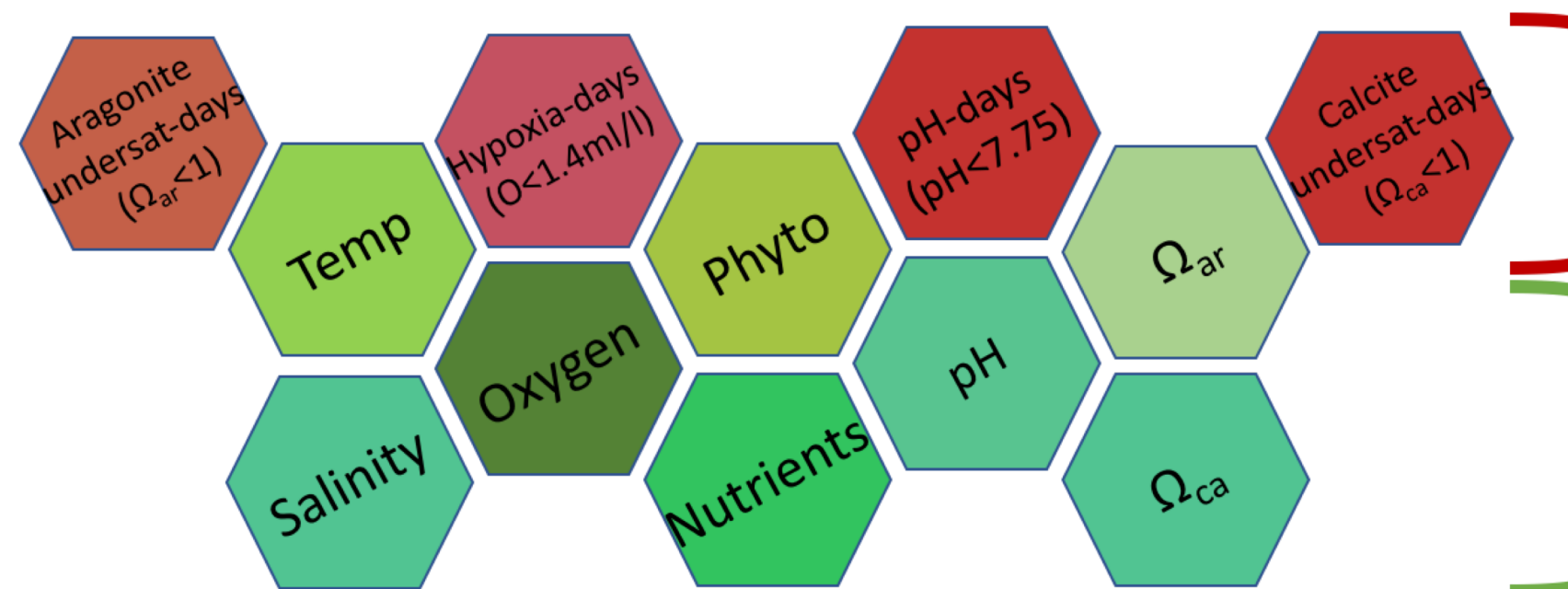
- Diel vertical migration (DVM)<sup>9</sup>
- Surface-following
- Passive
- Combination



More information in Norton et al., 2020, *Front. Mar. Sci.* 7:102.



a) **Severity Index:** Time and degree a threshold is surpassed



b) **Average conditions of exposure**

Figure 6. Two measures of environmental exposure history calculated along particle trajectories.<sup>6,10</sup>

## Behavior affects environmental exposure history

Behavior characterized larval depth habitat (Fig. 5), which determined environmental exposure history (Fig. 6). Particles with similar depth habitats experienced similar environmental conditions (e.g., EH-DVM30/EH-P1/EH-D15P; EH-DVM60/EH-P30), while particles with unique depth habitats experienced unique environmental conditions (e.g., EH-S1).

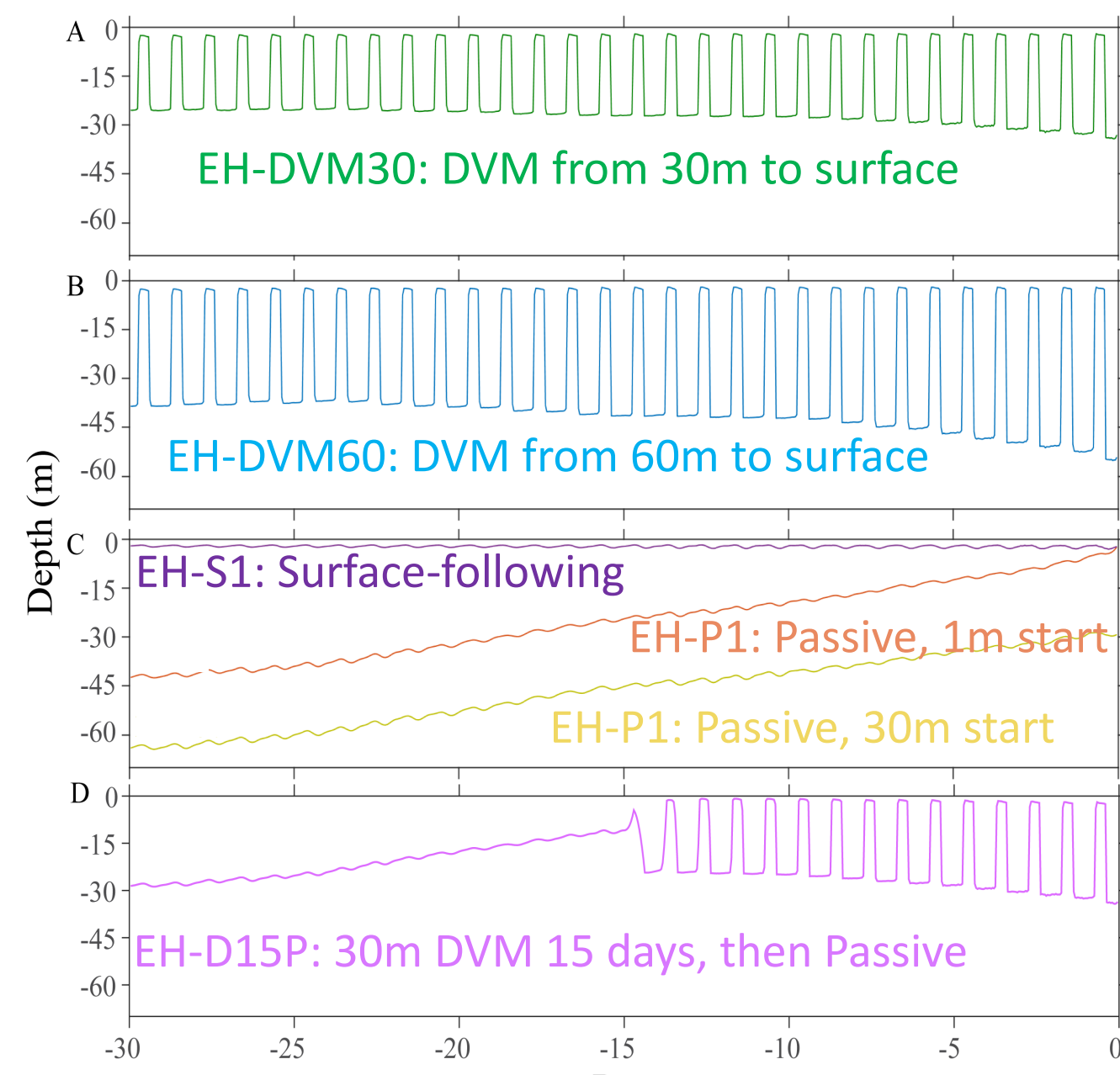
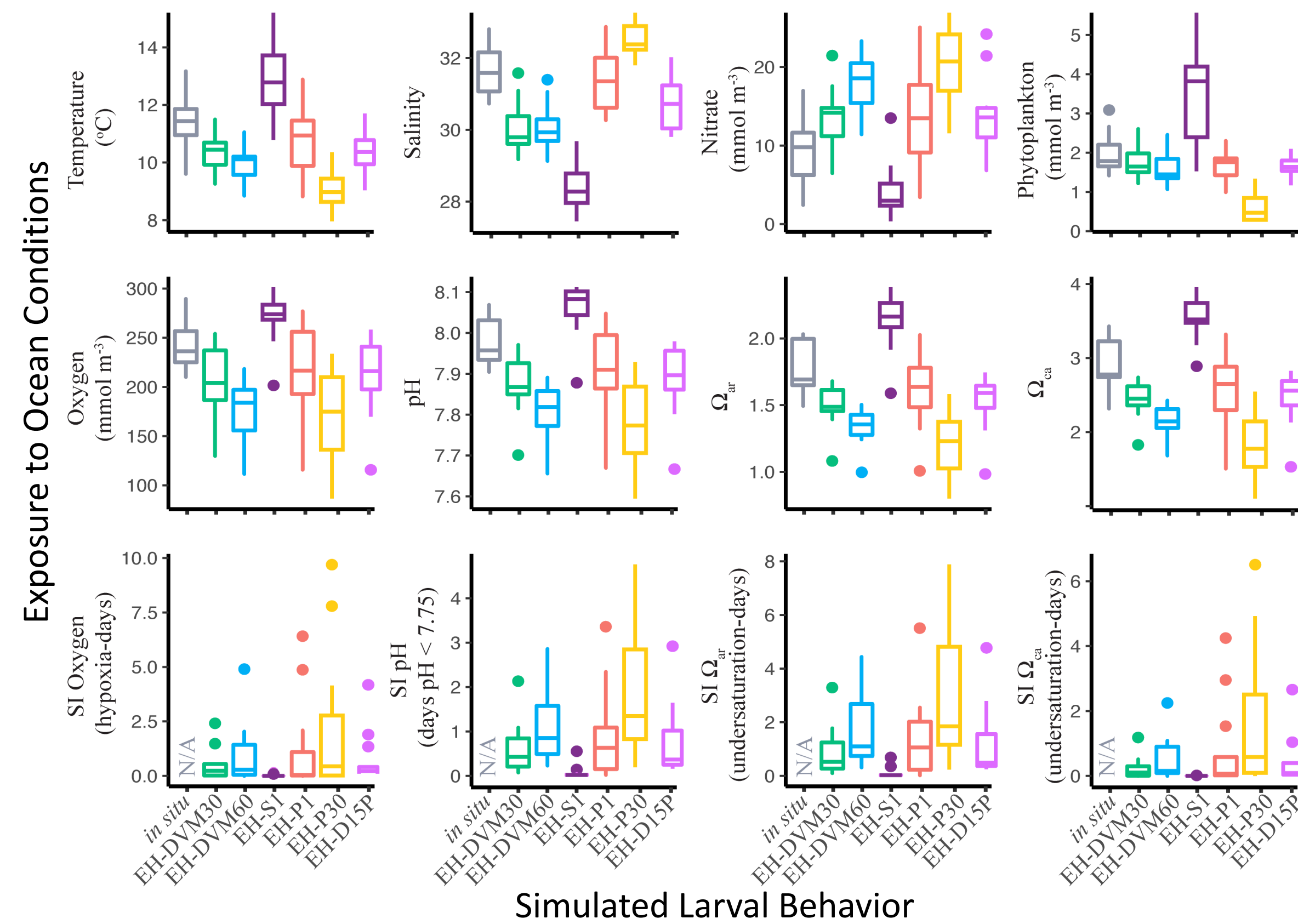


Figure 7. (Above) Simulated larval behaviors for exposure history experiments.

Figure 8. (Right) Exposure history differed among dispersal behaviors.



## Exposure history improves model fit and performance compared to *in situ* model

Table 1. Generalized linear models (GLMs) for *in situ* and exposure history (EH) behaviors. All exposure history GLMs had better fit (i.e., lower AICc) and performance (i.e., higher AUC) than the *in situ* model, regardless of which behavior was simulated. Predictor variables varied among models; significant predictors in **bold**, direction of correlation indicated (i.e., positive (+) or negative (-)).

Experiment	Predictors ( <b>bold p&lt;0.05</b> )	ΔAICc	in-sample AUC	Model fit metric: lower is better	Model performance metric: higher is better
<i>in situ</i>	-N	11.8	0.602		
EH-DVM30	+O	4.7	0.644		
EH-DVM60	+S, +O	5.3	0.650		
EH-S1	-T, -N, -SI Ωca	7.9	0.645		
EH-P1	+S, +O	0.0	0.658		
EH-P30	+P, -SI Ωar	1.9	0.625		
EH-D15P	+pH	1.7	0.657		

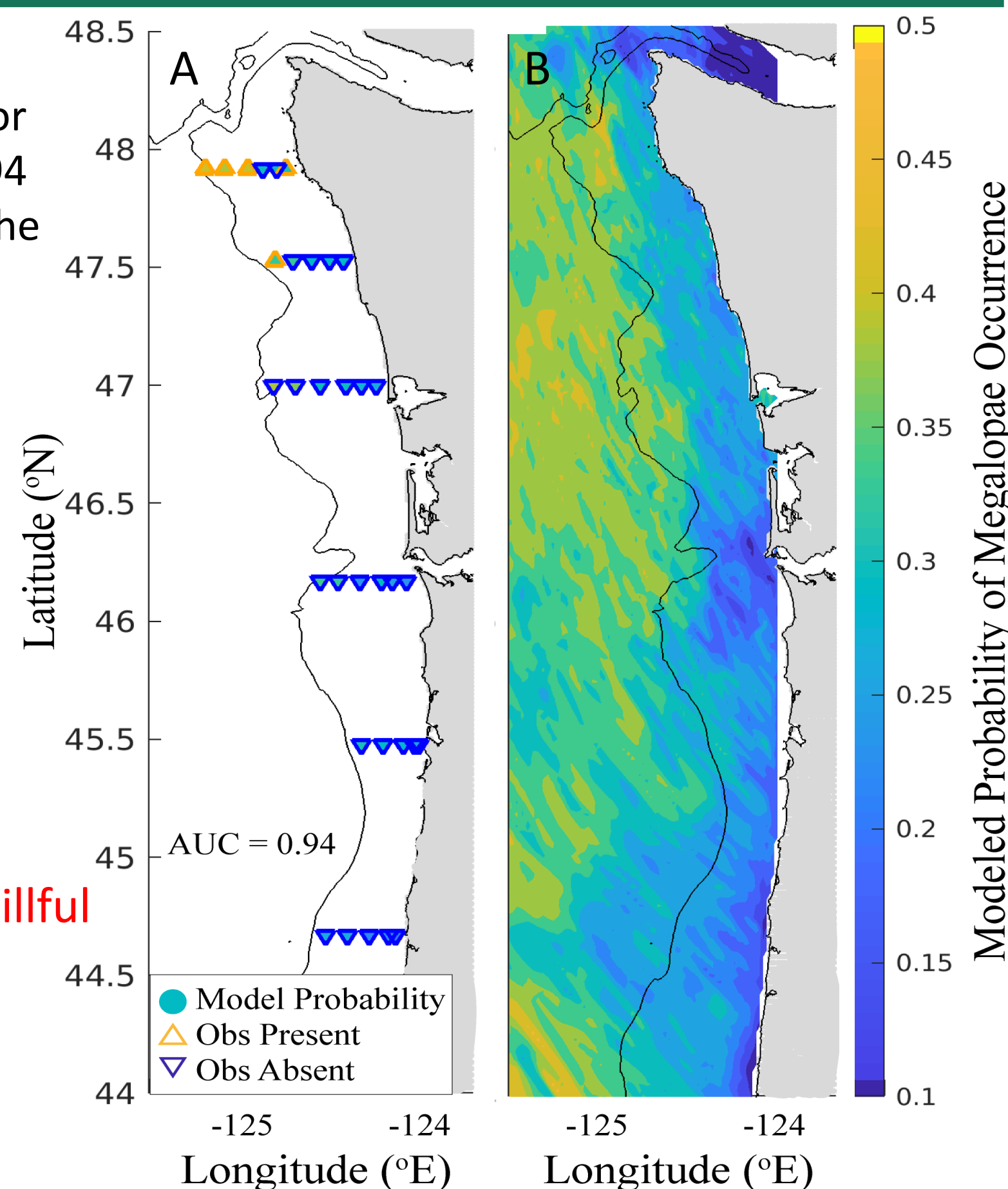
## Biological ensemble skillfully predicts megalopae occurrence

Table 2. The best models (see Table 1) were selected to compose a **biological ensemble**. Model performance (AUC) is shown for an out-of-sample year (2017) for each model individually and for the biological ensemble as a whole. An AUC of 0.94 indicates that probably of megalopae occurrence was correctly ranked at 94% of the stations.

Experiment	Equation ( <b>bold p&lt;0.05</b> )	2017 AUC
EH-DVM30	-3.01 + 0.109* <b>O</b>	0.814
EH-DVM60	-6.42 + 0.132* <b>S</b> + 0.00988* <b>O</b>	0.936
EH-S1	1.77 - 0.157*T - 0.0994*N - 79.5*(SI Ωca)	0.757
EH-P1	-11.0 + 0.248* <b>S</b> + 0.0111* <b>O</b>	0.914
EH-D15P	-34.9 + 4.32* <b>pH</b>	0.779
<b>Biological Ensemble:</b>		<b>0.943</b>

AUC > 0.5 is skillful

Figure 9. (Right) The biological ensemble predicting (A) megalopae occurrence compared to observed occurrence, and (B) habitat probabilities throughout the J-SCOPE domain, both for the out-of-sample test year 2017.



## Developing statistical models of megalopae occurrence

- Generalized linear models developed in Matlab (‘stepwiseglm’) using a logit link:

$$f(\mu) = \log\left(\frac{\mu}{1-\mu}\right) \quad \text{with} \quad \mu = \frac{e^{X_b}}{1 + e^{X_b}} \quad \text{and } X_b \text{ is a linear combination of predictor variables}$$

- Model fit: Akaike Information Criterion corrected for small sample sizes (AICc)
- Model performance: Area Under the (ROC) Curve (AUC)

**References:** <sup>1</sup><https://www.dfw.state.or.us/MRP/shellfish/commercial/crab/landings.asp>. <sup>2</sup>Shanks, A.L., 2013, *Fish. Oceanogr.* 22:263-272. <sup>3</sup>Siedlecki et al., 2016, *Sci. Rep.* 6: 27203. <sup>4</sup><http://www.nanoos.org/products/j-scope/home.php>. <sup>5</sup>Kaplan et al., 2016, *Fish. Oceanogr.* 25:15-27. <sup>6</sup>Bednarsek et al., 2017, *Prog. Oceanogr.* 145:1-24. <sup>7</sup>Malick et al., (in prep). <sup>8</sup>Schlag, Z.R., North, E.W. 2012, LTRANSv2 User Guide, UMCS. <sup>9</sup>Hobbs, R.C., Botsford, L.W. 1992, *Mar. Biol.* 112:417-428. <sup>10</sup>Hauri et al., 2013, *Geophys. Res. Lett.* 40:3424-3428.

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